

Two Arabidopsis Mutants That Overproduce Ethylene Are Affected in the Posttranscriptional Regulation of 1-Aminocyclopropane-1-Carboxylic Acid Synthase¹

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The Arabidopsis mutants *eto1* (ethylene overproducer) and *eto3* produce elevated levels of ethylene as etiolated seedlings. Ethylene production in these seedlings peaks at 60 to 96 h, and then declines back to almost wild-type levels. Ethylene overproduction in *eto1* and *eto3* is limited mainly to etiolated seedlings; light-grown seedlings and various adult tissues produce close to wild-type amounts of ethylene. Several compounds that induce ethylene biosynthesis in wild-type, etiolated seedlings through distinct 1-aminocyclopropane-1-carboxylic acid (ACC) synthase (ACS) isoforms were found to act synergistically with *eto1* and *eto3*, as did the ethylene-insensitive mutation *etr1* (ethylene resistant), which blocks feedback inhibition of biosynthesis. ACS activity, the rate-limiting step of ethylene biosynthesis, was highly elevated in both *eto1* and *eto3* mutant seedlings, even though RNA gel-blot analysis demonstrated that the steady-state level of ACS mRNA was not increased, including that of a novel Arabidopsis ACS gene that was identified. Measurements of the conversion of ACC to ethylene by intact seedlings indicated that the mutations did not affect conjugation of ACC or the activity of ACC oxidase, the final step of ethylene biosynthesis. Taken together, these data suggest that the *eto1* and *eto3* mutations elevate ethylene biosynthesis by affecting the posttranscriptional regulation of ACS.

The gaseous hormone ethylene has been shown to influence numerous plant growth and developmental processes, including germination, root-hair initiation, leaf and flower senescence and abscission, fruit ripening, nodulation, and the response to a wide variety of stresses (Mattoo and Suttle, 1991; Abeles et al., 1992). Much progress has been made in elucidating the mechanisms of ethylene perception and signal transduction (Ecker, 1995; Kieber, 1997a, 1997b), as well as the ethylene-biosynthetic pathway (Kende, 1993). However, to fully understand the mechanism of ethylene action, it is important to delineate how its biosynthesis is regulated. We have chosen 3-d-old etiolated Arabidopsis seedlings as a model system to unravel this circuitry. Here we describe the characterization of two mutants that affect the regulation of ethylene biosynthesis in etiolated seedlings.

Almost all plant tissues have the ability to make ethylene, although in most cases the amount made is very low.

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Ethylene production increases dramatically during a number of developmental events such as germination, leaf and flower senescence and abscission, and fruit ripening (Yang and Hoffman, 1984; Mattoo and Suttle, 1991; Abeles et al., 1992). A diverse group of factors modify the level of ethylene biosynthesis, and a major effect of most of these is to increase the steady-state level of ACS mRNA (see Olson et al., 1991, 1995; Rottmann et al., 1991; Botella et al., 1993, 1995), although there is accumulating evidence to suggest that this enzyme is also posttranslationally regulated (Nakajima et al., 1990; Spanu et al., 1994; Oetiker et al., 1997; Vogel et al., 1998b).

The ethylene-biosynthetic pathway (for review, see Yang and Hoffman, 1984; Kende, 1993) starts with the conversion of Met to AdoMet by the enzyme Met adenosyltransferase. ACS, which converts AdoMet to ACC (Adams and Yang, 1979), is the first committed and generally rate-limiting step in ethylene biosynthesis. ACS is encoded by a small gene family comprising at least three to six members in the species that have been closely examined. Distinct subsets of ACS genes are expressed in response to various developmental, environmental, and hormonal factors.

In Arabidopsis six ACS genes have been identified (ACS1–ACS6), two of which are nonfunctional (Liang et al., 1992, 1995; Van der Straeten et al., 1992; Vahala et al., 1998). Inhibition of protein synthesis by cycloheximide treatment induces expression of the functional genes, suggesting that they are under negative control (Liang et al., 1992). Wounding, auxin, LiCl, and anaerobiosis differentially induce these genes (Liang et al., 1992, 1996; Van der Straeten et al., 1992). ACS2 expression is higher in young, developing leaves and flowers compared with more mature tissues from these organs, and its expression is also correlated with the initial stages of lateral root formation (Rodrigues-Pousada et al., 1993). ACS5 is the major isoform involved in the production of ethylene in response to low doses of cytokinin in etiolated seedlings, and this regulation is primarily via a posttranscriptional mechanism (Vogel et al., 1998b). ACS4 transcription is induced by auxin, and several auxin-responsive elements have been identified upstream of the ACS4 coding region (Abel et al., 1995). The steady-state level of ACS6 transcript is increased by treatment with ozone (Vahala et al., 1998). The ACS3 gene is most likely a

Abbreviations: ACS, ACC synthase; AdoMet, S-adenosyl-Met.

pseudogene and *ACS1* encodes a nonfunctional ACS (Li-ang et al., 1995).

The final step of ethylene biosynthesis, the conversion of ACC to ethylene, is catalyzed by the enzyme ACC oxidase, which can also play an important role in regulating ethylene biosynthesis, especially under conditions of high ethylene production (Nadeau et al., 1993; Kim and Yang, 1994; Tang et al., 1994; Barry et al., 1996; Lasserre, 1996; Mekhedov and Kende, 1996). ACC oxidase, like ACS, appears to be encoded by a gene family whose members are differentially regulated in a number of plants. There are several ACO genes in *Arabidopsis* (Newman et al., 1994), at least one of which is induced by ethylene itself (Gomez-Lim et al., 1993).

ACC can be conjugated to an inactive form, malonyl-ACC, by the enzyme ACC malonyltransferase (Amrhein et al., 1981; Hoffman et al., 1982; Kionka and Amrhein, 1984). A second ACC conjugate, 1-(γ -L-glutamylamino) cyclopropane-1-carboxylic acid, has also been identified (Martin et al., 1995), although recent evidence suggests that this is a much less abundant conjugate (Peiser and Yang, 1998). There is some evidence that the level of ACC conjugation is regulated, which may contribute to control of ethylene production (Jiao et al., 1986).

Treatment of etiolated seedlings with ethylene results in a morphology known as the triple response. In *Arabidopsis* the triple response consists of shortening and radial swelling of the hypocotyl, inhibition of root elongation, and exaggeration of the curvature of the apical hook (see Fig. 1). This response has been used to identify mutants disrupted in ethylene perception and signaling (Bleecker et al., 1988; Guzman and Ecker, 1990; Kieber et al., 1993), as well as mutants affected in the regulation of ethylene biosynthesis. Mutants in the latter class fall into two categories: (a) those that fail to induce ethylene in response to a particular inducer (cytokinin-insensitive mutants, *Cin*; Vogel et al., 1998a, 1998b); and (b) those that overproduce ethylene (ethylene overproducer, *Eto*; Guzman and Ecker, 1990; Kieber et al., 1993). Three *Eto* loci have been identified: *eto1* is inherited as a recessive mutation, and *eto2* and *eto3* are dominant. *eto2* was recently found to be the result of a disruption of the carboxy-terminal 11 amino acids of ACS5 (Vogel et al., 1998b). Here we describe the physiological characterization of the *eto1* and *eto3* mutants. This analysis suggests that these mutations are affected in the posttranscriptional regulation of ACS.

MATERIALS AND METHODS

Plant Lines and Growth Conditions

The Columbia ecotype of *Arabidopsis* was used in this study. Seeds were surface-sterilized as described previously (Vogel et al., 1998b), resuspended in a suitable volume of top agar (0.8% low-melt agarose), and spread onto Murashige and Skoog agar (Murashige and Skoog salts [GIBCO-BRL], 2% Suc, and 0.8% agar, pH 5.7). Seeds were cold treated for 4 d (4°C), exposed to light for 2 h, and then moved to a dark incubator at 23°C. The time at which the vials were moved to 23°C was designated as time 0. Adult

plants were grown in potting soil (Metro Mix 250, Grace-Sierra, Boca Raton, FL) under continuous illumination at 23°C. The *eto1-1* and *eto3* mutants and the *eto1-1/etr1-3* double mutant were identified previously (Guzman and Ecker, 1990; Kieber et al., 1993; Roman et al., 1995). The *eto1-3* allele was isolated from an x-ray-mutagenized population (ecotype Columbia) (Kieber et al., 1993). This allele was used for all of the experiments, with the exception of the *eto1/etr1* double-mutant analysis, which used the *eto1-1* allele. The *eto1-3/ein2* double mutant was obtained by crossing an *eto1-3* homozygote to a *ein2-1* homozygote. The F₁ population was allowed to self, and seedlings that were phenotypically Eto⁻ were then selected in the F₂ line. Tall progeny from these individuals were then selected and self-set seed was collected. A line that retested as being ethylene insensitive and that overproduced ethylene was identified and used in further experiments.

Ethylene Measurement

Hormones and CuSO₄ were added to about 20 2-d-old etiolated seedlings that were grown in 22-mL GC vials containing 3 mL of Murashige and Skoog agar by pipetting 200 μ L of solution on top of the seedlings. An equal volume of water plus solvent was added to the control treatments. These vials were then flushed with hydrocarbon-free air and capped, and the accumulated ethylene measured 24 to 48 h later, as described previously (Vogel et al., 1998b). Ethylene production was normalized to the number of seedlings in each vial and the time between capping and sampling. All observations are from at least three replicates, and each experiment was repeated at least once with comparable results. To measure ethylene from adult plants, tissues were detached, weighed, and then placed in 22-mL vials containing 3 mL of Murashige and Skoog agar. The vials were flushed with hydrocarbon-free air, sealed, and incubated in the light for the indicated times. The amount of ethylene produced by light-grown seedlings was determined by putting capped vials in a lighted growth chamber for 72 h.

ACS Assays

ACS was assayed from 3-d-old etiolated seedlings as described previously (Peck and Kende, 1995), with some modifications. Sterilized seeds were plated on filter paper on Murashige and Skoog agar (10,000 seeds per 150-mm plate), cold incubated (4°C) for 4 d, and then moved to a dark chamber for 3 d at 23°C. Ten grams of tissue was added to 15 mL of buffer A (250 mM phosphate buffer, pH 8.0, 10 μ M pyridoxal phosphate, 1 mM EDTA, 2 mM PMSF, and 5 mM DTT) and the sample was homogenized on ice for 4 min (maximum speed with a PowerGen 700 homogenizer [Fisher Scientific]). The sample was centrifuged at 15,000g for 15 min and the supernatant was respun at 15,000g for 15 min. One milliliter of the supernatant was placed into a 22-mL GC tube and 100 μ L of 5 mM AdoMet was added. This was incubated for 1 h at 22°C. The ACC formed was converted to ethylene by addition of 100 μ L of 20 mM HgCl₂, followed by 100 μ L of a 1:1 mixture of

saturated NaOH:bleach (Lizada and Yang, 1979). The tubes were capped immediately after addition of the NaOH/bleach and incubated on ice for 10 min. Five milliliters of headspace was removed with a syringe and injected into a new vial, and the ethylene was measured as described previously (Vogel et al., 1998b). All reactions were done in triplicate and compared with controls, to which AdoMet was not added. Protein concentration was determined using the Bradford assay as described by the manufacturer (Bio-Rad).

RNA-Blot Analysis

Total RNA was prepared as described previously (Ausubel et al., 1994) and poly(A⁺) RNA was isolated using Oligotex-dT resin, as described by the manufacturer (Qiagen, Chatsworth, CA). Five micrograms of poly(A⁺) mRNA was separated on an agarose gel, blotted to a nylon membrane, and hybridized to radiolabeled probes as described previously (Ausubel et al., 1994). Fragments corresponding to each ACS gene and β -tubulin were obtained by amplifying each from Arabidopsis genomic DNA using PCR with oligonucleotide primers specific for each gene, or in the case of ACS6, the insert from the expressed sequence tag clone FAI88 (see below) was used. The signals were quantified with a phosphor imager and normalized to the level of the β -tubulin control. This analysis was repeated once with comparable results.

Isolation of ACS7

We searched the Arabidopsis expressed sequence tag database (Newman et al., 1994) for sequences similar to those of ACS1 to ACS6. Three clones (FAI88, 288D2T7, and 240L12T7), corresponding to an identical gene, were identified that were similar to the previously identified Arabidopsis ACS genes, but encoded a novel ACS isoform. Sequence analysis revealed that these ACS7 cDNA clones were missing the 5' portion of the coding region (see "Results").

RESULTS

Ethylene Biosynthesis in the *eto1* and *eto3* Mutants Is Developmentally Regulated

The *eto1* and *eto3* mutants display a constitutive triple-response phenotype as etiolated seedlings (Fig. 1) caused by an overproduction of ethylene. We analyzed ethylene biosynthesis from wild-type and Eto mutant etiolated seedlings at various intervals during the first several days of growth (Fig. 2). There was almost no detectable ethylene produced by wild-type seedlings during the early stages of germination (<24 h), followed by a low, stable level during the next 6 d. Ethylene biosynthesis in *eto1-3* seedlings rose quickly between 48 and 60 h, and remained at a steady, elevated level for the next 60 h, after which it gradually declined to close to wild-type levels. *eto3* mutant seedlings displayed a similar pattern, although their peak production was about 2.5-fold higher and the rate of decline of pro-

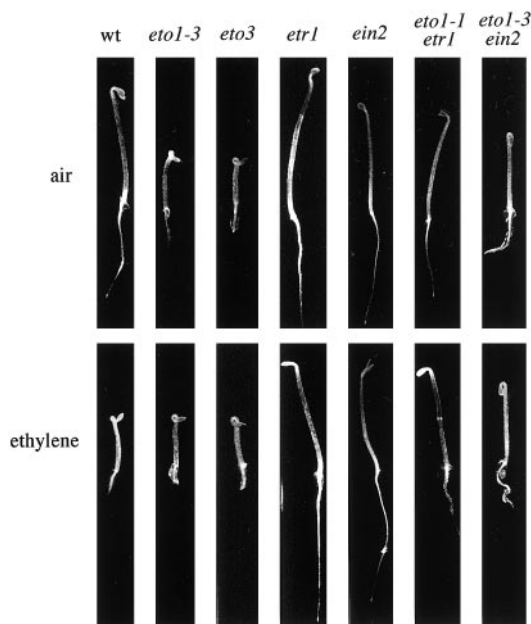


Figure 1. Phenotypes of 3-d-old etiolated wild-type (wt), single-mutant, and double-mutant Arabidopsis seedlings (as indicated) grown in air (top panels) or 10 $\mu\text{L L}^{-1}$ ethylene (bottom panels). Representative seedlings were picked and photographed.

duction was increased compared with that observed in *eto1-3* seedlings. By d 9, both mutants produced close to wild-type levels of ethylene.

To determine if the decline in ethylene biosynthesis was caused by feedback regulation, we examined the time course of ethylene biosynthesis in the double-mutant seedlings of *eto1-3* and the mutation *ein2* ethylene insensitive (Guzman and Ecker, 1990). The initial rate of increase in ethylene biosynthesis was the same for both *eto1-3* and *eto1-3/ein2* seedlings (Fig. 2B). However, the double mutant continued to increase its rate of ethylene biosynthesis beyond 60 h, the point at which the rate in *eto1-3* seedlings peaked. The peak of biosynthesis in the double mutants was close to what one would predict from an additive interaction, suggesting that these mutations act independently to regulate ethylene biosynthesis. The rate of ethylene biosynthesis in double-mutant seedlings also returned to close to that observed in wild-type seedlings, although the rate of decline was somewhat slower than in the *eto1-3* single mutant. This suggests that the decline in biosynthesis is not the result of negative-feedback regulation from the elevated ethylene levels but, rather, may reflect a developmental change in the regulation of ethylene biosynthesis (although feedback regulation may alter the rate at which this occurs).

We examined ethylene production from various adult tissues to determine if any were affected by the Eto mutations. Both *eto1-3* and *eto3* affected ethylene biosynthesis almost exclusively in etiolated seedlings: light-grown seedlings (not shown), adult leaves, flowers, and siliques from *eto1-3* and *eto3* mutants produced close to wild-type levels of ethylene (Fig. 3). Thus, these mutants are either specific

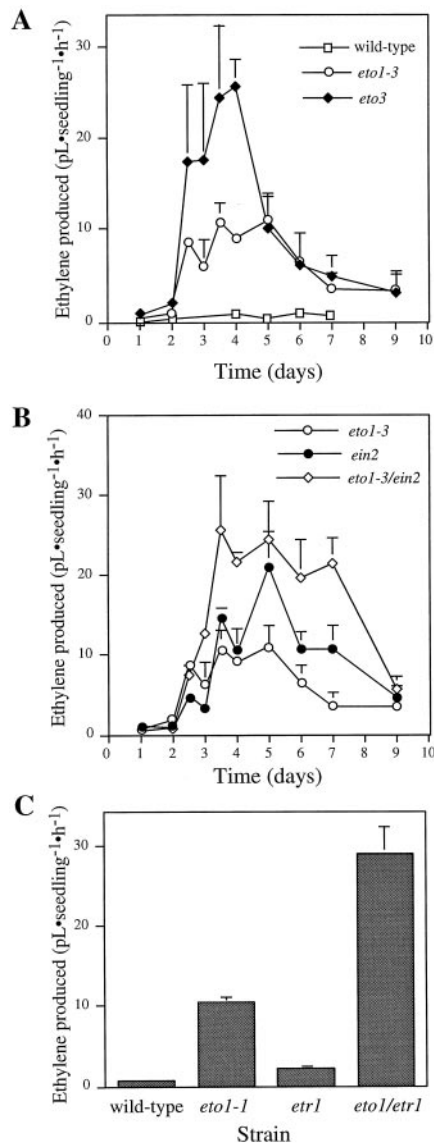


Figure 2. Ethylene biosynthesis by wild-type, single-mutant, and double-mutant etiolated *Arabidopsis* seedlings. Seedlings were grown for various times on Murashige and Skoog agar in GC vials and the accumulated ethylene was measured. A, Time course of ethylene production by etiolated wild-type, *eto1-3*, and *eto3* mutant seedlings. B, Time course of ethylene production by etiolated *eto1-3*, *ein2*, and *eto1-3/ein2* double-mutant seedlings. The data points in A and B represent the ethylene accumulated during the time intervals 0 to 24, 24 to 48, 48 to 60, 60 to 72, 72 to 84, 84 to 96, 120 to 144, 144 to 168, and 168 to 216 h. The second time point in each interval was plotted as the value for the x axis. C, Ethylene accumulated by etiolated wild-type *eto1-1*, *etr1*, and *eto1-1/etr1* double-mutant seedlings from 72 to 96 h. All values are means (\pm SD) of three replicates.

for etiolated seedlings or are involved in regulating ethylene biosynthesis only in the dark.

Interaction of the Eto Mutants and Various Inducers of Ethylene Biosynthesis

Ethylene biosynthesis in etiolated *Arabidopsis* seedlings is strongly induced by a number of plant hormones, as well

as by the cupric ion. In some cases the target of these factors has been demonstrated to be distinct ACS genes (Liang et al., 1992; Abel et al., 1995; Vogel et al., 1998b). We examined the interaction of these factors and the *eto1-3* and *eto3* mutations to begin to address how these signaling pathways are related. The concentration of each inducer was chosen as the concentration that gave the peak of induction in wild-type etiolated seedlings (Woeste et al., 1999).

Both *eto1-3* and *eto3* displayed a synergistic interaction with auxin, cytokinin, cupric ion, and 24-epibrassinolide (Fig. 4). The level of ethylene produced was generally close to multiplicative. For example, the *eto1-3* mutation elevated ethylene biosynthesis approximately 15-fold above wild-type levels, 5 μ M BA increased ethylene approximately 4-fold in etiolated, wild-type seedlings, and BA-treated *eto1-3* seedlings made close to 60-fold more ethylene than untreated, wild-type seedlings. In general, the synergism of these inducers with the *eto3* mutation was slightly less than that observed with *eto1-3*. This synergism indicates that the *eto1-3* and *eto3* mutations may act interdependently with these factors in regulating ethylene biosynthesis. Interestingly, exogenous ABA strikingly dampened the ethylene overproduction observed in the Eto mutants, although it did not appear to affect ethylene production in wild-type seedlings.

Interaction with Mutants Affected in Ethylene Biosynthesis

Ethylene-insensitive mutations produce elevated levels of ethylene, most likely because they block the feedback inhibition of ethylene biosynthesis in vegetative *Arabidopsis* tissue (Guzman and Ecker, 1990). To determine the effect of ethylene feedback on the Eto mutants, we constructed double mutants of *eto1* and the ethylene-insensitive mutants *ein2* and *etr1-3* (ethylene resistant) (Fig. 1). The average length of hypocotyls from the double mutants was not significantly different from that of each ethylene-insensitive parent (not shown), which indicates that the *etr1-3* and *ein2* mutations are epistatic to *eto1*. This

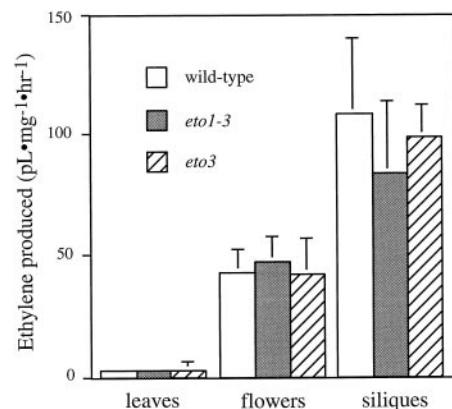


Figure 3. Ethylene produced by adult tissues of wild-type, *eto1-3*, and *eto3* mutant plants growing in soil at 23°C under continuous illumination. Ethylene measurements were as described in "Materials and Methods." Values are means (\pm SD) of three replicates.

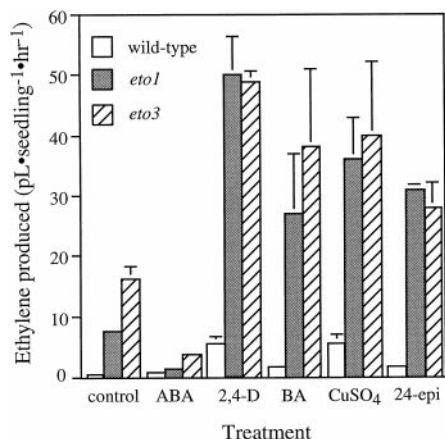


Figure 4. Ethylene produced by wild-type and mutant etiolated *Arabidopsis* seedlings in response to treatment with ABA (75 μM), 2,4-D (160 μM), BA (5 μM), CuSO_4 (20 mM), and 1 μM 24-epibrassinolide (24-epi). For all treatments except BA, seedlings were grown in GC vials, and 200 μL of the solutions of the indicated concentrations was added at 48 h. The vials were flushed with hydrocarbon-free air, sealed, and returned to the dark at 23°C. The ethylene that accumulated during the next 24 h was measured. For BA treatment, the seedlings were germinated on Murashige and Skoog medium supplemented with 5 μM BA and the ethylene production during the first 72 h of germination was measured. Values are means ($\pm\text{SD}$) of three replicates.

confirms the expectation that mutations defective in the perception of ethylene act downstream of those affecting ethylene biosynthesis (Roman et al., 1995). However, under the conditions that we used, the hypocotyl lengths of *ein2* etiolated seedlings were much more variable than those of either the wild-type or *eto1-3/ein2* etiolated seedlings (not shown). It is interesting that the apical hooks of the *eto1-3/ein2* double-mutant etiolated seedlings generally appeared to be more closed than those of the *ein2* single parents (Fig. 1), which have a hook that is less angled than that of wild-type etiolated seedlings.

When combined with the *eto1-1* mutation, the *etr1-3* mutation displayed a synergistic interaction in terms of the amount of ethylene produced (Fig. 2C). However, the amount of ethylene produced by the *eto1-3/ein2* double mutant appeared to be close to additive relative to the parental seedlings (Fig. 2B). This difference may reflect a branch in the ethylene-response pathway or perhaps subtle differences between the *eto1-1* and *eto1-3* alleles.

eto1 and *eto3* Etiolated Seedlings Have Elevated Levels of ACS Activity

One likely target of these ethylene-overproducing mutants is ACS, the rate-limiting step of ethylene biosynthesis. Previous work demonstrated that ACS is the target of various inducers of ethylene biosynthesis in etiolated *Arabidopsis* seedlings by both transcriptional and posttranscriptional mechanisms (Liang et al., 1992, 1996; Van der Straeten et al., 1992; Abel et al., 1995; Vogel et al., 1998b). We assayed the level of ACS in crude extracts from wild-type and *eto1-3* and *eto3* etiolated seedlings (Fig. 5). Both

mutants showed high elevated levels of ACS activity compared with wild-type etiolated seedlings, which had barely detectable levels of ACS activity. This indicates that increases in ACS activity may be responsible for the elevated ethylene biosynthesis observed in the mutant seedlings.

The formation of ACC is generally the rate-limiting step of ethylene biosynthesis, but regulation may also be mediated by changes in ACC oxidase levels or by changes in the amount of ACC that is conjugated. To address this issue, we examined the ability of intact wild-type and mutant seedlings to convert exogenous ACC to ethylene, which should reflect the level of ACC oxidase activity minus any ACC that becomes conjugated (Fig. 6). Both *eto1-3* and *eto3* mutants were indistinguishable from wild-type seedlings in their conversion of ACC to ethylene over a wide range of exogenous ACC concentrations. Consistent with this, *eto1-3* and *eto3* seedlings contained wild-type or slightly elevated levels of ACC *N*-malonyltransferase and γ -glutamyltranspeptidase activity (not shown). These data suggest that these mutants elevate ethylene biosynthesis primarily by increasing ACS activity.

eto1 and *eto3* Etiolated Seedlings Have Wild-Type Levels of ACS mRNA

Elevation of ethylene biosynthesis has often been correlated with increases in the steady-state levels of ACS mRNAs, although in a few cases posttranscriptional control has also been demonstrated. To determine if elevated ACS mRNA contributes to the increased ethylene biosynthesis observed in *eto1* and *eto3* etiolated seedlings, we analyzed ACS mRNA levels by northern blotting (Fig. 7). We analyzed the expression of ACS2, ACS4, ACS5, and ACS6, the three previously identified, active ACS genes in *Arabidopsis*. In addition, we analyzed a novel ACS gene

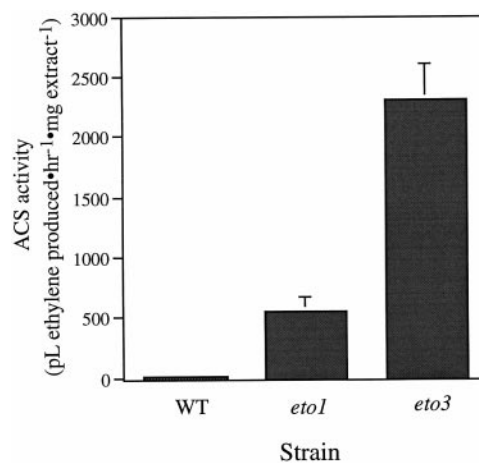


Figure 5. ACS activity in crude extracts from 3-d-old, etiolated wild-type (WT), *eto1*, and *eto3* mutant seedlings. The enzyme was assayed by incubating crude extracts in buffer with or without AdoMet, and then measuring the amount of ACC formed by converting it to ethylene (see "Materials and Methods"). The activity was calculated by subtracting the amount of ethylene produced in the absence of added AdoMet and then normalizing to the protein concentration of each sample. Values are means ($\pm\text{SD}$) of three replicates.

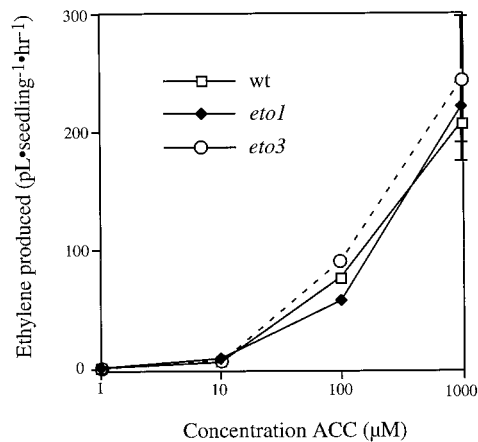


Figure 6. Ethylene produced by etiolated wild-type (wt), *eto1*, and *eto3* Arabidopsis seedlings in response to varying concentrations of supplemented ACC. Seedlings were grown on 3 mL of Murashige and Skoog agar in GC vials. ACC of the indicated concentrations, in a total volume of 200 μ L, was added to the seedlings 48 h after moving to 23°C. The vials were then flushed with hydrocarbon-free air and sealed, and the ethylene produced during the next 24 h was measured. Values are means (\pm SD) of three replicates.

that we found by searching the Arabidopsis expressed sequence tag database (Newman et al., 1994), which we have named ACS7 (Fig. 8).

The ACS7 cDNA clones all appear to lack the 5' end of the gene, because none was as large as the size predicted from RNA gel-blot analysis (approximately 1.6 kb) and the open reading frame continued to the 5' end of the longest sequence. However, the 3' end of the gene is intact and an in-frame stop codon is present, as is a short poly(A⁺) tail. A comparison of the predicted amino acid sequence of ACS7, derived from the sequence of a non-full-length cDNA expressed sequence tag clone, with the other Arabidopsis ACS genes reveals that this clone lacks the variable carboxy-terminal extension present in other ACS proteins (Fig. 8). This carboxy-terminal domain has been demonstrated in at least two cases to negatively regulate the function of the ACS protein (Li and Mattoo, 1994; Vogel et al., 1998).

Overall, the level of expression of all five ACS genes was very low, in most cases just above the level of detection using 10 μ g of poly(A⁺) mRNA. The steady-state level of mRNA for ACS5 in *eto1-3* and *eto3* etiolated seedlings was close to the level observed in wild-type seedlings. This was also confirmed by quantitative reverse-transcriptase PCR analysis (not shown). ACS2 and ACS4 steady-state levels were actually lower in the *Eto* mutants, perhaps reflecting negative feedback from the elevated ethylene levels. There was no detectable ACS6 or ACS7 expression in either wild-type or mutant etiolated seedlings, although treatment with the protein-synthesis inhibitor cycloheximide resulted in high levels of ACS7 expression (not shown), as is the case with the other Arabidopsis ACS genes (Liang et al., 1992). These results indicate that the increased ACS activity is not caused by elevated gene expression, but is likely attributable to a posttranscriptional mechanism.

DISCUSSION

The *eto1* and *eto3* mutants were isolated as seedlings that displayed a constitutive triple-response phenotype caused by an elevation of ethylene biosynthesis. We demonstrate that this ethylene overproduction is brought about by increased ACS activity, and that this is most likely the result of posttranscriptional regulation. This conclusion is based on measurements of ACS activity and mRNA levels, as well as the interactions with various inducers of ethylene biosynthesis. This is consistent with previous reports of posttranscriptional regulation of ACS in etiolated Arabidopsis seedlings (Vogel et al., 1998b), and suggests that this may be a major mechanism for modulating the level of ethylene biosynthesis in this tissue. Alternatively, it is possible that an additional ACS gene(s) is present in Arabidopsis and that the transcription of this gene is elevated in the *Eto* mutants. However, if this were the case, one would predict an additive interaction of these mutations with inducers that act through independent ACS genes, which is clearly not the case with either auxin or cytokinin. Thus, the model that is most consistent with the data presented here is that the *Eto* mutations affect the posttranscriptional regulation of ACS.

One possible mechanism for this posttranscriptional regulation is that ACS protein could be modified to increase its activity or stability, and the *Eto* mutations affect this modification. Protein phosphorylation has been found to play a

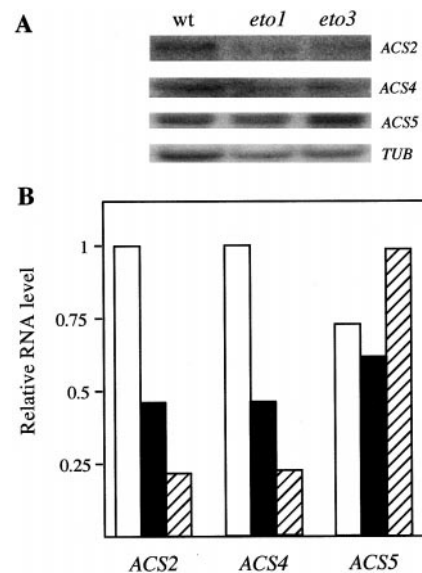


Figure 7. RNA-blot analysis of ACS mRNA levels in etiolated seedlings. Five micrograms of poly(A⁺) RNA from 3-d-old wild-type (open bars), *eto1* (closed bars), and *eto3* (hatched bars) seedlings was separated by agarose-gel electrophoresis, blotted to a nylon membrane, and hybridized to an ACS2, ACS4, ACS5, or β -tubulin (*TUB*) probe. A, The original images of the blots. B, The quantification of the blots in A. The signal from each band was quantified using a phosphor imager. The value for each ACS band was divided by its β -tubulin loading control. The highest level in each set was assigned a value of 1, and the other values are expressed relative to this. The ACS5 blot was from an independent RNA blot and was normalized to its own β -tubulin loading control (not shown).

ABA has been linked to both decreased ACC oxidase levels and increased ACC conjugation (McKeon et al., 1982; Corbinau et al., 1989; Tan and Thimann, 1989). The difference in the effect of ABA on *Eto* mutants versus its effect on wild-type etiolated seedlings can be explained if regulation by ABA is only required under conditions of high ethylene production, conditions not normally found in germinating wild-type *Arabidopsis* seedlings. The *Eto* mutations do not display any other obvious defects in ABA responses. ABA may act to suppress excess ethylene production downstream of the wild-type gene product of both *Eto* mutations.

The emerging picture from these and other studies is that posttranscriptional events play an important role in regulating ethylene biosynthesis. The interactions of the *Eto* mutations with other regulators of ethylene biosynthesis suggest that the pathways regulating this biosynthetic pathway are complex, reflecting the multitude of regulatory inputs that affect ethylene biosynthesis. Cloning of the genes corresponding to the *eto1* and *eto3* mutations should shed further light on the role that these genes play in regulating ethylene biosynthesis.

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LITERATURE CITED

- Abel S, Nguyen MD, Chow W, Theologis A (1995) *ACS4*, a primary indoleacetic acid-responsive gene encoding 1-aminocyclopropane-1-carboxylate synthase in *Arabidopsis thaliana*. *J Biol Chem* **270**: 19093–19099
- Abeles FB, Morgan PW, Saltveit ME Jr (1992) Ethylene in Plant Biology. Academic Press, San Diego, CA
- Adams DO, Yang SF (1979) Ethylene biosynthesis: identification of 1-aminocyclopropane-1-carboxylic acid as an intermediate in the conversion of methionine to ethylene. *Proc Natl Acad Sci USA* **76**: 170–174
- Amrhein N, Schneebeck D, Skorupka H, Tophof S, Stockigt J (1981) Identification of a major metabolite of the ethylene precursor 1-aminocyclopropane-1-carboxylic acid in higher plants. *Naturwissenschaften* **68**: 619–620
- Ausubel FM, Brent R, Kingston RE, Moore DD, Seidman JG, Smith JA, Struhl K (1994) Current Protocols in Molecular Biology. John Wiley & Sons, New York
- Avni A, Baily BA, Mattoo AK, Anderson JD (1994) Induction of ethylene biosynthesis in *Nicotiana tabacum* by a *Trichoderma viride* xylanase is correlated to the accumulation of 1-aminocyclopropane-1-carboxylic acid (ACC) synthase and ACC oxidase transcripts. *Plant Physiol* **106**: 1049–1055
- Barry CS, Blume B, Bouzayen M, Cooper W, Hamilton AJ, Grierson D (1996) Differential expression of the 1-aminocyclopropane-1-carboxylate oxidase gene family of tomato. *Plant J* **9**: 525–535
- Bleecker AB, Estelle MA, Somerville C, Kende H (1988) Insensitivity to ethylene conferred by a dominant mutation in *Arabidopsis thaliana*. *Science* **241**: 1086–1089
- Botella JR, Arteca RN, Frangos JA (1995) A mechanical strain-induced 1-aminocyclopropane-1-carboxylate synthase gene. *Proc Natl Acad Sci USA* **92**: 1595–1598
- Botella JR, Schlaghauser CD, Arteca JM, Arteca RN, Phillips AT (1993) Identification of two new members of the 1-aminocyclopropane-1-carboxylate synthase-encoding multigene family in mung bean. *Gene* **123**: 249–253
- Corbinau F, Rudnicki RM, Come D (1989) ACC conversion to ethylene by sunflower seeds in relation to maturation, germination and thermodormancy. *Plant Growth Regul* **8**: 105–115
- Ecker JR (1995) The ethylene signal transduction pathway in plants. *Science* **268**: 667–675
- Finlayson SA, Lee IJ, Morgan PW (1998) Phytochrome B and the regulation of circadian ethylene production in sorghum. *Plant Physiol* **116**: 17–25
- Gomez-Lim MA, Valdes-Lopez V, Cruz-Hernandez A, Saucedo-Arias LJ (1993) Isolation and characterization of a gene involved in ethylene biosynthesis from *Arabidopsis thaliana*. *Gene* **134**: 217–221
- Guzman P, Ecker JR (1990) Exploiting the triple response of *Arabidopsis* to identify ethylene-related mutants. *Plant Cell* **2**: 513–523
- Hoffman NE, Yang SF, McKeon T (1982) Identification and metabolism of 1-(malonylamino)cyclopropane-1-carboxylic acid as a major conjugate of 1-aminocyclopropane-1-carboxylic acid, an ethylene precursor in higher plants. *Biochem Biophys Res Commun* **104**: 765–770
- Jiao X-Z, Philosoph-Hadas S, Su L-Y, Yang SF (1986) The conversion of 1-(malonylamino)cyclopropane-1-carboxylic acid to 1-aminocyclopropane-1-carboxylic acid in plant tissues. *Plant Physiol* **81**: 637–641
- Kende H (1993) Ethylene biosynthesis. *Annu Rev Plant Physiol Plant Mol Biol* **44**: 283–307
- Kieber JJ (1997a) The ethylene response pathway in *Arabidopsis*. *Annu Rev Plant Physiol Plant Mol Biol* **48**: 277–296
- Kieber JJ (1997b) The ethylene signal transduction pathway in *Arabidopsis*. *J Exp Bot* **48**: 211–218
- Kieber JJ, Rothenburg M, Roman G, Feldmann KA, Ecker JR (1993) *CTR1*, a negative regulator of the ethylene response pathway in *Arabidopsis*, encodes a member of the Raf family of protein kinases. *Cell* **72**: 427–441
- Kim W, Yang S (1994) Structure and expression of cDNAs encoding 1-aminocyclopropane-1-carboxylate oxidase homologs isolated from excised mung bean hypocotyls. *Planta* **194**: 223–229
- Kionka C, Amrhein N (1984) The enzymatic malonylation of 1-aminocyclopropane-1-carboxylic acid in homogenates of mung bean hypocotyls. *Planta* **194**: 226–235
- Lasserre E, Bouquin T, Hernandez JA, Bull J, Pech JC, Balague C (1996) Structure and expression of three genes encoding ACC oxidase homologs from melon (*Cucumis melo* L.). *Mol Gen Genet* **251**: 81–90
- Li N, Mattoo AK (1994) Deletion of the carboxyl-terminal region of 1-aminocyclopropane-1-carboxylic acid synthase, a key protein in the biosynthesis of ethylene, results in catalytically hyperactive monomeric enzyme. *J Biol Chem* **269**: 6908–6917
- Liang X, Abel S, Keller JA, Shen NF, Theologis A (1992) The 1-aminocyclopropane-1-carboxylate synthase gene family of *Arabidopsis thaliana*. *Proc Natl Acad Sci USA* **89**: 11046–11050
- Liang X, Oono Y, Shen NF, Köhler C, Li K, Scolnik PA, Theologis A (1995) Characterization of two members (*ACS1* and *ACS3*) of the 1-aminocyclopropane-1-carboxylate synthase gene family of *Arabidopsis thaliana*. *Gene* **167**: 17–24
- Liang X, Shen NF, Theologis A (1996) Li^+ -regulated 1-aminocyclopropane-1-carboxylate synthase gene expression in *Arabidopsis thaliana*. *Plant J* **10**: 1027–1036
- Lizada MCC, Yang SF (1979) A simple and sensitive assay for 1-aminocyclopropane-1-carboxylic acid. *Anal Biochem* **100**: 140–145
- Martin MN, Cohen JD, Saftner RA (1995) A new 1-aminocyclopropane-1-carboxylic acid-conjugating activity in tomato fruit. *Plant Physiol* **109**: 917–926
- Mattoo AK, Suttle JC (1991) The Plant Hormone Ethylene. CRC Press, Boca Raton, FL
- McKeon TA, Hoffman NE, Yang SF (1982) The effect of plant-hormone pretreatments on ethylene production and synthesis of 1-aminocyclopropane-1-carboxylic acid in water-stressed wheat leaves. *Planta* **155**: 437–443

- Mekhedov SI, Kende H** (1996) Submergence enhances expression of a gene encoding 1-aminocyclopropane-1-carboxylate oxidase in deepwater rice. *Plant Cell Physiol* **37**: 531–537
- Nadeau JA, Zhang XS, Nair H, O'Neill SD** (1993) Temporal and spatial regulation of 1-aminocyclopropane-1-carboxylate oxidase in the pollination-induced senescence of orchid flowers. *Plant Physiol* **103**: 31–39
- Nakajima N, Nakagawa N, Imaseki H** (1990) Molecular size of wound-induced 1-aminocyclopropane-1-carboxylate synthase from *Cucurbita maxima* Duch. and change of translatable mRNA of the enzyme after wounding. *Plant Cell Physiol* **29**: 989–998
- Newman T, de Bruijn FJ, Green P, Keegstra K, Kende H, McIntosh L, Ohlrogge J, Raikhel N, Somerville S, Thomashow M, and others** (1994) Genes galore: a summary of methods for accessing results from large-scale partial sequencing of anonymous *Arabidopsis* cDNA clones. *Plant Physiol* **106**: 1241–1255
- Oetiker JH, Olsen DC, Shiu OY, Yang SF** (1997) Differential induction of seven 1-aminocyclopropane-1-carboxylate synthase genes by elicitor in suspension cultures of tomato (*Lycopersicon esculentum*). *Plant Mol Biol* **34**: 275–286
- Olson DC, Oetiker JH, Yang SF** (1995) Analysis of *LE-ACS3*, a 1-aminocyclopropane-1-carboxylic acid synthase gene expressed during flooding in the roots of tomato plants. *J Biol Chem* **270**: 14056–14061
- Olson DC, White JA, Edelman L, Harkins RN, Kende H** (1991) Differential expression of two genes for 1-aminocyclopropane-1-carboxylate synthase in tomato fruits. *Proc Natl Acad Sci USA* **88**: 5340–5344
- Peck SC, Kende H** (1995) Sequential induction of the ethylene biosynthetic enzymes by indole-3-acetic acid in etiolated peas. *Plant Mol Biol* **28**: 293–301
- Peiser G, Yang SF** (1998) Evidence for 1-(malonylamino) cyclopropane-1-carboxylic acid being the major conjugate of 1-aminocyclopropane-1-carboxylic acid in tomato fruit. *Plant Physiol* **116**: 1527–1532
- Rodrigues-Pousada RA, De Rycke R, Dedonder A, van Caeneghem W, Engler G, van Montagu M, Van der Straeten D** (1993) The *Arabidopsis* 1-aminocyclopropane-1-carboxylate synthase gene 1 is expressed during early development. *Plant Cell* **5**: 897–911
- Roman G, Lubarsky B, Kieber JJ, Rothenberg M, Ecker JR** (1995) Genetic analysis of ethylene signal transduction in *Arabidopsis thaliana*: five novel mutant loci integrated into a stress response pathway. *Genetics* **139**: 1393–1409
- Rottmann WH, Peter GF, Oeller PW, Keller JA, Shen NF, Nagy BP, Taylor LP, Campbell AD, Theologis A** (1991) 1-Aminocyclopropane-1-carboxylate synthase in tomato is encoded by a multigene family whose transcription is induced during fruit and floral senescence. *J Mol Biol* **222**: 937–961
- Spanu P, Grosskopf DG, Felix G, Boller T** (1994) The apparent turnover of 1-aminocyclopropane-1-carboxylate synthase in tomato cells is regulated by protein phosphorylation and dephosphorylation. *Plant Physiol* **106**: 529–535
- Tan Z-Y, Thimann KV** (1989) The roles of carbon dioxide and abscisic acid in the production of ethylene. *Physiol Plant* **75**: 13–19
- Tang X, Gomes AMTR, Bhatia A, Woodson WR** (1994) Pistil-specific and ethylene-regulated expression of 1-aminocyclopropane-1-carboxylate oxidase gene in petunia flowers. *Plant Cell* **6**: 1227–1239
- Vahala J, Schlaghaufer CD, Pell EJ** (1998) Induction of an ACC synthase cDNA by ozone in light-grown *Arabidopsis thaliana* leaves. *Physiol Plant* **103**: 45–50
- Van der Straeten D, Rodrigues-Pousada RA, Villarreal R, Hanley S, Goodman HM, van Montagu M** (1992) Cloning, genetic mapping, and expression analysis of an *Arabidopsis thaliana* gene that encodes 1-aminocyclopropane-1-carboxylate synthase. *Proc Natl Acad Sci USA* **89**: 9969–9973
- Vogel JP, Schuerman P, Woeste KW, Brandstatter I, Kieber JJ** (1998a) Isolation and characterization of *Arabidopsis* mutants defective in induction of ethylene biosynthesis by cytokinin. *Genetics* **149**: 417–427
- Vogel JP, Woeste KW, Theologis A, Kieber JJ** (1998b) Recessive and dominant mutations in the ethylene biosynthetic gene *ACS5* of *Arabidopsis* confer cytokinin insensitivity and ethylene overproduction, respectively. *Proc Natl Acad Sci USA* **95**: 4766–4771
- Woeste K, Vogel JP, Kieber JJ** (1999) Factors regulating ethylene biosynthesis in etiolated *Arabidopsis thaliana* seedlings. *Physiol Plant* (in press)
- Wright STC** (1980) The effect of plant growth regulator treatments on the levels of ethylene emanating from excised turgid and wilted wheat leaves. *Planta* **148**: 381–388
- Yang SF, Hoffman NE** (1984) Ethylene biosynthesis and its regulation in higher plants. *Annu Rev Plant Physiol* **35**: 155–189
- Yoshii H, Imaseki H** (1981) Biosynthesis of auxin-induced ethylene: effects of indole-3-acetic acid, benzyladenine and abscisic acid on endogenous levels of 1-aminocyclopropane-1-carboxylic acid (ACC) and ACC synthase. *Plant Cell Physiol* **22**: 369–379