

Interaction of Auxin and ERECTA in Elaborating Arabidopsis Inflorescence Architecture Revealed by the Activation Tagging of a New Member of the YUCCA Family Putative Flavin Monooxygenases¹

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The aboveground body of higher plants has a modular structure of repeating units, or phytomers. As such, the position, size, and shape of the individual phytomer dictate the plant architecture. The Arabidopsis (*Arabidopsis thaliana*) *ERECTA* (*ER*) gene regulates the inflorescence architecture by affecting elongation of the internode and pedicels, as well as the shape of lateral organs. A large-scale activation-tagging genetic screen was conducted in Arabidopsis to identify novel genes and pathways that interact with the *ER* locus. A dominant mutant, *super1-D*, was isolated as a nearly complete suppressor of a partial loss-of-function allele *er-103*. We found that *SUPER1* encodes *YUCCA5*, a novel member of the YUCCA family of flavin monooxygenases. The activation tagging of *YUCCA5* conferred increased levels of free indole acetic acid, increased auxin response, and mild phenotypic characteristics of auxin overproducers, such as elongated hypocotyls, epinastic cotyledons, and narrow leaves. Both genetic and cellular analyses indicate that auxin and the ER pathway regulate cell division and cell expansion in a largely independent but overlapping manner during elaboration of inflorescence architecture.

The aboveground body of higher plants is a consequence of the continual activity of the shoot apical meristem (SAM), which generates repeating units called phytomers. Each phytomer is composed of a node, stem (internode), leaf (lateral organ), and axillary bud, the latter of which allows branching. Modification of the position, size, and shape of the individual phytomer provides immense variations in plant architecture. Such diversity in plant architecture has significance in the domestication and breeding of crop plants. For instance, the maize (*Zea mays*) *teosinte branched1* gene regulates branching pattern, and alteration in the *teosinte branched1* expression level played a pivotal role in the domestication of maize (spp. *mays*) from its wild ancestor, Teosinte (maize spp. *Parviglumis*; Doebley et al., 1997). Another example is semi-dwarf varieties of cereals, in which reduction in the

internode length has directly contributed to dramatic yield increase (Chrispeels and Sadava, 1994; Peng et al., 1999).

The model plant Arabidopsis (*Arabidopsis thaliana*) forms a typical rosette. During the vegetative stage, the Arabidopsis plant produces rosette leaves with no apparent internodal elongation. As the SAM acquires reproductive characteristics, the primary inflorescence stem elongates rapidly (bolting). First, the SAM gives rise to a shoot that has both vegetative and reproductive characteristics, including bracts (cauline leaves), axillary buds, and branches. Subsequently, the SAM generates multiple floral meristems, each of which differentiates a flower at the tip and a pedicel at the base, while the SAM itself maintains its indeterminate state (Schultz and Haughn, 1991).

Molecular-genetic studies have highlighted the *in vivo* role of phytohormones and growth regulators for normal growth of the inflorescence. For instance, mutants defective in biosynthesis or signaling pathways of auxin, gibberellin, brassinosteroids, and sporamin conferred dwarfism due to reduced cell elongation and/or cell division in the internode (Feldmann et al., 1989; Lincoln et al., 1990; Leyser et al., 1993; Li et al., 1996; Li and Chory, 1997; Azpiroz et al., 1998; Hedden and Proebsting, 1999; Hanzawa et al., 2000; Fridborg et al., 2001; Olszewski et al., 2002). Conversely, elevated response to these phytohormones, either by mutations in the downstream components of the signaling pathways or overexpression of the biosynthetic enzymes, led to extra elongation of the inflores-

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cence (Wang et al., 2001, 2002; Zhao et al., 2001; Olszewski et al., 2002).

Loss-of-function mutations in the Arabidopsis *ERECTA* (*ER*) gene dramatically modify inflorescence architecture due to altered organ shape and internodal elongation pattern (Torii et al., 1996; Komeda et al., 1998). Compared to the wild type, the young *erecta* (*er*) inflorescence forms flattened and somewhat umbellate floral bud clusters due to reduced internodal and pedicel elongation. At seed set, *er* forms a compact inflorescence with short and thick internodes, pedicels, and siliques. The *ER* gene encodes a Leu-rich repeat receptor-like kinase (RLK; Torii et al., 1996). Other members of the Leu-rich repeat-RLK superfamily are known to regulate diverse aspects of development, defense against pathogens, and wounding response, and their known ligands vary from short peptides (e.g. phytoalexin, systemin, and flagellin peptide) and proteins (CLAVATA3) to steroids (e.g. brassinolide; Matsubayashi et al., 2001; Ryan et al., 2002; Carles and Fletcher, 2003; Diévar and Clark, 2004; Torii, 2004). Although the ligands and downstream targets of *ER* are not known, recent studies revealed that *ER* and its two closely related paralogs interact in a synergistic manner in promoting cell proliferation, leading to aboveground organ growth (Shpak et al., 2003, 2004).

ER exhibits strong genetic interactions with multiple phytohormone and growth regulatory pathways. For example, *ER* was identified as a strong suppressor of dwarfism caused by two gibberellin response mutants, *gai* and *shi* (Peng et al., 1997; Fridborg et al., 2001). Indeed, the height of the *shi* inflorescence was nearly 6 times greater in the presence of functional *ER* (Fridborg et al., 2001). A similar effect of *ER* was observed for the dwarf phenotype of the auxin response mutant *transport inhibitor resistance3* (*tir3*)/*attenuated shade avoidance1* (Kanyuka et al., 2003). In either case, the cellular basis of *ER*-mediated suppression has not been investigated. *ER* also shows dramatic interaction with *BREVIPEDICELLUS* in specifying inflorescence architecture (Douglas et al., 2002). *BREVIPEDICELLUS* encodes KNAT1, a KNOTTED-like homeodomain protein that promotes internodal growth via modulating the expression of the lignin biosynthesis genes (Mele et al., 2003).

To better understand how *ER* promotes inflorescence growth, it is necessary to identify genes that interact with the *ER* pathway. We initiated an activation-tagging screen to isolate dominant, extragenic suppressors of the *er* inflorescence phenotype using a partial loss-of-function allele *er-103*. The strategy is based on a random insertion of 4× cauliflower mosaic virus (CaMV) 35S enhancers that, if inserted in cis-regulatory elements, will lead to up-regulation of nearby open reading frames (Kakimoto, 1996; Weigel et al., 2000). We discovered that the activation tagging of *YUCCA5*, a previously undescribed member of *YUCCA* family genes encoding a putative flavin monooxygenase, led to elevated auxin levels and a nearly complete suppression of *er-103* inflorescence elongation defects. Our detailed developmental and cellular analyses high-

lighted the complex interplay of auxin and the *ER*-mediated pathway in the elaboration of inflorescence architecture.

RESULTS

Isolation of suppressor of *er1-Dominant* as a Suppressor of *er-103*

To identify genes whose overexpression suppresses the *er* phenotype, a gain-of-function activation-tagging screen was performed using the intermediate allele *er-103* (Torii et al., 1996). Approximately 6,000 T1 Basta-resistant plants were visually screened for suppressed flower-clustering phenotype at bolting. One line, 38May2, displayed an inflorescence phenotype nearly identical to that of the wild-type *ER* (Fig. 1, A–C). A genetic analysis of the subsequent T2 generation revealed that the suppressor is dominant and tightly linked to Basta resistance, indicating that it is activation tagged by the 4 × CaMV enhancer (see “Material and Methods”). The mutant was named *suppressor of er1-Dominant* (*super1-D*).

super1-D Suppresses Internodal and Pedicel Elongation Defects of *er-103*

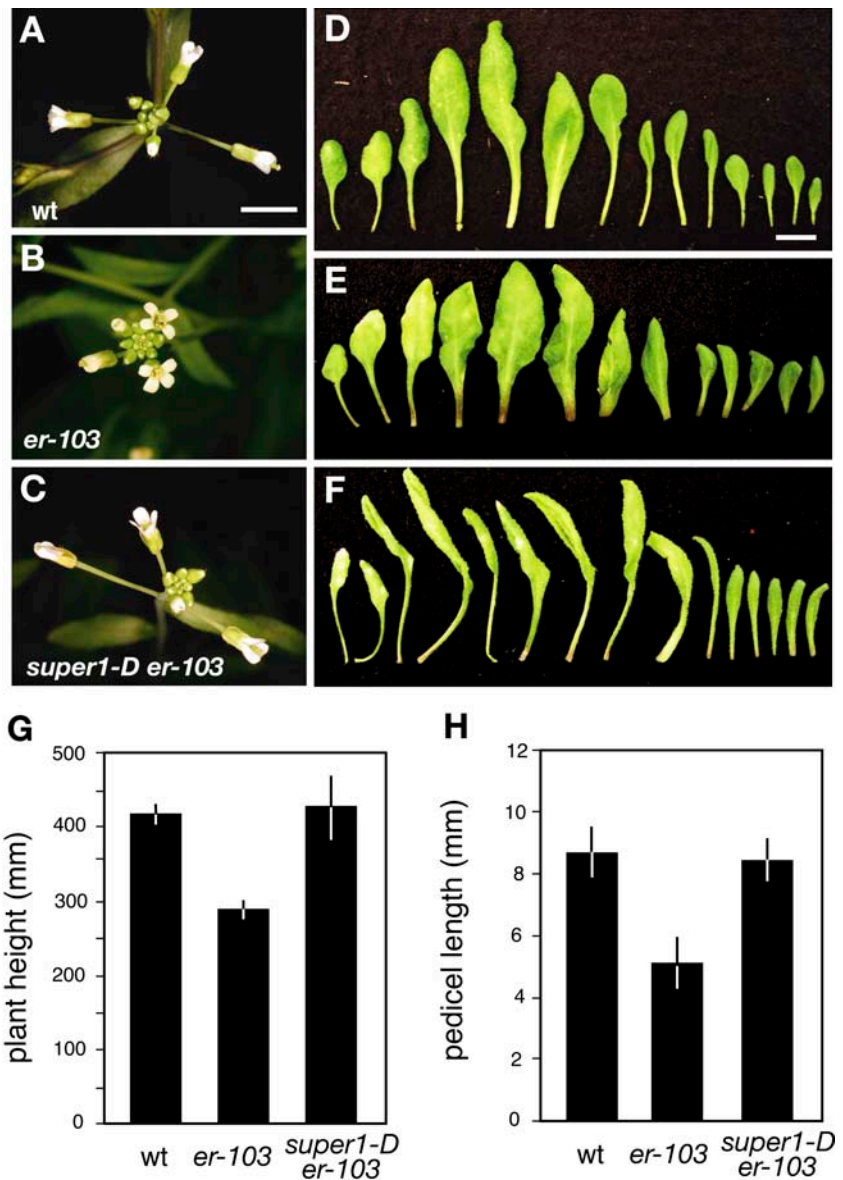
We subsequently analyzed the morphological phenotype of *super1-D er-103* during vegetative and reproductive development (Fig. 1). Unlike wild-type plants, *er-103* plants develop a compact rosette leaf with a short petiole and round blade (Fig. 1, D and E; Torii et al., 1996). The *super1-D* mutation suppressed the rosette phenotype of *er-103* and further conferred characteristic leaf morphology including elongated petioles and long, narrow blades that often curled down (Fig. 1F).

The inflorescence architecture of *super1-D er-103* highly resembles that of the wild type. Flower buds of the wild type fold on to each other to cover the SAM, and older flowers have elongated pedicels (Fig. 1A). In contrast, the inflorescence tip of *er-103* forms a characteristic disc-shaped, flat surface due to defective elongation of internodes and pedicels (Fig. 1B). *super1-D* suppressed the flower-clustered phenotype of *er-103* (Fig. 1C). Furthermore, mature *super1-D er-103* plants are as tall as wild-type plants (Fig. 1G), and elongation of *er-103* pedicels is fully rescued by *super1-D* (Fig. 1H). In addition, *super1-D* conferred reduced fertility (data not shown). These results indicate that overexpression of *SUPER1* by activation tagging suppresses stem and pedicel elongation defects of *er* but also confers additional developmental phenotypes, such as altered leaf shape and reduced fertility.

Genetic Interaction of *er* and *super1-D* Mutants

To understand the nature of genetic interaction, the *super1-D* mutation was subsequently introduced into

Figure 1. *super1-D* is a genetic suppressor of *er-103*. A to C, Inflorescence top view of wild-type (A), *er-103* (B), and *super1-D er-103* (C) plants at bolting. Flowers of wild-type and *super1-D er-103* plants display dramatic pedicel elongation, while the inflorescence tip of *er-103* forms a characteristic flat, disc shape due to reduced internodal and pedicel elongation. Images are taken under the same magnification. Scale bar, 5 mm. D to F, Rosette leaves of 4-week-old wild type (D), *er-103* (E), and *super1-D er-103* (F). Leaves were dissected from the primary shoot axis and laid out in the sequence of development (left, base; right, apex). Images are taken under the same magnification. Scale bar, 1 cm. G and H, Morphometric analysis of full-grown 7-week-old wild-type, *er-103*, and *super1-D er-103* plants. G, Plant height ($n = 8$). H, Lengths of mature pedicels on the main stem ($n = 35$; seven measurements per stem). Bars represent the mean; error bars = SD.



wild-type *ER* and null *er-105* allele backgrounds. It has been shown that the severity of *er* mutations directly reflects reduction in plant height, pedicel length, and fruit (silique) length in a quantitative manner (Torii et al., 1996, 2003). Wild-type *ER* plants develop the longest stems, pedicels, and siliques; null *er-105* plants develop the shortest; and *er-103* plants are intermediate (Fig. 2). As shown in Figure 2, A and B, the *super1-D* mutation conferred extra elongation of stems and pedicels both in *ER* and *er-105* plants, suggesting the largely additive nature of interaction. Interestingly, the degree of stem and pedicel elongation was greater in *er* mutants than in a wild type (For stem length, 17%, 50%, and 43% increase in *ER*, *er-103*, and *er-105*, respectively; for pedicel length, 19%, 32%, and 41% increase in *ER*, *er-103*, and *er-105*, respectively.) Thus, *er* mutants may be more sensitive to the organ elongation effects of *super1-D* (see "Discussion").

In contrast to stems and pedicels, *er* and *super1-D* exhibited distinct interaction in fruit development. While the wild-type *ER* allele suppressed the fertility defects of *super1-D er-103*, the null *er-105* allele enhanced the fertility defect. Notably, *super1-D er-105* plants produced short siliques that contain very few seeds (Fig. 2C and data not shown). Thus, a functional *ER* pathway is required for proper fruit development in the *super1-D* background. From these findings, we conclude that *SUPER1* acts in pathways independent from *ER*, with possible interplay during the development of specific organs, such as the silique.

***SUPER1* Encodes a YUCCA-Like Putative Flavin Monooxygenase**

To gain insights into the molecular nature of the *super1-D* phenotype, we cloned the genomic DNA

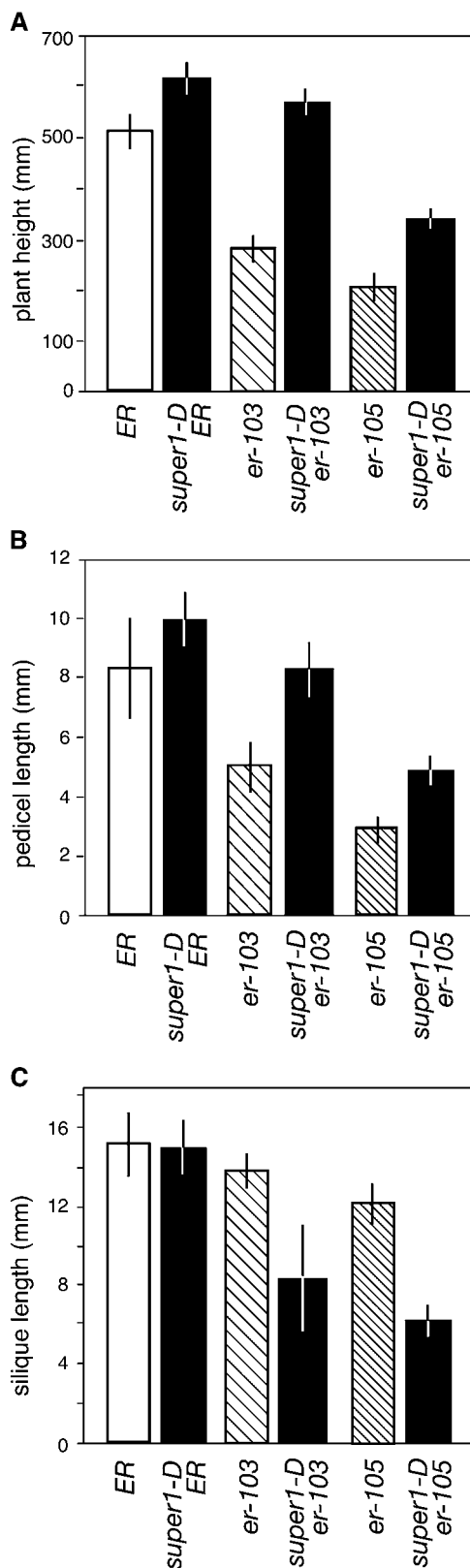


Figure 2. Genetic interaction of *super1-D* with different *er* alleles. **A**, Effects of *super1-D* mutation on final height of wild-type *ER*, intermediate *er-103*, and null *er-105* alleles. $n = 10$ per genotype, error bars = sd . **B**, Effects of *super1-D* mutation on pedicel length of wild-type *ER*, intermediate *er-103*, and null *er-105* alleles. $n = 40$ per genotype, error

fragment flanking the right border of the T-DNA insertion (Fig. 3A). Sequencing analysis revealed that the CaMV35S 4 \times enhancer elements were inserted approximately 5.6 kb upstream of an open reading frame on Chromosome V (bacterial artificial chromosome F6B6). This corresponds to At5g43890/BAA98069, which encodes a 424-amino acid putative flavin-containing monooxygenase (Fig. 3A). It belongs to the *YUCCA* family genes implicated in biosynthesis of auxins, with BLAST scores of 1e-127, 1e-162, and 1e-128 to Arabidopsis *YUCCA* (At4g32540), *YUCCA3* (At1g04610), and petunia (*Petunia hybrida*) *FLOOZY* (*FLZ*), respectively (Zhao et al., 2001; Tobeña-Santamaria et al., 2002). A subsequent, reverse transcription (RT)-PCR analysis revealed that *super1-D er-103* inflorescence accumulates transcripts of At5g43890 at a much higher level than *er-103*, strongly suggesting that *SUPER1* encodes this *YUCCA*-like putative flavin monooxygenase (Fig. 3B). Similar to *YUCCA*, the *SUPER1* protein possesses conserved FAD-binding (GAGPSG) and NADH-binding (GCGNSG) motifs (Fig. 3C). The open reading frames of *YUCCA*, *YUCCA2*, and *FLZ* are intercepted by three introns at the same exact positions (Tobeña-Santamaria et al., 2002). In contrast, *SUPER1* does not possess any introns. The gene was renamed *YUCCA5* following the nomenclature of the *YUCCA* family genes.

Overexpression of *YUCCA5* Confers Suppression of the *er-103* Phenotype

To gain direct evidence that the suppression of the *er-103* phenotype is due to *YUCCA5* overexpression, the full-length cDNA of *YUCCA5* was placed under the constitutive *CaMV35S* promoter and transformed into *er-103* plants. As shown in Figure 4, the transgenic plants recapitulated the suppressed inflorescence phenotype of *er-103*. Furthermore, additional phenotypes, such as narrow leaf blades and reduced fertility, were also recapitulated (data not shown). From these results, we conclude that elevated expression of *YUCCA5*, a new member of the *YUCCA* family of putative flavin monooxygenases, confers all aspects of the *super1-D* phenotype.

YUCCA5 Is Normally Expressed in Roots and Young Vegetative Tissues

We next investigated the expression levels of *YUCCA5* in different organs by RT-PCR. *YUCCA5* was predominantly expressed in roots and young vegetative tissues, while it was under detectable levels in reproductive tissues, such as inflorescence stems, pedicels, and floral buds (Fig. 5). Our result was consistent with the expression profiles of *YUCCA5* obtained using the Arabidopsis full genome chip array (Affymetrix ATH1: analysis by AtGenExpress [[bars = \$sd\$. **C**, Effects of *super1-D* mutation on silique length of wild-type *ER*, intermediate *er-103*, and null *er-105* alleles. \$n = 40\$ per genotype, error bars = \$sd\$.](http://</p>
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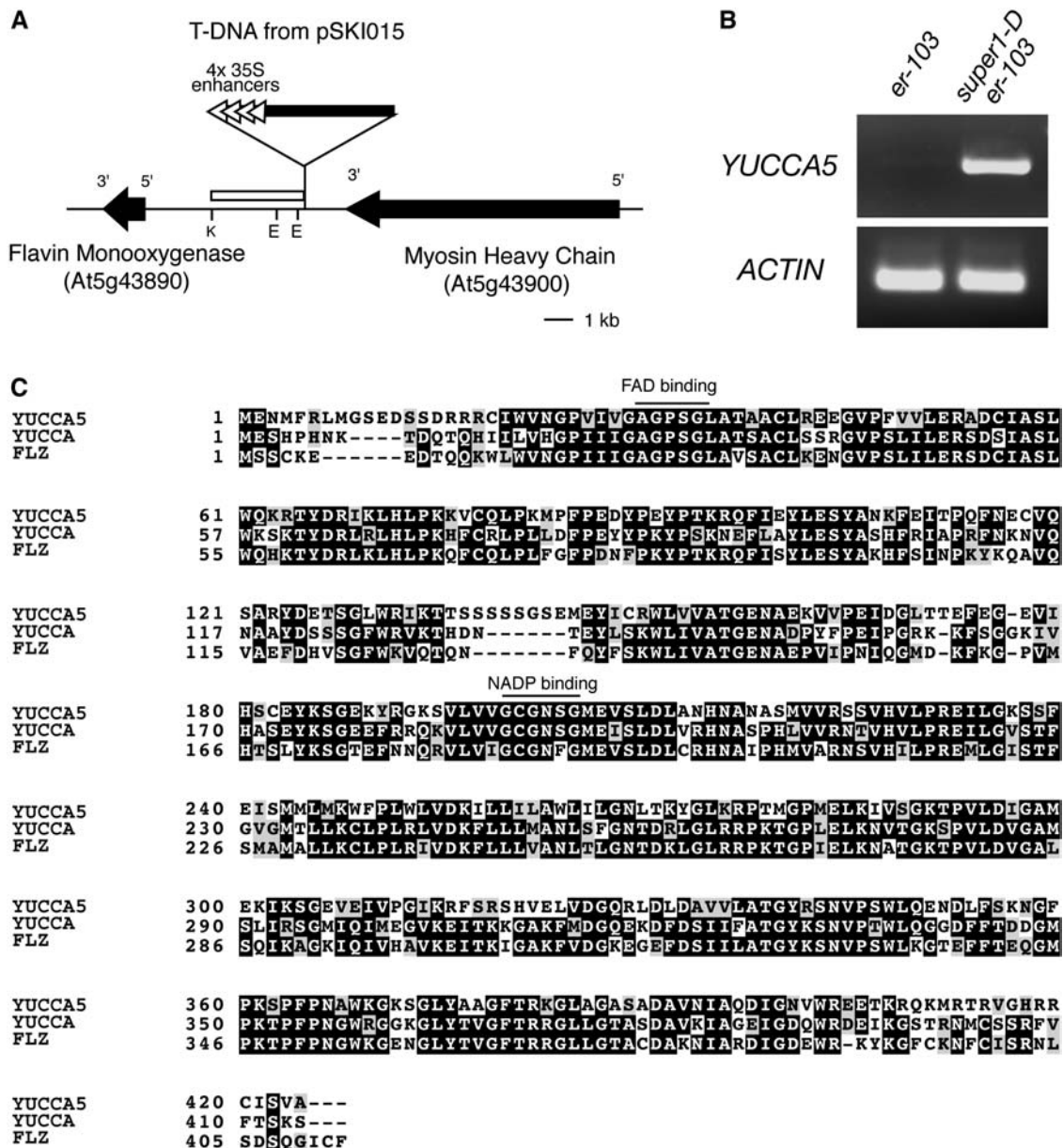


Figure 3. *SUPER1* encodes YUCCA5, a previously undescribed member of YUCCA family putative flavin monooxygenases. A, Genomic organization at the T-DNA insertion site. The black arrows indicate open reading frames in the direction of 5' to 3'. The orientations of 4 × 35S enhancers are indicated by white triangles. The white bar indicates a flanking sequence recovered by inverse-PCR. K, *Kpn*I; E, *Eco*R I. B, Semiquantitative RT-PCR analysis shows that YUCCA5 transcript levels are elevated in *super1-D er-103* inflorescence. The actin fragment was amplified simultaneously as a control. C, Amino acid sequence alignment of YUCCA5 with YUCCA and FLZ, two well-studied members of the YUCCA family of putative flavin monooxygenases. The conserved FAD- and NADP-binding motifs are underlined.

web.uni-frankfurt.de/fb15/botanik/mcb/AFGN/atgenex.htm]; virtual northern analysis by Expression Angler [http://bbc.botany.utoronto.ca/ntools/cgi-bin/ntools_expression_angler.cgi].

Overexpression of YUCCA5 by *super1-D* Results in Elevated Auxin Level and Response

It has been shown that elevated expression of YUCCA, YUCCA3, and FLZ results in overproduction

of auxin, and that recombinant YUCCA protein can catalyze the rate-limiting step in Trp-dependent auxin biosynthesis (Zhao et al., 2001; Tobeña-Santamaria et al., 2002). To test whether YUCCA5 plays a role in auxin biosynthesis, we measured the amounts of free indole acetic acid (IAA) in seedlings of wild type, *er-103*, and *super1-D er-103*. As shown in Figure 6A, the *super1-D* mutation led to a roughly 70% increase in free IAA amount in germinating seedlings. On the other hand, *er-103* seedlings had normal amounts of free



Figure 4. Expression of *YUCCA5* cDNA driven by the dual CaMV 35S promoter recapitulated the *er-103* suppression phenotype. Shown are inflorescences of 5-week-old wild-type (A), *er-103* (B), and transgenic *er-103* (C) plants expressing *CaMV35S::YUCCA5*. Images are taken under the same magnification. Double arrow bars indicate the internode length between the inflorescence tip and the node-bearing flower turning into stage 17. Scale bar, 5 mm.

IAA (Fig. 6A), suggesting that growth defects by *er-103* mutation are not caused by the reduction in free auxin levels.

To understand the relationship between elevated auxin levels and the suppressed *er-103* inflorescence phenotype by *super1-D*, we sought to visualize cellular auxin distribution and response using genetic tools. For this purpose, we used a reporter construct that expresses the β -glucuronidase (GUS) gene under the 0.6-kb promoter of auxin-responsive gene *AtAux2-11/IAA4* (Wyatt et al., 1993). This promoter can respond to a range of auxin concentration from 10^{-8} to 10^{-4} M (Wyatt et al., 1993). The well-known reporter construct, *DR5::GUS* (Ulmasov et al., 1997), did not drive strong GUS expression in the inflorescence and floral tissues; thus, it was not used in this study (data not shown).

The *super1-D* mutation led to increased *AtAux2-11::GUS* activity both in seedlings and inflorescences. In seedlings, the elongation zone of hypocotyls and petioles, as well as developing leaves, showed extensive staining (Fig. 6B). Fully expanded cotyledons also displayed increased GUS activity. At the reproductive stage, inflorescence stems at the tip and elongating pedicels exhibited strong staining (Fig. 6B). GUS activity diminished in older pedicels and stems that are no longer elongating (Fig. 6B; data not shown). These results demonstrate that overexpression of *YUCCA5* by *super1-D* leads to elevated levels of free auxin and, importantly, that increased auxin distribution and response in elongating regions of internodes and pedicels likely cause the suppressed *er-103* phenotype.

er Mutation Does Not Significantly Alter Auxin Response

While *super1-D* dramatically suppressed the elongation defects of *er-103*, the wild-type plants with additional *super1-D* mutation exhibited only a modest increase in inflorescence and pedicel elongation (Fig. 2). This is not due to reduction in *YUCCA5* overexpression by outcrossing into wild-type plants, as *YUCCA5* transcripts are equally up-regulated in the

wild-type *ER* background (data not shown). One possibility is that *er* mutant plants may be defective in responding to normal levels of endogenous auxin to promote organ growth, but they can respond to increased auxin levels resulting from *YUCCA5* overexpression. To test this hypothesis, we analyzed induction kinetics of *Aux2-11::GUS* by exogenous IAA in wild-type and *er-103* plants. A 6-h incubation with IAA resulted in increased GUS activity both in wild-type and *er-103* seedlings carrying *Aux2-11::GUS* (Fig. 7). While the GUS induction was most striking in the root tips, entire rosettes displayed some increase in the GUS staining (Fig. 7A and data not shown). However, we did not detect any clear difference in GUS induction levels between wild-type and *er-103* seedlings, either by quantitative GUS activity assay or by GUS tissue staining. These results indicate that the *er* mutation does not significantly alter auxin response and further imply that suppression of the *er* phenotype by *YUCCA5* involves interaction at downstream processes, such as cell proliferation and elongation.

super1-D Is Epistatic to *er* during Seedling Development and Confers Hypocotyl Cell Elongation

The activation-tagged allele of *YUCCA* was originally isolated from its seedling-long hypocotyls and epinastic cotyledons, both of which are characteristics of auxin overproduction (Zhao et al., 2001). We next tested whether *super1-D* also confers seedling auxin overproduction phenotypes. For this purpose, the phenotypic severity was compared between *super1-D* and a recapitulated *yucca* line, which drives *YUCCA* cDNA under the *CaMV35S* promoter (Zhao et al., 2001). The original *yucca-D* plants are nearly sterile (Zhao et al., 2001) and thus were not used for the analysis. The *super1-D* mutation indeed conferred elongated hypocotyls (Figs. 5B and 6A). However, hypocotyl length of *super1-D* was shorter than that of *yucca* recapitulated lines (Student's *t* test; *t* value = 4.143, *P* = 0.0003). Unlike *yucca* recapitulated lines, *super1-D* seedlings did not exhibit severe cotyledon epinasty, suggesting that the effects conferred by activation-tagged *YUCCA5* are rather moderate (Fig. 8A).

We next analyzed the genetic interaction of *er* and *super1-D* during seedling development. We found that the null *er-105* mutation resulted in a very subtle yet

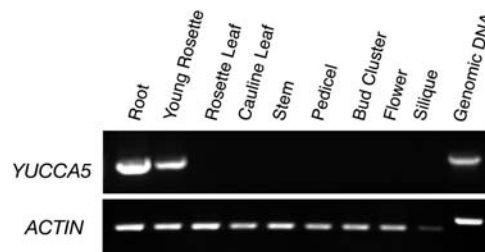


Figure 5. Expression of *YUCCA5* in different organs of wild-type Arabidopsis plants. The actin cDNA fragment was amplified as a control for 26 cycles. *YUCCA5* cDNA was amplified for 45 cycles.

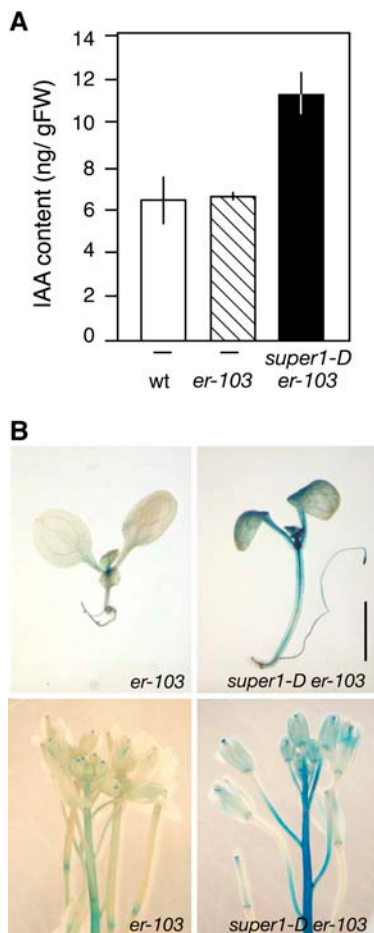


Figure 6. Activation-tagged *YUCCA5* by *super1-D* mutation confers elevated levels of auxin and increased auxin distribution and response in elongating tissues. **A**, Free IAA content (nanogram/gram fresh weight) in seedlings of wild type, *er-103*, and *super1-D er-103*. Error bars = *sd*. **B**, The reporter construct *AtAux2-11::GUS* visualizing auxin distribution and response in 6-d-old seedlings (top) and 5-week-old inflorescence tips (bottom) of *er-103* (left) and *super1-D er-103* (right), respectively. Scale bar, 2 mm.

statistically significant reduction in hypocotyl length ($P = 0.0013$). Overexpression of *YUCCA5* by the *super1-D* mutation led to elongated hypocotyls regardless of the presence or absence of *er* mutations (Fig. 8, A and B). Thus, with respect to the long hypocotyl phenotype, *super1-D* appears epistatic to *er*. The cellular basis of *super1-D*-mediated hypocotyl elongation was subsequently investigated. As shown in Figure 8C, both epidermal and cortex cell files show significant elongation under the *super1-D* background. The results indicate that *super1-D* confers a long hypocotyl phenotype by increasing cell elongation.

Cellular Phenotype of Mature Pedicels

Unlike hypocotyl elongation, *er* and *super1-D* exhibited largely additive effects as well as complex interactions during internode and pedicel elongation, and this led to a complete rescue of *er-103* inflores-

cence architecture (Figs. 1 and 2). To understand the cellular basis of the phenotypic rescue, we next examined the cellular phenotype of mature pedicels. It has been shown that the short pedicel phenotype of *er-105* is due primarily to a reduced number of cortex cells, which are abnormally expanded (Shpak et al., 2003). As shown in Figure 9, the severity of the *er* mutation correlated with an increase in the size of both the epidermal pavement and cortex cells. Interestingly, the effects of *super1-D* for these two cell types were completely distinct. The *super1-D* mutation led to elongation of the pedicel epidermal pavement cells of wild-type *ER*, *er-103*, and *er-105* ($P = 0.0058$, $P = 0.0005$, and $P = 0.042$ for the comparison of *ER* and *super1-D ER*, *er-103* and *super1-D er-103*, and *er-105* and *super1-D er-105*, respectively; Fig. 9G). The extent of increased epidermal cell elongation of *er* mutants by *super1-D* roughly correlated with the increased pedicel length (Figs. 2B and 9G). By contrast, the presence of *super1-D* had no effects on cortex cell size ($P = 0.48$, $P = 0.66$, and $P = 0.79$ for the comparison of *ER* and *super1-D ER*, *er-103* and *super1-D er-103*, and *er-105* and *super1-D er-105*, respectively; Fig. 9H). The results suggest that *super1-D* suppresses pedicel elongation defects of *er* by promoting elongation of the epidermal

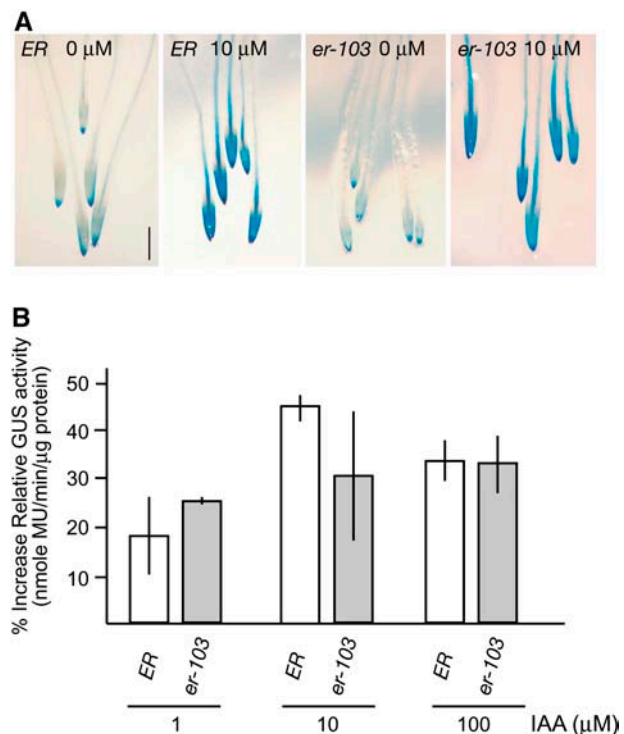


Figure 7. Induction of GUS activity in wild-type and *er-103* seedlings carrying *AtAux2-11::GUS* reporter construct. **A**, Histochemical staining of GUS activity in the root tip of wild-type *ER* and *er-103* after a 6-h treatment with 10 μM IAA. Images are taken at the same magnification. Scale bar, 0.5 mm. **B**, Quantitative GUS fluorimetric activity assays of wild-type *ER* and *er-103* seedlings treated with 1, 10, and 100 μM IAA for 6 h. Bar graphs represent the mean of two independent induction experiments. For each induction, the enzymatic reactions were repeated three times per genotype. Bars = *se*.

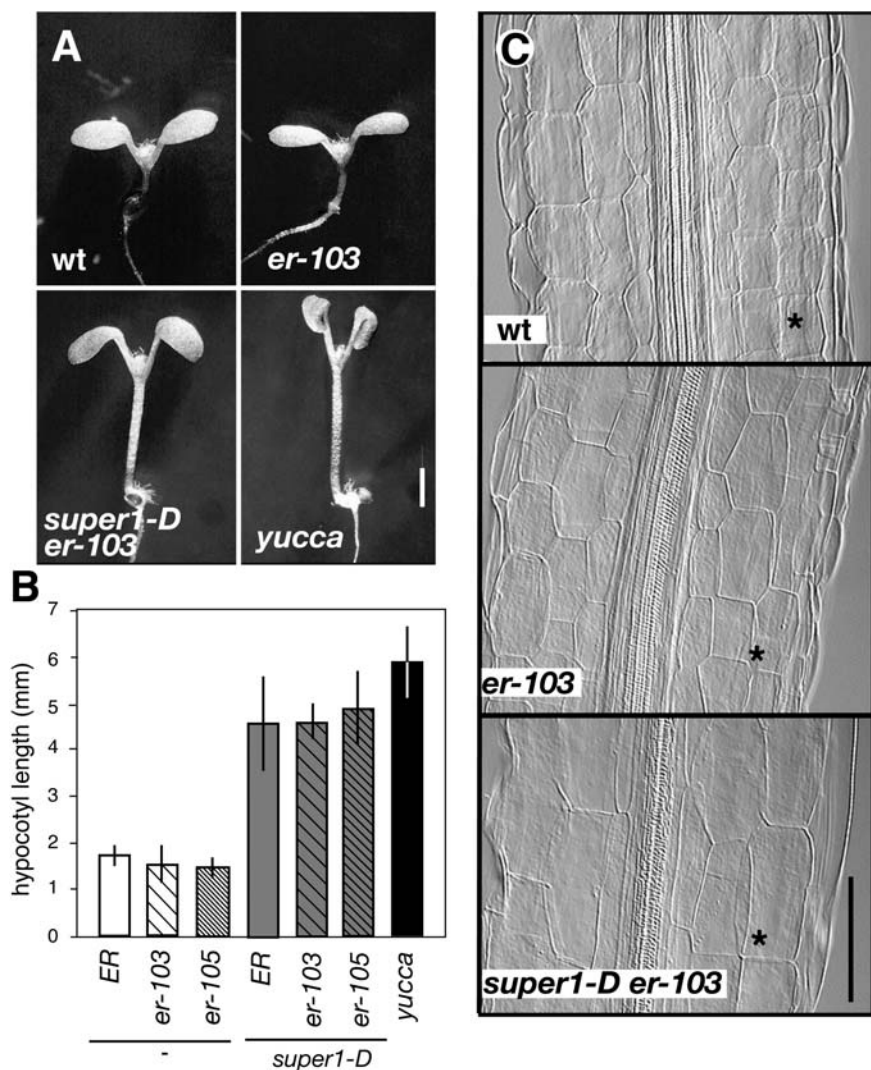


Figure 8. The seedling phenotype of *super1-D* is epistatic to *er*, and *super1-D* confers hypocotyl cell elongation. A, Six-day-old seedlings of wild type (wt: top left), *er-103* (top right), *super1-D er-103* (bottom left), and recapitulated *yucca* line (bottom right). Scale bar, 2 mm. B, Hypocotyl length of the wild-type *ER*, intermediate *er-103*, null *er-105* seedlings with and without the presence of *super1-D* mutation. Hypocotyl length of the recapitulated *yucca* line was plotted as a comparison. $n = 16$, error bars = SD. C, Cellular morphology of hypocotyls of 6-d-old wild type (wt: top), *er-103* (middle), and *super1-D er-103* (bottom) seedlings. The cortex tissue is indicated by asterisks. Images are taken at the same magnification. Scale bar, 100 μm .

pavement cells, which is accompanied by an increase in cortex cell numbers.

DISCUSSION

In this study, we isolated a dominant suppressor of *er* by activation tagging and further identified that suppression of the *er* inflorescence phenotype was due to overexpression of *YUCCA5*, a previously undescribed member of the *YUCCA* family of putative flavin monooxygenases. Our findings highlight an intersection of auxin and ER-RLK signaling that is important for plant growth and elaboration of inflorescence architecture.

Activation Tagging of *YUCCA5* Confers Elevated Auxin Levels and Elongated Inflorescence

Among 10 *Arabidopsis* *YUCCA* family members, *SUPER1/YUCCA5* belongs to a subfamily with two

additional members, At1g04180 (AAC16744) and BAS3/At4g28720 (CAA22980). These subfamily members possess either no intron (*YUCCA5* and *BAS3*) or one intron (At1g04180), as opposed to three introns in conserved locations in orthotypical *YUCCA* members (e.g. *YUCCA2*, *THREAD/YUCCA4*, and petunia *FLZ*; Tobeña-Santamaria et al., 2002). Our finding, that overexpression of *YUCCA5* by *super1-D* mutation conferred increased free IAA levels, extends the role of this *YUCCA* subfamily for auxin biosynthesis. In contrast, activation tagging of *BAS3*, the closest homolog of *YUCCA5*, had been reported to confer short hypocotyls, a phenotype opposite to *super1-D* (Zhao et al., 2001).

Although their IAA contents have not been quantified, the activation tagging/overexpression of other *YUCCA* members, namely *YUCCA2*, *YUCCA3*, and *THREAD/YUCCA4* also conferred a signature phenotype of *yucca*: elongated hypocotyls, epinastic leaves, severely weak and lanky inflorescences, increased apical dominance, and reduced fertility (Zhao et al.,

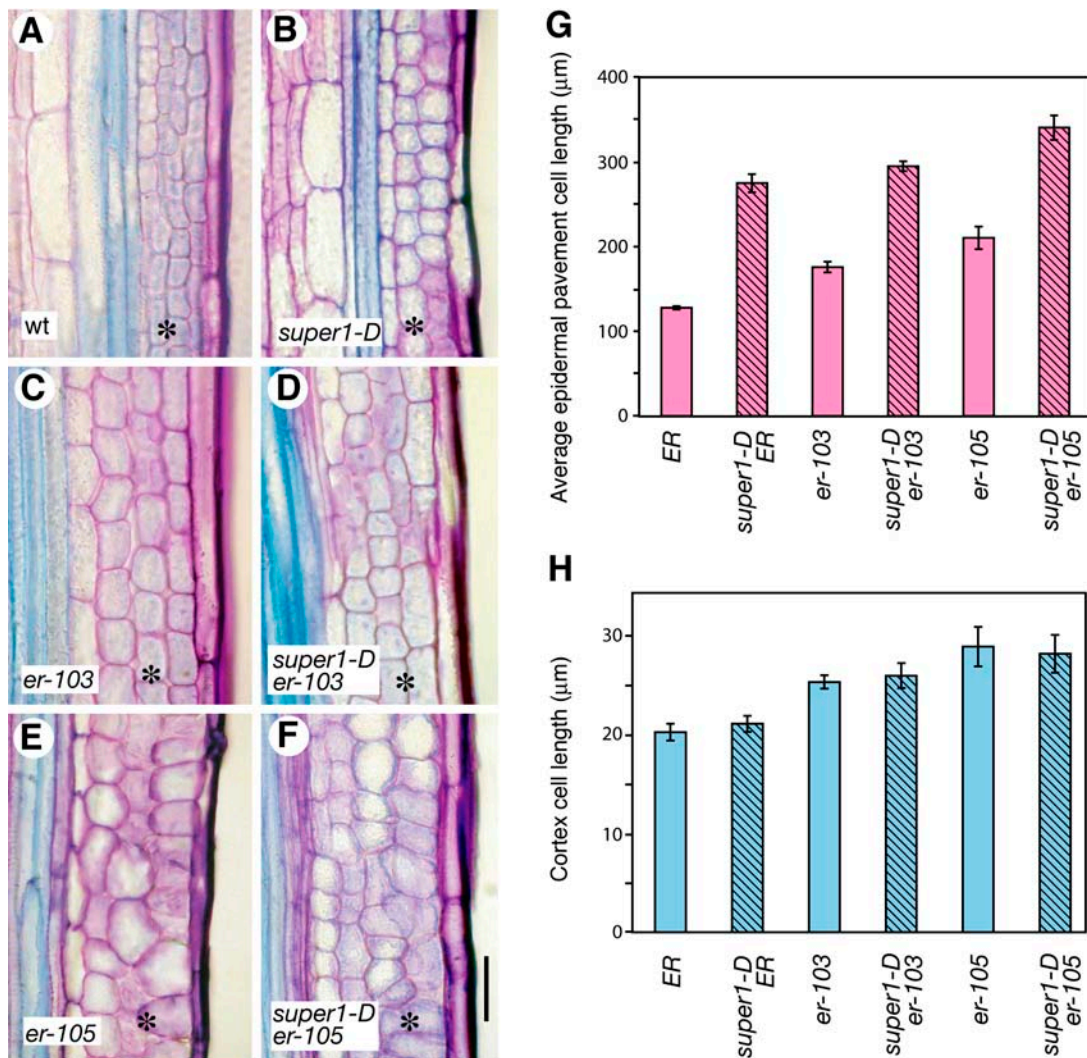


Figure 9. Cellular basis of *super1-D*-mediated pedicel elongation. A to F, Longitudinal sections of mature pedicels from wild type (wt; A), *super1-D* (B), *er-103* (C), *super1-D er-103* (D), *er-105* (E), and *super1-D er-105* (F). The *er* mutations led to a reduced number of enlarged cortex cells (asterisks), while *super1-D* mutation promotes pedicel elongation without increasing the cortex cell size. Bars = 50 μm. G, Average epidermal pavement cell length. For each genotype, pedicel epidermal strips were prepared from three individual plants. Fifteen pavement cells were measured from each epidermal strip. Bars = SE. H, Cortex cell length. Bar represent the average ($n = 20$ for each genotype). Bars = SE.

2001; Marsch-Martinez et al., 2002). In contrast, the phenotype of *super1-D* was notably milder. For instance, the *super1-D* hypocotyls were shorter than that of *yucca-D*, and the *super1-D* cotyledons did not exhibit strong epinasty. Similarly, the adult phenotypes of *super1-D* in the presence of functional *ER* are vigorous and fertile with larger rosettes and inflorescences.

The difference in the growth phenotype between *super1-D* and other *YUCCA* overexpressors may reflect redundant but distinct roles of each *YUCCA* member during plant growth and development. One possibility is that the activation-tagged *YUCCA5* leads to a relatively moderate increase in free IAA amounts in the inflorescence. This may be because the *YUCCA5* enzyme is not as active as the other members. Alternatively, the *super1-D* mutation may not drive high ex-

pression of *YUCCA5* as compared to other activation-tagged *YUCCA* genes. In wild-type plants, *YUCCA5* is predominantly expressed in young vegetative tissues and roots (Fig. 5). Further analysis to correlate tissue-specific accumulation of activation-tagged transcripts and IAA among different *yucca* family mutants will be necessary to clarify these two possibilities. The recently developed technique of microscale IAA quantification in tissue sections (Mori et al., 2005) may enable direct comparison of tissue-specific auxin accumulation to the developmental phenotypes.

It has been reported that loss-of-function mutations in *YUCCA* and *YUCCA2* confer no visible phenotype, most likely due to redundancy (Zhao et al., 2001). Therefore, a targeted disruption of multiple *YUCCA*-family genes including *YUCCA5* by generating higher-

order mutant/cosuppression lines may be required to further address the developmental roles of *YUCCA*-family genes and their interaction with *ER*.

Intersection of ER- and Auxin-Mediated Control of Inflorescence Architecture

While our screen did not identify novel ER-signaling components, it highlighted the intersection of ER- and auxin-mediated organ growth and internodal elongation, which in turn translates to inflorescence architecture. Previous studies have shown that both ER and auxin are required for elongation of the inflorescence and that they display synergism. For instance, a large-scale genetic screen for clustered, *er*-like inflorescence phenotype led to the isolation of 10 novel *er-like1* mutants that are allelic to *tir3* (Lease et al., 2001). Conversely, *er* was identified as an enhancer of *attenuated shade avoidance1*, which is also allelic to *tir3* (Kanyuka et al., 2003). *tir3* was originally isolated as a mutant resistant to the auxin polar transport inhibitors (Ruegger et al., 1997). *TIR3* encodes a calossin-like protein BIG that is required for normal auxin distribution, presumably by affecting the cellular localization of the PIN family auxin efflux carriers (Gil et al., 2001). The complete suppression of the compact inflorescence phenotype of *er-103* by activation tagging of *YUCCA5* provides supporting evidence for complementary action of ER and auxin in promoting inflorescence elongation.

The effects of *super1-D* in promoting elongation of inflorescences and pedicels, as well as reducing fertility, are more pronounced in *er* plants than in wild-type plants (see Fig. 2), implying that *er* plants may be more sensitive to elevated levels of auxin. The mechanism by which *er* mutation confers elevated response to auxin-mediated growth is not clear. It is highly unlikely that ER directly affects auxin biosynthesis or homeostasis for the following reasons: First, *er* mutations do not significantly alter the endogenous levels of free IAA (Fig. 4). Second, the *er* mutations do not affect expression of auxin-responsive promoter activity of *AtAux2-11::GUS* during vegetative and inflorescence development under normal growth condition (data not shown; Wyatt et al., 1993; Abel et al., 1994; Ulmasov et al., 1997). Third, both wild-type and *er-103* mutant plants responded similarly to the IAA application (Fig. 7), suggesting that the *er* mutations do not confer reduced sensitivity to the auxins. Finally, it has been reported that a single recessive mutation in ecotype Landsberg *er* confers inflorescence stem segments a responsiveness to IAA-stimulated cell elongation via activation of the plasma membrane proton pump, so-called acid growth (Soga et al., 2000). We repeated their experiments using *er* alleles in Landsburg as well as Columbia ecotypes, and ruled out the possibility that the *er* mutation is responsible for increased acid growth (S. Bemis and K. Torii, unpublished data).

The *super1-D* mutation conferred elongation of the epidermal pavement cells of both wild-type and *er* mutant pedicels to a similar extent (Fig. 9), while

elongation of wild-type pedicels was not nearly as dramatic as *er* mutant pedicels (Fig. 2). This implies that it is not simply the extent of the epidermal cell elongation that specifies the overall elongation of pedicels as a whole. Unlike the epidermis, the cortex cell size was not altered by the *super1-D* mutation (Fig. 9). Therefore the sensitivity of *er* to *super1-D* may involve auxin-mediated cell proliferation.

Perhaps both auxin and ER promote inflorescence growth via largely independent pathways but with overlaps in their ultimate downstream processes (e.g. cell proliferation). Thus the excessive activity of either pathway may partly mask the loss of the other. Plant growth is under the control of both environmental stimuli and developmental programs. Multiple environmental and endogenous pathways may overlap or be redundant. Recently, the cross talk of hormone-signaling pathways has emerged as a central theme in plant growth regulation, as exemplified by the recent finding that interaction of auxin and brassinosteroids in promoting seedling growth occurs via transcriptional control of downstream targets (Goda et al., 2004; Nemhauser et al., 2004). Further studies, such as a genome-wide comparison of gene expression by auxin- and ER-signaling pathways may reveal the molecular basis of their intersections during growth and development of inflorescence.

MATERIALS AND METHODS

Plant Materials and Growth Conditions

Arabidopsis (*Arabidopsis thaliana* ecotype Col) was used as the wild type. The *yucca* recapitulated line is a generous gift from Dr. Yunde Zhao (University of California at San Diego, La Jolla, CA) and Dr. Joanne Chory (Salk Institute, La Jolla, CA), and the *AtAux2-11::GUS* line is a generous gift from Dr. Masao Tasaka (NAIST, Ikoma, Japan). *er-103*, *er-105*, *yucca*, and *AtAux2-11::GUS* lines are all in the Col background. Plants were grown on soil mixtures (Sunshine Mix4:Vermiculite:Perlite = 2:1:1 with 0.85 mg/cm³ of Osmocot14-14-14) under 18/6 light-dark cycle at 21°C. For plate cultures, seeds were surface sterilized with 30% bleach and 0.01% Triton X-100 (Sigma) for 12 min and subsequently washed five times with sterile distilled water. Seedlings were grown in Murashige and Skoog media supplemented with 1% (w/v) Suc.

Genotyping

The *er-105* mutation was detected by using a primer pair ERg2248 (AAG AAG TCA TCT AAA GAT GTG A) and *er-105rc* (AGC TGA CTA TAC CCG ATA CTG A), and the absence of the wild-type allele was confirmed by a primer pair ERg2248 and ERg3016rc (AGA ATT TTC AGG TTT GGA ATC TGT). *er-103* plants were genotyped using a primer pair ERg3588 (GAC TTG TCC TAC AAT CAG CTA ACT) and ERg4056rc (TAG ACC AGT CAG CTT GTT ACT GTG). The amplified fragment was subsequently digested with *Nla*III to detect 3 cleaved amplified polymorphic sequences (CAPS): Bands obtained for wild type are 209 bp, 150 bp, 70 bp, and 65 bp; bands for *er-103* are 274 bp, 150 bp, and 70 bp.

Isolation of *super1-D* Mutant and Recovery of the T-DNA Flanking Sequence

er-103 plants were vacuum infiltrated with the vector pSKI015 (a generous gift from Dr. Detlef Weigel; Weigel et al., 2000). *super1-D* was isolated as a suppressor from batch 38. An analysis of the subsequent generation revealed a segregation of Basta resistance to Basta-sensitive approximately 2:1 (sup-

pressed phenotype: *er-103* phenotype = 43:21; BastaR:BastaS = 43:21; all suppressed plants were BastaR and all *er-103* phenotype plants were BastaS), indicating that it is activation tagged and presumably homozygous lethal. This was further confirmed by analysis of the F3 populations, where none of the populations derived from 15 independent *super1-D* plants carried homozygous Basta resistance. Therefore, morphological analysis was performed using the *super1-D/+* plants.

The genomic DNA was isolated from 2-week-old *super1-D* plants using Nucleon Phytopure plant DNA extraction kit (Amersham Pharmacia LKB). The T-DNA flanking sequence of *super1-D* was recovered by the inverse-PCR (iPCR) as described by Li et al. (2001), except that both *EcoRI* and *KpnI* were used to digest the *super1-D* genomic DNA. The amplified fragments were cloned into pCR2.1-TOPO vector (Invitrogen) and subjected to sequencing.

RNA and RT-PCR Analysis

Total RNA was prepared by using an RNeasy mini kit (Qiagen) and subsequently treated with DNaseI (Invitrogen) prior to the RT-PCR reaction. cDNA was synthesized from 0.2 mg of the total RNA by using Thermoscript reverse transcriptase (Invitrogen) and a random hexamer as a primer. One-fifth of the synthesized cDNA was used for the PCR reaction with a *YUCCA5* gene-specific primer pair: May2FMO5 (cggaattccc ATG GAG AAC ATG TTT AGG CT) and May2FMO3rc (ccggatcc CTA AGC AAC TGA AAT GCA TC). The Arabidopsis actin (*ACT2*) gene was amplified as a template control using a primer pair: ACT2-1 (GCC ATC CAA GCT GTT CTC TC) and ACT2-2 (GCT CGT AGT CAA CAG CAA CAA).

Cloning of the *YUCCA5* Coding Sequence and Generation of Transgenic Plants

The *YUCCA5* cDNA was cloned into the *EcoRI* and *BamHI* sites of pSP73 (Promega) and the sequence was confirmed. The plasmid was partially digested with *NcoI* and *BamHI* and inserted into pRTL2-GUS to replace the *GUS* gene. This construct, which drives the *YUCCA5* expression under a dual-enhanced CaMV35S promoter, was cloned into the *HindIII* site of pPZP222 (Hajdukiewics et al., 1994) to generate pEJH101. The plasmid was introduced into *Agrobacterium* strain GV3101 by electroporation and transformed into Arabidopsis *er-103* plants by vacuum infiltration. Selection of the transgenic plants and segregation analysis in the next generation were conducted as described by Shpak et al. (2003).

Histological Analysis, Microscopy, and Cell Length Measurement

Fixation, embedding, and sectioning of tissues for light microscopy using Olympus BX40 were performed as described by Shpak et al. (2003). The GUS staining was performed according to Sessions et al. (1999). For clearing of hypocotyls and pedicels, tissues were placed in 9:1 ethanol:acetic acid solution at room temperature overnight, rehydrated through an ethanol series (85%, 70%, 50%, 30%, 15%) and then stored in chloral hydrate:water:glycerol 8:1:1. The cleared specimens were viewed under the differential interference contrast microscope Nikon Optishot equipped with a Sony DXC-960MD CCD video camera. The cortex cell length was measured directly on the photographic images of plastic sections. For measuring the epidermal pavement cell length, the pedicel epidermis was stripped with scapel, photographed under phase contrast microscopy, and analyzed with Adobe Photoshop 7.

Auxin Treatment and GUS Activity Assay

Two-week-old seedlings were transferred from plates to full-strength Murashige and Skoog liquid media and incubated for 5 d with shaking (100 rpm) at 18-h-light/6-h-dark condition. Seedlings were then treated with 0, 1, 10, or 100 μM IAA (Sigma) for 6 h with shaking. Subsequently, 150 to 200 mg of plant tissue was ground in liquid nitrogen and homogenized in 500 μL extraction buffer containing 50 mM sodium phosphate (pH 7.0), 10 mM EDTA, 0.1% sodium laurylsarcosine, 0.1% Triton X-100, and 10 mM β -mercapto-ethanol. The extracts were cleared by centrifugation at 12,000g at 4°C for 5 min. The protein content of the extracts was determined using a Bio-Rad Protein Assay kit according to the manufacturer's instructions. To measure GUS activity, 3 μL protein extract was mixed with 475 μL 2 mM 4-methylumbelliferyl β -D-glucuronide (in extraction buffer), and incubated at 37°C for

45 min. The reaction was stopped by mixing 50 μL of the mixture with 200 μL 0.2 M sodium carbonate. Fluorescence of the samples was measured at 360 (excitation) and 460 (emission) nm using a SynergyHT Multi-Detection microplate reader (BioTek).

Quantification of Free IAA Level

For gas chromatography, single-ion-monitoring mass spectrometry (GC-SIM-MS) analyses, fresh plant material (corresponding to a pool of 10 seedlings grown on agar plates for 7 d under continuous light) was carefully weighed, frozen in liquid nitrogen, and stored at -80°C . For extraction of free IAA, the material was ground in liquid nitrogen using a mortar and pestle. After addition of 0.6 ng of [^{13}C]IAA (Cambridge Isotope Lab) as an internal standard, the material was extracted in 80% acetone and 0.1 mg/mL 2,6-Di-tert-butyl-4-methylphenol for 60 min. After centrifugation, the supernatant was collected. The pellet was reextracted again for 90 min, and the supernatant was brought to a water phase in a rotary evaporator. IAA was partially purified from the residual aqueous solution by partitioning using ether, and then purified by HPLC connecting with a fluorometer (Hitachi FL Detector L-7485; 280 nm excitation, 355 nm emission). The HPLC was performed using a Nucleosil N(CH₃)₂ column (Senshu) with an isocratic system of 100% methanol and 0.03% acetic acid. The purified IAA fraction was dried under a nitrogen stream and trimethylsilylated with *N*-methyl-*N*-trimethylsilyltri-fluoroacetamide at 60°C for 15 min.

Splitless injections were made into a GC-single-ion-monitoring-MS system (QP5050A, Shimadzu), equipped with a capillary column (DB-1, 0.25 mm i.d. \times 30 m, film thickness 0.25 μm ; J&W Scientific). A linear temperature gradient was applied from 80°C to 280°C with an increase of 20°C/min. The injection temperature of the GC was 250°C, the ion source temperature of the MS was 250°C, and a helium flow of 1.2 mL/min was applied. The ionization potential was 70 electron volt, and the scan time was 0.2 s. The percentages of molecules of IAA labeled with ^{13}C were calculated from the relative intensities of *m/z* 202 to 208 and 319 to 325 ions after subtraction of background.

Upon request, all novel materials described in this publication will be made available in a timely manner for noncommercial research purposes, subject to the requisite permission from any third-party owners of all or parts of the material. Obtaining any permissions will be the responsibility of the requestor.

Sequence data from this article can be found in the GenBank/EMBL data libraries under accession number DQ159070.

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LITERATURE CITED

- Abel S, Oeller PW, Theologis A (1994) Early auxin-induced genes encode short-lived nuclear proteins. *Proc Natl Acad Sci USA* **91**: 326–330
- Azpiroz R, Wu Y, LoCascio JC, Feldmann KA (1998) An Arabidopsis brassinosteroid-dependent mutant is blocked in cell elongation. *Plant Cell* **10**: 219–230
- Carles CC, Fletcher JC (2003) Shoot apical meristem maintenance: the art of a dynamic balance. *Trends Plant Sci* **8**: 394–401
- Chrispeels MJ, Sadava DE (1994) Plants, Genes, and Agriculture. Jones and Bartlett, Boston, pp 298–327
- Diévarit A, Clark SE (2004) LRR-containing receptors regulating plant development and defense. *Development* **131**: 251–261

- Doebley J, Stec A, Hubbard L (1997) The evolution of apical dominance in maize. *Nature* **386**: 485–488
- Douglas SJ, Chuck G, Dengler RE, Pelecanda L, Riggs CD (2002) *KNAT1* and *ERECTA* regulate inflorescence architecture in *Arabidopsis*. *Plant Cell* **14**: 547–558
- Feldmann KA, Marks MD, Christianson ML, Quatrone RS (1989) A dwarf mutant of *Arabidopsis* generated by T-DNA insertion. *Science* **243**: 1351–1354
- Fridborg I, Kuusk S, Robertson M, Sundberg E (2001) The *Arabidopsis* protein SHI represses gibberellin responses in *Arabidopsis* and barley. *Plant Physiol* **127**: 937–948
- Gil P, Dewey E, Friml J, Zhao Y, Snowden KC, Putterill J, Palme K, Estelle M, Chory J (2001) BIG: a calossin-like protein required for polar auxin transport in *Arabidopsis*. *Genes Dev* **15**: 1985–1997
- Goda H, Sawa S, Asami T, Fujioka S, Shimada Y, Yoshida S (2004) Comprehensive comparison of auxin-regulated and brassinosteroid-regulated genes in *Arabidopsis*. *Plant Physiol* **134**: 1555–1573
- Hajdukiewicz P, Svab Z, Maliga P (1994) The small, versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol Biol* **25**: 989–994
- Hanzawa Y, Takahashi T, Michael AJ, Burtin D, Long D, Pineiro M, Coupland G, Komeda Y (2000) *ACAULIS5*, an *Arabidopsis* gene required for stem elongation, encodes a spermine synthase. *EMBO J* **19**: 4248–4256
- Hedden P, Proebsting WM (1999) Genetic analysis of gibberellin biosynthesis. *Plant Physiol* **119**: 365–370
- Kakimoto T (1996) CKI1, a histidine kinase homolog implicated in cytokinin signal transduction. *Science* **274**: 982–985
- Kanyuka K, Praekelt U, Franklin KA, Billingham OE, Hooley R, Whitelam GC, Halliday KJ (2003) Mutations in the huge *Arabidopsis* gene BIG affect a range of hormone and light responses. *Plant J* **35**: 57–70
- Komeda Y, Takahashi T, Hanzawa Y (1998) Development of inflorescences in *Arabidopsis thaliana*. *J Plant Res* **111**: 283–288
- Lease KA, Wen J, Li J, Doke JT, Liscum E, Walker JC (2001) A mutant *Arabidopsis* heterotrimeric G-protein beta subunit affects leaf, flower, and fruit development. *Plant Cell* **13**: 2631–2641
- Leyser HM, Lincoln CA, Timpte C, Lammer D, Turner J, Estelle M (1993) *Arabidopsis* auxin-resistance gene *AXR1* encodes a protein related to ubiquitin-activating enzyme E1. *Nature* **364**: 161–164
- Li J, Chory J (1997) A putative leucine-rich repeat receptor kinase involved in brassinosteroid signal transduction. *Cell* **90**: 929–938
- Li J, Lease KA, Tax FE, Walker JC (2001) BRS1, a serine carboxypeptidase, regulates BRI1 signaling in *Arabidopsis thaliana*. *Proc Natl Acad Sci USA* **98**: 5916–5921
- Li J, Nagpal P, Vitart V, McMorris TC, Chory J (1996) A role for brassinosteroids in light-dependent development of *Arabidopsis*. *Science* **272**: 398–401
- Lincoln C, Britton JH, Estelle M (1990) Growth and development of the *axr1* mutants of *Arabidopsis*. *Plant Cell* **2**: 1071–1080
- Marsch-Martinez N, Greco R, Van Arkel G, Herrera-Estrella L, Pereira A (2002) Activation tagging using the En-I maize transposon system in *Arabidopsis*. *Plant Physiol* **129**: 1544–1556
- Matsubayashi Y, Yang H, Sakagami Y (2001) Peptide signals and their receptors in higher plants. *Trends Plant Sci* **6**: 573–577
- Mele G, Ori N, Sato Y, Hake S (2003) The *knotted1-like* homeobox gene *BREVIPEDICELLUS* regulates cell differentiation by modulating metabolic pathways. *Genes Dev* **17**: 2088–2093
- Mori Y, Nishimura T, Koshiba T (2005) Vigorous synthesis of indole-3-acetic acid in the apical very tip leads to a constant basipetal flow of the hormone in maize coleoptile. *Plant Science* **168**: 467–473
- Nemhauser JL, Mockler TC, Chory J (2004) Interdependency of brassinosteroid and auxin signaling in *Arabidopsis*. *PLoS Biol* **2**: E258
- Olszewski N, Sun TP, Gubler F (2002) Gibberellin signaling: biosynthesis, catabolism, and response pathways. *Plant Cell Suppl* **14**: S61–S80
- Peng J, Carol P, Richards DE, King KE, Cowling RJ, Murphy GP, Harberd NP (1997) The *Arabidopsis* GAI gene defines a signaling pathway that negatively regulates gibberellin responses. *Genes Dev* **11**: 3194–3205
- Peng J, Richards DE, Hartley NM, Murphy GP, Devos KM, Flintham JE, Beales J, Fish LJ, Worland AJ, Pelica F, et al (1999) 'Green revolution' genes encode mutant gibberellin response modulators. *Nature* **400**: 256–261
- Ruegger M, Dewey E, Hobbie L, Brown D, Bernasconi P, Turner J, Muday G, Estelle M (1997) Reduced naphthylphthalamic acid binding in the tir3 mutant of *Arabidopsis* is associated with a reduction in polar auxin transport and diverse morphological defects. *Plant Cell* **9**: 745–757
- Ryan C, Pearce G, Scheer JM, Moura DS (2002) Polypeptide hormones. *Plant Cell (Suppl)* **14**: S251–S264
- Schultz EA, Haughn GW (1991) LEAFY, a homeotic gene that regulates inflorescence development in *Arabidopsis*. *Plant Cell* **3**: 771–781
- Sessions A, Weigel D, Yanofsky MF (1999) The *Arabidopsis thaliana* *MERISTEM LAYER 1* promoter specifies epidermal expression in meristems and young primordia. *Plant J* **20**: 259–263
- Shpak ED, Berthiaume CT, Hill EJ, Torii KU (2004) Synergistic interaction of three ERECTA-family receptor-like kinases controls *Arabidopsis* organ growth and flower development by promoting cell proliferation. *Development* **131**: 1491–1501
- Shpak ED, Lakeman MB, Torii KU (2003) Dominant-negative receptor uncovers redundancy in the *Arabidopsis* ERECTA leucine-rich repeat receptor-like kinase signaling pathway that regulates organ shape. *Plant Cell* **15**: 1095–1110
- Soga K, Wakabayashi K, Hoson T, Kamisaka S (2000) Flower stalk segments of *Arabidopsis thaliana* ecotype Columbia lack the capacity to grow in response to exogenously applied auxin. *Plant Cell Physiol* **41**: 1327–1333
- Tobeña-Santamaría R, Bliok M, Ljung K, Sandberg G, Mol JNM, Souer E, Koes R (2002) FLOOZY of petunia is a flavin mono-oxygenase-like protein required for the specification of leaf and flower architecture. *Genes Dev* **16**: 753–763
- Torii KU (2004) Leucine-rich repeat receptor kinases in plants: structure, function, and signal transduction pathways. *Int Rev Cytol* **234**: 1–46
- Torii KU, Hanson LA, Josefsson CAB, Shpak ED (2003) Regulation of inflorescence architecture and organ shape by the *ERECTA* gene in *Arabidopsis*. In T Sekimura, ed, *Morphogenesis and Patterning in Biological Systems*. Springer-Verlag, Tokyo, pp 153–164
- Torii KU, Mitsukawa N, Oosumi T, Matsuura Y, Yokoyama R, Whittier RE, Komeda Y (1996) The *Arabidopsis* ERECTA gene encodes a putative receptor protein kinase with extracellular leucine-rich repeats. *Plant Cell* **8**: 735–746
- Ulmasov T, Murfett J, Hagen G, Guilfoyle TJ (1997) Aux/IAA proteins repress expression of reporter genes containing natural and highly active synthetic auxin response elements. *Plant Cell* **9**: 1963–1971
- Wang ZY, Nakano T, Gendron J, He J, Chen M, Vafeados D, Yang Y, Fujioka S, Yoshida S, Asami T, et al (2002) Nuclear-localized BZR1 mediates brassinosteroid-induced growth and feedback suppression of brassinosteroid biosynthesis. *Dev Cell* **2**: 505–513
- Wang ZY, Seto H, Fujioka S, Yoshida S, Chory J (2001) BRI1 is a critical component of a plasma-membrane receptor for plant steroids. *Nature* **410**: 380–383
- Weigel D, Ahn JH, Blazquez MA, Borevitz JO, Christensen SK, Fankhauser C, Ferrandiz C, Kardailsky I, Malanchruvil EJ, Neff MM, et al (2000) Activation tagging in *Arabidopsis*. *Plant Physiol* **122**: 1003–1013
- Wyatt RE, Ainley WM, Nagao RT, Conner TW, Key JL (1993) Expression of the *Arabidopsis* *AtAux2-11* auxin-responsive gene in transgenic plants. *Plant Mol Biol* **22**: 731–749
- Zhao Y, Christensen SK, Fankhauser C, Cashman JR, Cohen JD, Weigel D, Chory J (2001) A role for flavin monooxygenase-like enzymes in auxin biosynthesis. *Science* **291**: 306–309