Axillary Meristem Development. Budding Relationships between Networks Controlling Flowering, Branching, and Photoperiod Responsiveness

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Morphology in many animals is preordained during embryonic development and remains unchanged by environment. In contrast, vast differences in phenotype can occur in plants of identical genotype in different environments. Being sessile organisms, plants must rely on morphological and physiological plasticity to cope with a variable environment. The basis for this ability is the maintenance of numerous pluripotent stem cell clusters called meristems.

During embryonic development, plants produce a shoot and a root apical meristem. It is unclear whether axillary meristems are produced de novo in leaf axils or whether they are derived from the apical meristem of the primary shoot (Gribić and Bleecker, 2000). Variation in the timing of the initiation and development of axillary meristems can be observed by comparing the rosette crucifer, Arabidopsis, with the caulescent legume, pea (Pisum sativum). Arabidopsis has delayed axillary meristem initiation, causing some nodes to be devoid of axillary meristems (Gribić and Bleecker, 2000). Subsequent development of these meristems may also be delayed such that a pronounced axillary bud is often not observed. Pea develops axillary meristems at most nodes along its stem, and development usually proceeds apparently uninhibited up to the stage of a dormant bud that consists of several undeveloped leaves and internodes.

The nodes at which axillary meristems and/or branches occur along the stem are influenced by photoperiod in pea (Arumingtyas et al., 1992; Napoli et al., 1999) and Arabidopsis (Gribić and Bleecker, 2000; Stirnberg et al., 2002). The formation of basal branches in pea is enhanced under short photoperiods (Fig. 1, A and B). Bud outgrowth at upper nodes in pea often occurs at the onset of flowering and may also be, directly or indirectly, under photoperiod control. Although the formation of branches in Arabidopsis is somewhat constrained until the floral transition, this species shows similar photoperiod responses in the node of axillary bud initiation and development to those observed for branching in pea (Gribić and Bleecker, 2000; Stirnberg et al., 2002).

Determination of the axillary meristem as either vegetative or floral is a key step in regulating plant architecture and involves interactions among genotype, environmental cues, and endogenous phytohormone-like signals. Once the identity of an axillary meristem is determined, a further developmental program acts locally to maintain that determination and to prescribe organ identity within the axillary structure. However, long and short-range signals control not only the specification and maintenance of meristem identity but also organ outgrowth, thus exerting a major influence on whether axillary meristems reach their potential to form a mature branch or an inflorescence-bearing fruit.

The suitability of pea for investigating long-distance signaling makes it a valuable tool for elucidating the coordinate regulation of axillary meristem development. Its long internodes separating nodes in vegetative and reproductive zones and its ample root size for xylem sap extraction make it suitable for endogenous and exogenous phytohormone studies at several developmental stages. Moreover, in contrast to Arabidopsis, which does not respond to exogenous auxin after decapitation (Cline, 1996), pea shows a typical strong apical dominance phenotype and does respond to auxin after decapitation (e.g. Beveridge et al., 2000). Whereas grafting has only recently been successfully applied to Arabidopsis (Turnbull et al., 2002), pea is readily amenable to many different graft unions, allowing the production of genetic chimeras without the complication of adventitious rooting.

The phenotypes of various mutants discussed herein indicate that apical and axillary meristems are, to some extent, independently regulated. Genetic and physiological analysis of flowering time and shoot...
architecture mutants in garden pea has identified three interacting networks (Fig. 2). Two of these, the vegetative and floral meristem networks, are devoted specifically to axillary meristem identity and/or subsequent development. A third, the photoperiod network, controls the developmental strategy of the whole plant in response to daylength and coordinately regulates both vegetative traits and flowering. This Update will discuss these regulatory networks with an emphasis on the involvement of long-distance signals.

**VEGETATIVE MERISTEM DEVELOPMENT NETWORK**

In many plants, and particularly in weakly branching monopodial plants such as pea, rapid outgrowth of axillary buds after decapitation allows the plant to maintain vigorous growth under competitive conditions and may be essential to provide replacement sites for reproductive development. However, it is also clear that this response must be regulated, because indiscriminate bud outgrowth could quickly lead to deleterious shading and might also divert resources away from reproductive structures developing elsewhere on the plant. This implies communication among axillary buds and between axillary buds and the shoot tip. Gene expression and protein profiling studies in pea have revealed that removal of the shoot tip induces axillary buds to enter a transition state between dormancy and growth (Fig. 2A) and that whether buds subsequently revert back to the dormant state or proceed to sustained growth is influenced by the state of other buds (Stafstrom et al., 1998; Shimazato-Sato and Mori, 2001).

Until recently, the most widely accepted hypothesis on the role of systemic signals in regulating bud outgrowth was that auxin derived in shoot tips and young leaves acts indirectly to inhibit branching by decreasing cytokinin supply to buds (e.g., Cline, 1994). Central to recent progress to expand this simplistic hypothesis has been the identification of mutants that differ from the wild type (WT) primarily due to enhanced development of vegetative axillary meristems. The most comprehensive genetic and physiological analysis of shoot branching in plants has been performed with the *ramosus* series of branching mutants in pea (Arumingtyas et al., 1992; Rameau et al., 2002). Studies with these *rms* mutants have revealed a more complex regulatory network by demonstrating involvement of long-distance signals in addition to auxin and cytokinins (Beveridge et al., 2000; Fig. 2A).

Nonallelic mutants *rms1* to *rms5* exhibit increased branching at basal and aerial nodes, whereas *rms6* mutants branch at basal nodes only. The *rms* mutants enhance rather than override the ontogenetic variation in tendency for bud outgrowth exhibited by WT plants. As in WT plants, the pattern of bud outgrowth in mutants *rms1* to *rms5* remains strongly influenced by photoperiod, with a decrease in basal branching and an increase in aerial branching under long days (LD) compared with short days (SD; Fig. 3; Arumingtyas et al., 1992).

**rms** Mutants Reveal Involvement of Novel Signals

Grafting studies with three of the pea mutants (*rms1*, *rms2*, and *rms3*) have demonstrated clear roles for long-distance signals in the control of bud outgrowth and have shown regulation by genes acting in shoot and/or stem and root (Beveridge et al., 1997; Morris et al., 2001). Similar results have been obtained with recently isolated Arabidopsis branching mutants, *max1* and *max3*, and the *dad1* branching mutant of petunia (*Petunia hybrida*), providing further evidence that axillary bud outgrowth is not under control of the shoot tip alone (Napol, 1996; Turnbull et al., 2002). Grafting experiments in pea have
shown that RMS1 and RMS5 may act in the same biochemical pathway for a signal that acts like an inhibitor and moves only acropetally in shoots, presumably through the xylem (Foo et al., 2001; Morris et al., 2001; Fig. 2A).

The rms mutants have been used to determine the possible phytohormone basis of these graft-transmissible signals (for review, see Beveridge, 2000; Morris et al., 2001). The signal regulated by RMS1 and RMS5 is unlikely to be either cytokinin or auxin for several reasons. For example, rms1 and rms5 plants have greatly reduced, rather than elevated, xylem sap cytokinin concentrations (Beveridge, 2000; Morris et al., 2001). Also auxin and auxin precursors are not thought to be carried in the xylem. Moreover, auxin content in the shoot of rms mutants is typically elevated, not reduced, and exogenous auxin does not restore a WT phenotype to rms plants (Beveridge et al., 1997, 2000; Beveridge, 2000). These results indicate that RMS1 and RMS5 may regulate a novel signal (Fig. 2A). The recent cloning of RMS1 after isolation of the MAX4 sequence from the new max series of branching mutants from Arabidopsis and isolation of a MAX4 homolog from Medicago truncatula (K. Sorefan, J. Booker, K. Haurogne, M. Goussot, E. Foo, S. Chatfield, C. Beveridge, C. Rameau, and O. Leyser, unpublished data) will be reported elsewhere and opens new avenues to test this hypothesis.

The sequence of an additional MAX gene, MAX2, has been reported and encodes an F-box protein that may be involved in signal transduction (Woo et al., 2001; Stirnberg et al., 2002). The obvious candidates for a pea ortholog of MAX2 are therefore genes such as RMS3 or RMS4 that appear to act mostly in the shoot and that are proposed to control the response to signals involved in branching control (for review, see Beveridge, 2000; Fig. 2A).

What Is the Role of Auxin?

The auxin inhibition of bud outgrowth in decapitated plants appears to require the long-distance signal regulated by RMS1 (Fig. 2A). Decapitated rms1 mutant shoots can only respond to exogenous auxin when grafted to WT rootstocks (Beveridge et al., 2000). Moreover, RMS1 may be auxin regulated because RMS1 expression drops after decapitation and is restored by exogenous auxin (E. Foo, C. Beveridge, and C. Rameau, unpublished data).

The possibility that other phytohormones or environmental cues may directly or indirectly regulate

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**Figure 2.** Regulatory networks controlling vegetative and floral meristem development in pea. The vegetative (A), photoperiod response (B), and reproductive (C) development networks are shown. Arrows between the networks show hypotheses of where the points of coordinate regulation may occur.

**Figure 3.** Lateral lengths at nodes of decapitated WT (pea cv Parvus) and intact rms1-1 and rms2-2 plants (left to right). Plants received 8 h of natural daylight followed by darkness or light extension to 18 h supplied by a 1:1 mixture of fluorescent (40-W, white) and incandescent (100-W) lights providing an intensity of 25 to 30 μmol m⁻² s⁻¹ at the pot top. Intact WT plants did not produce laterals greater than 1 cm. Nodes are counted from the cotyledonary node as node 1. Decapitation was performed below the highest expanded leaf, 7 d before scoring. All plants were scored on d 36 when intact mutant plants under LD and SD had 13 to 14 and 11 leaves expanded, respectively. The first flower opened under LD at node 16. Data are presented as mean ± se; n = 5–6.
RMS1 expression, RMS1 protein stability, precursor availability, or product degradation and therefore affect levels of the shoot-to-root signal regulated by RMS1, should be investigated. This could reveal how bud outgrowth may be regulated via long-distance signals that do not cause auxin-related pleiotropic effects.

In pea, the bushy mutant and several late flowering mutants under certain conditions (Fig. 1B and see below) have a pleiotropic highly branched and dwarfed phenotype (Beveridge et al., 2001; Symons et al., 2002). In contrast to rms mutants, which have vigorous shoot tip growth and do not have depleted auxin levels, these pleiotropic phenotypes are related to weak shoot tip growth and may be at least partly attributed to reduced endogenous auxin levels (Beveridge et al., 2001; Symons et al., 2002).

In addition to the auxin-regulation of RMS1, auto-regulation of bud outgrowth may also involve auxin-independent modulation of xylem sap cytokinin content via a shoot-to-root signal. Several of the rms mutants show strongly reduced xylem sap cytokinin concentration. Graft combinations of WT and rms3 or rms4 reveal that the branching phenotype of the shoot is associated with the rate of cytokinin export from the roots, regardless of the root genotype, implying involvement of a shoot-to-root feedback signal (Beveridge et al., 1997; Beveridge, 2000; Fig. 2A). This shoot-to-root signal is unlikely to be indole-3-acetic acid, because indole-3-acetic acid levels and transport are not greatly affected in these genotypes. RMS2 may regulate the feedback signal because rms2 plants have elevated xylem sap cytokinin content, and double mutants show that rms1 and rms5 do not cause reduced xylem sap cytokinin content in the presence of rms2 (Beveridge et al., 1997; Morris et al., 2002).

**Figure 4.** Typical pleiotropic effects of photoperiod response in pea. A, Node of flower initiation (NFI) and number of reproductive nodes (RN); B, length of the primary stem between nodes 1 to 9; C, node of flower opening relative to the node of the highest expanded leaf; D, number of branches at nodes 1 to 3. The highly photoperiod responsive (SN DNE PPD HR: H63) line was grown under 8 h of daylight extended to 12 or 16 h with incandescent light at 55 μmol m⁻² s⁻¹ or to 24 h with incandescent light at 3 μmol m⁻² s⁻¹. Data are means ± se for six plants per treatment. *, Plants grown under 12 h were scored well before senescence and would have developed considerably more reproductive nodes (RN) than shown.

**PHOTOPERIOD RESPONSE NETWORK**

The effects of photoperiod on the initiation of flowering are well known. Perhaps less widely recognized is that, in many species, including temperate LD plants such as Arabidopsis and pea and SD plants such as common bean (*Phaseolus vulgaris*), photoperiod also affects vegetative shoot architecture (Figs. 1 and 3) and a range of other vegetative and reproductive characteristics (Wallace et al., 1993; Fig. 4).

Under inductive conditions for flowering, the resources of the plant are directed toward rapid completion of the life cycle. The growth habit under noninductive photoperiods can be understood as a strategy that prevents the plant from investing too much energy in reproduction under unfavorable conditions and prepares it to exploit a subsequent improvement in conditions by increasing the photosynthetic area and the number of sites available for reproduction. The differences in phenotype between plants grown in SD and LD cannot be explained solely by the earlier flowering and enhanced sink activity of developing flowers and fruits in plants grown under LD, because effects of photoperiod on vegetative traits can be clearly seen even in mutants that fail to initiate flowers under any photoperiod (Reid and Murfet, 1984; Kelly and Davies, 1988).

**Photoperiod Response Genes Control Long-Distance Signal(s)**

Although photoperiod controls many different traits, genetic analyses show that responsiveness to photoperiod depends on a common mechanism. Recessive mutations that reduce or eliminate photoperiod responsiveness (day-neutral) are known in several legumes. In most cases, these mutants are early flowering and display a “constitutively reproductive” growth habit, having lost the ability to stimulate vegetative growth and inhibit flowering under noninductive conditions. Mutants known from LD species include sn (Fig. 1, A and B), dne, and ppd in pea, dn in sweet pea, and sn in lentil (*Lens culinaris*;
Murfet, 1971; King and Murfet, 1985; Ross and Murfet, 1988; Sarker et al., 1999). The ppd mutant of
common bean, a SD species, also shows a similar phenotype (Wallace et al., 1993).

The involvement of long-distance signaling in the
control of photoperiod responses has been a topic
of interest for several decades. Day-neutral mutants in
pea and sweet pea have been used to establish a
genetic basis for long-distance signaling in photope-
riod responsiveness. For example, day-neutral mu-
tants sn, dne, and ppd show a delay in flowering and
an enhancement of vegetative vigor when grafted
onto WT rootstocks, suggesting that the mutations
somehow disrupt the supply of a mobile signal to
the apex by interfering with its synthesis or trans-
port (Murfet, 1985). Studies with these mutants have
also reinforced the idea that the mobile signal con-
trols multiple aspects of development, including
flowering and branching (Fig. 2). Grafting experi-
ments have also shown that the inhibitory influence
of leaves declines with leaf age (Reid and Murfet,
1977).

Studies with two other genes that affect photope-
riod response indicate that developmental and
tissue-specific regulation of the level of the mobile
signal is an important feature of the photoperiod
response in pea. Dominant alleles of the HIGH RE-
SPONSE (HR) gene occur in many primitive acces-
sions and increase the size of the photoperiod re-
sponse for flowering, mainly by delaying flowering
under SD (Murfet, 1985). Results from grafting ex-
periments suggest that rather than increasing inhib-
itor production, HR extends the time span over
which leaves remain inhibitory (Reid and Murfet,
1977). Another gene, EARLY (E), decreases inhibitor
production and acts only in the cotyledons. It may
cause early floral initiation in some genetic back-
grounds, followed by a period of inflorescence abor-
tion or vegetative reversion after the cotyledons se-
sesce and the photoperiod response of the shoot
becomes dominant (Murfet, 1985).

Mutants have been used to investigate the role of
specific photoreceptors in the regulation of long-
distance signaling. Plants deficient in phytochrome A
(phyA) are very similar to WT when grown under
SD, but under LD, they flower late and show the full
range of pleiotropic characteristics typical of WT
plants grown in SD (Weller et al., 1997). Leafy phyA
rootstocks delay flowering in WT scions under LD,
confirming that phyA contributes to the down-
regulation of a mobile inhibitor of flowering under
LD (Fig. 2B). However, flowering of phyA mutants is
still strongly promoted by day extensions with cer-
tain light sources, and at least one cryptochrome is
probably also involved in this response (Weller et al.,
2001). Phytochrome B is required for inhibition of
flowering under SD, but does not affect photoperiod
responsiveness for other traits, and its inhibitory ef-
fects on flowering are not graft transmissible (Weller
et al., 2001; Fig. 2C).

Comparing Photoperiod Response Systems in
Pea and Arabidopsis

Apart from PHYA, none of the pea genes involved
in the photoperiod response have been cloned, and
the nature of the inhibitory signal is still unknown.
Further progress in understanding the functions of
these genes will come from comparisons with Arabi-
dopsis, where the photoperiod pathway is well char-
acterized and many genes involved in the photope-
riod response have been cloned. In Arabidopsis, light
and the circadian clock interact to regulate the ex-
pression of genes that specifically promote or inhibit
flowering (Mouradov et al., 2002). This pathway con-
ists of genes that either promote or inhibit flower-
ing. Among the genes that promote flowering are
those encoding known photoreceptors (PHYA and
CRY2), putative photoreceptors involved in light sig-
aling to the clock (ZTL, FKF, and LKP2) and genes
pecific for the floral transition (CO, FT, FD, and FE).
The genes that inhibit flowering all appear to func-
tion close to clock mechanism (LHY, CCA1, TOC1,
ELF3, and ELF4).

The fact that most of the known photoperiod re-
ponse genes in pea have an inhibitory effect on
flowering (SN, DNE, PPD, and HR) raises the possi-
bility that this system in pea might correspond to the
circadian system in Arabidopsis. The Arabidopsis
circadian system is implicated in the control of a
wide range of processes, so this possibility is consis-
tent with the fact that the pea genes have pleiotrop-
effects. Although the pea system has frequently been
discussed as if it controlled a single mobile signal, it
might equally represent a mechanism for the regula-
tion of a large number of genes, some of which could
be associated with long-distance signaling in specific
developmental responses (Fig. 2).

FLORAL MERISTEM DEVELOPMENT NETWORK

Genetic and physiological studies indicate that the
photoperiod response network interacts with net-
works of genes that are specific to axillary bud out-
growth or reproductive development (Fig. 2). That is,
genomes acting in these vegetative or reproductive spe-
cific networks do not affect photoperiod responsiv-
ness. Like bud outgrowth, the transition to flowering
involves specific long-distance signals.

Although the transition to flowering in legumes
may appear to be simply the replacement of vegeta-
tive axillary meristems with reproductive axillary
meristems, the primary shoot apical meristem must
first make the transition from a vegetative to an
inflorescence meristem (Fig. 1C). In pea, inflores-
cence fate can be determined as early as eight nodes
before a floral meristem forms (Ferguson et al., 1991).

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Commitment to a program of inflorescence development is thus separable and distinct from determination for floral development. Analysis of mutants has identified genes that may be involved in these transitions, and grafting studies are being used to separate those genes acting in long-distance signaling from those acting locally.

A Mobile Signal Is Specifically Required for Transition to Flowering

In addition to the inhibitory signal involved in photoperiod responsiveness, there is a large body of physiological evidence from many species supporting the existence of a mobile flowering stimulus (Murfet, 1971). Genetic evidence for such a promoter has more recently been demonstrated in pea (Beveridge and Murfet, 1996). Recessive mutants at the GI GAS (GI) locus can show a photoperiod response in flowering node, but show a large delay in flowering under SD, and flower late or not at all under LD. In addition to differing in flowering response to photoperiod, gi mutants differ from the late flowering phyA mutants, because vegetative traits in gi plants respond normally to photoperiod. GI is proposed to have a role in long-distance signaling (Fig. 2C) because flowering can be partially restored to gi mutant scions by grafting to a WT stock (Beveridge and Murfet, 1996).

Development of gi mutant plants in LD proceeds relatively normally until around the time of flower initiation in WT, after which the mutants appear to lose apical dominance and become highly branched at aerial nodes (Fig. 1B). The change in growth pattern occurs earlier and is more severe in an sn background, and it may thus reflect the natural decline in the activity of the photoperiod pathway in the absence of inflorescence development (Murfet, 1985; Beveridge et al., 2001).

A Module of Genes with Overlapping Functions Acts Locally to Specify Inflorescence and Floral Identity

Like GI, the LATE-FLOWERING (LF) gene appears to specifically affect the transition to flowering. Recessive mutants at the LF locus flower earlier than WT in both LD and SD (Fig. 1, A and B). Whereas GI controls a mobile stimulus, LF has an inhibitory effect and acts only in the shoot (Murfet, 1985). Plants carrying extreme-late, dominant LF alleles show a reduced apical dominance phenotype under LD that is very similar to that seen in gi mutants. Also, like GI, LF alleles have an essentially additive interaction with SN. These observations suggest that GI and LF may act in the same flowering-specific pathway (Fig. 2C). Consistent with this suggestion, a strong lf allele is completely epistatic to gi under SD (Taylor, 1997). The GI-LF interaction may therefore define an important point at which a mobile signal from the leaves could interact with a genetic module controlling inflorescence meristem identity in the shoot apex.

Severe mutants at the VEGETATIVE 1 (VEGI) and VEGETATIVE 2 (VEG2) loci never flower and yet show a photoperiod response for vegetative traits. In SD, these mutants have a relatively normal appearance, apart from the failure to initiate secondary inflorescences. In LD, they display the same aerial branching phenotype as gi mutants (Fig. 1B) and plants carrying extreme-late LF alleles. The non-flowering phenotypes of veg1 and veg2 plants cannot be overcome by grafting to WT, indicating that like LF, VEG1 and VEG2 act locally in the shoot apex. Both veg1 and veg2 are epistatic to LF (Reid and Murfet, 1984; Taylor, 1997), suggesting that these genes are required for LF function, which may be to repress VEG1 and VEG2 expression in newly initiated axillary meristems (Fig. 2C).

Another gene that acts locally to control inflorescence and flower development downstream of LF is DETERMINATE (DET). DET appears to maintain the indeterminacy of growth in the primary shoot inflorescence meristem (II; see Fig. 1C; Singer et al., 1990). Development of det mutants proceeds normally until one or two essentially normal axillary secondary inflorescences have been produced (II; Fig. 1C), each bearing one or more individual flowers and terminating in a stub. After this, the det shoot apex itself develops the characteristics of a secondary inflorescence, producing a flower and terminating in a stub (Fig. 1C), whereas the WT primary shoot inflorescence meristem remains indeterminate. This suggests that DET acts specifically in the main shoot apex, to suppress secondary inflorescence development, possibly by excluding VEG1 and VEG2 activity (Fig. 2C). The det mutant phenotype suggests that DET may be homologous to TFL and CEN from Arabidopsis and Antirrhinum sp., respectively (Bradley et al., 1997).

Three homologs of floral meristem identity genes in other species have been characterized in pea: PROLIFERATING INFLOWER ICE MERISTÈM (PIM), STAMINA PISTILLOIDA (STP), and UNIFOLIATA (UNI; Hofer et al., 1997; Taylor et al., 2001, 2002). The pea homolog of the MADS-box gene AP1 from Arabidopsis, PIM, specifies floral meristem identity but has minimal influence on the phenotype of the secondary inflorescence (Fig. 2C), pim mutants develop a secondary inflorescence with a terminal stub, but the third-order branch develops as an inflorescence rather than a flower (Fig. 1C). Repetitive inflorescence branching leads to formation of an aberrant flower. The extent and pattern of the branching in later order branches of pim mutants depends on photoperiod, providing more evidence that the photoperiod system also operates at relatively late stages of flower development (Taylor et al., 2002).

The apparent independence of floral and inflorescence developmental networks is shown by the continuation of inflorescence development in the ab-
cence development. The strongest mental genes can have on vegetative and inflorescence development.

PIM also has a minimal role in secondary inflorescence commitment and architecture in pea, particularly photoperiod-responsive late lines is likely to lead to new loci in pea, whereas work in pea has characterized long-distance signaling mechanisms that are yet to be identified in Arabidopsis. The legume genomics projects (see VandenBosch and Stacey, 2003; this issue) will facilitate this comparative analysis by enabling rapid cloning of pea homologs of genes cloned in other species.

New opportunities to understand plant development arise from recognizing the high degree of regulation among different parts of the developmental network. Here, we have identified coordinate regulation of flowering and branching by a photoperiod response module. Studies of the properties of the entire developmental network (including modules not covered here) are likely to yield new knowledge about how perturbations in one part of the network affect other parts. Computational approaches to model and test hypotheses concerning components of the system and their interactions may be useful to conceptualize the whole system and to incorporate a rapidly increasing body of knowledge.

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CONCLUDING REMARKS

Long-distance signals are involved in the regulation of many aspects of axillary meristem development, including vegetative axillary meristem development and the determination and development of primary and secondary inflorescence meristems. In the control of vegetative branching, long-distance signals form an autoregulatory loop whereby the outgrowth of buds feeds back to down-regulate the outgrowth of other buds. It is interesting to note that autoregulation by long-distance signals is also an important part of the control of nodule meristem development (see Szczygowski and Amyot, 2003; this issue). A long-awaited breakthrough in this field of research will be identification of novel long-distance signals (see Dixon and Sumner, 2003; this issue). It will also be important to understand how the major phytohormones such as auxin interact with these signals and contribute to the coordinated regulation of multiple aspects of plant development.

An enhanced rate of progress on the research of all aspects of axillary meristem development in legumes is likely to result from integrated studies among different species. For example, work in Arabidopsis suggests that mutagenesis for flowering mutants in pea, particularly photoperiod-responsive late lines is likely to lead to new loci in pea, whereas work in pea has characterized long-distance signaling mechanisms that are yet to be identified in Arabidopsis. The legume genomics projects (see VandenBosch and Stacey, 2003; this issue) will facilitate this comparative analysis by enabling rapid cloning of pea homologs of genes cloned in other species.
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