Sequencing around 5-Hydroxyconiferyl Alcohol-Derived Units in Caffeic Acid O-Methyltransferase-Deficient Poplar Lignins\textsuperscript{1}[OA]

Fachuang Lu*, Jane M. Marita, Catherine Lapierre, Lise Jouanin, Kris Morreel, Wout Boerjan, and John Ralph

Department of Biochemistry and Department of Energy Great Lakes Bioenergy Research Center, University of Wisconsin, Madison, Wisconsin 53706 (F.L., J.R.); United States Dairy Forage Research Center, United States Department of Agriculture Agricultural Research Service, Madison, Wisconsin 53706 (J.M.M.); Institut Jean-Pierre Bourgin, INRA-AgroParisTech, UMR 1318, 78026 Versailles cedex, France (C.L., L.J.); Department of Plant Systems Biology, VIB, 9052 Ghent, Belgium (K.M., W.B.); and Department of Plant Biotechnology and Genetics, Ghent University, 9052 Ghent, Belgium (K.M., W.B.)

Caffeic acid O-methyltransferase (COMT) is a bifunctional enzyme that methylates the 5- and 3-hydroxyl positions on the aromatic ring of monolignol precursors, with a preference for 5-hydroxyconiferaldehyde, on the way to producing sinapyl alcohol. Lignins in COMT-deficient plants contain benzodioxane substructures due to the incorporation of 5-hydroxyconiferyl alcohol (5-OH-CA), as a monomer, into the lignin polymer. The derivatization followed by reductive cleavage method can be used to detect and determine benzodioxane structures because of their total survival under this degradation method. Moreover, partial sequencing information for 5-OH-CA incorporation into lignin can be derived from detection or isolation and structural analysis of the resulting benzodioxane products. Results from a modified derivatization followed by reductive cleavage analysis of COMT-deficient lignins provide evidence that 5-OH-CA cross couples (at its \(\beta\)-position) with syringyl and guaiacyl units (at their O-4-positions) in the growing lignin polymer and then either coniferyl or sinapyl alcohol, or another 5-hydroxyconiferyl monomer, adds to the resulting 5-hydroxyguaiacyl terminus, producing the benzodioxane. This new terminus may also become etherified by coupling with further monolignols, incorporating the 5-OH-CA integrally into the lignin structure.

Lignins are polymeric aromatic constituents of plant cell walls, constituting about 15% to 35% of the dry mass (Freudenberg and Neish, 1968; Adler, 1977). Unlike other natural polymers such as cellulose or proteins, which have labile linkages (glycosides and peptides) between their building units, lignins’ building units are combinatorially linked with strong ether and carbon-carbon bonds (Sarkanen and Ludwig, 1971; Harkin, 1973). It is difficult to completely degrade lignins. Lignins are traditionally considered to be dehydrogenative polymers derived from three monolignols, \(p\)-coumaryl alcohol \(1\H\) (which is typically minor), coniferyl alcohol \(1\G\), and sinapyl alcohol \(1\S\) (Fig. 1; Sarkanen, 1971). They can vary greatly in their composition in terms of their plant and tissue origins (Campbell and Sederoff, 1996). This variability is probably determined and regulated by different activities and substrate specificities of the monolignol biosynthetic enzymes from different sources, and by the carefully controlled supply of monomers to the lignifying zone (Sederoff and Chang, 1991).

Recently there has been considerable interest in genetic modification of lignins with the goal of improving the utilization of lignocellulosics in various agricultural and industrial processes (Baucher et al., 2003; Boerjan et al., 2003a, 2003b). Studies on mutant and transgenic plants with altered monolignol biosynthesis have suggested that plants have a high level of metabolic plasticity in the formation of their lignins (Sederoff et al., 1999; Ralph et al., 2004). Lignins in angiosperm plants with depressed caffeic acid O-methyltransferase (COMT) were found to derive from significant amounts of 5-hydroxyconiferyl alcohol (5-OH-CA) monomers \(1\Sh\) (Fig. 1) substituting for the traditional monomer, sinapyl alcohol \(1\S\) (Marita et al., 2001; Ralph et al., 2001a, 2001b; Jouanin et al., 2004; Morreel et al., 2004b). NMR analysis of a lignin from COMT-deficient poplar (\textit{Populus} spp.) has revealed that novel benzodioxane structures are formed through \(\beta\)-O-4 coupling of a monolignol with 5-hydroxyguaiacyl units (resulting from coupling of...
5-OH-CA), followed by internal trapping of the resultant quinone methide by the phenolic 5-hydroxyl (Ralph et al., 2001a). When the lignin was subjected to thioacidolysis, a novel 5-hydroxyguaiacyl monomer 2 (Fig. 1) was found in addition to the normal guaiacyl and syringyl thioacidolysis monomers (Jouanin et al., 2000). Also, a new compound 3G (Fig. 1) was found in the dimeric products from thioacidolysis followed by Raney nickel desulfurization (Lapierre et al., 2001; Goujon et al., 2003).

Further study with the lignin using the derivatization followed by reductive cleavage (DFRC) method also confirmed the existence of benzodioxane structures, with compounds 4 (Fig. 1) being identified following synthesis of the authentic parent compounds 9 (Fig. 2). However, no 5-hydroxyguaiacyl monomer could be detected in the DFRC products. These facts imply that the DFRC method leaves the benzodioxane structures fully intact, suggesting that the method might therefore be useful as an analytical tool for determining benzodioxane structures that are linked by β-O-4 ethers. Using a modified DFRC procedure, we report here on results that provide further evidence for the existence of benzodioxane structures in lignins from COMT-deficient plants, that 5-OH-CA is behaving as a rather ideal monolignol that can be integrated into plant lignins, and demonstrate the usefulness of the DFRC method for determining these benzodioxane structures.

RESULTS AND DISCUSSION

Benzodioxane Products Released by DFRC

Our previous NMR and DFRC studies on COMT-deficient poplar lignins showed that the 5-OH-CA monomer cross couples with guaiacyl or syringyl lignin units into the growing lignin oligomer followed by further coupling with monolignols producing benzodioxane structures (Marita et al., 2001, 2003b; Ralph et al., 2001a, 2001b; Jouanin et al., 2004; Morreel et al., 2004a, 2004b; for review, see Boerjan et al., 2003b; Ralph et al., 2004). When COMT-deficient poplar lignins or whole plant cell wall fractions were submitted to DFRC degradation, a dimeric product containing the benzodioxane structure was released and identified to be compound 4G (Fig. 1; Lapierre et al., 2001). Thioacidolysis also produced the corresponding benzodioxane products 3G/S; substantial amounts of monomer 2, which may be the cleavage product of benzodioxanes, were also detected. The fact that 5-hydroxyconiferyl acetate has not been found in DFRC degradation products of COMT-deficient poplar lignins implies that all of the 5-OH-CA monomer is incorporated into lignin as benzodioxane structures and these structures totally survive DFRC conditions (without cleavage to monomeric products). To test this absolute survivability, lignin dimeric models 13 (Fig. 3) were synthesized and subjected to DFRC degradation. The results (Fig. 3) showed that DFRC of benzodioxane compound 13Gf (with a free phenol) gave benzodioxane compound 14Gf with the hydroxyls on the A unit acetylated and the hydroxyl on the B unit reduced to methyl; a tiny amount of compound 15Gf with all hydroxyls acetylated was also produced, but no monomeric units. With compound 13Ge that models the internal (etherified) benzodioxane structures in lignin, similar results were obtained, i.e. the major product was compound 14Ge and minor amounts of compound 15Ge were also detected. The behaviors of compounds 13Gf and 13Ge during each step of DFRC treatment were monitored by NMR.
Acetyl bromide in acetic acid cleanly acetylated \( \gamma \)-hydroxyls and free phenols, and brominated the benzylic hydroxyl groups. Zinc reduced the benzylic bromide on the B ring to methyl. The minor benzyl acetate remaining on the B unit was the result of hydrolysis of the benzylic bromide during the zinc reduction step, followed by acetylation. In general, the results shown in Figure 3 confirmed that benzodioxane structures totally survived DFRC conditions, suggesting that DFRC can be used to detect and determine benzodioxane structures in lignins of COMT-deficient plants, and also from F5H up-regulated plants such as Arabidopsis \( (Arabidopsis\ thaliana)\) that have been shown to contain lignins with benzodioxane structures (Ralph et al., 2001b).

**Partial Sequencing around Benzodioxanes in COMT-Deficient Poplar Lignins**

When a COMT-deficient poplar lignin was degraded by the standard DFRC procedure, the benzodioxane marker compound 4G (Fig. 1) was detected by gas chromatography-mass spectrometry (GC-MS). The double bond present signifies that a \( \beta \)-O-4 bond was cleaved; the phenolic end may also result from cleaving a 4-O-\( \beta \) ether, or it could have already been free phenolic. But the absence of the corresponding analog 4s raised a question as to whether the syringyl benzodioxane was not in this lignin, or if the marker compound 4s simply could not get through the GC. With synthetic compound 4s, we found that 4s could not survive the GC conditions because of its thermolability, but the more thermally stable trimethylsilyl (TMS)-derivatized 9sf and 9se (Fig. 2), analogs of 4s, proved to be amenable to GC quantitative analysis. So a modified DFRC procedure was developed (Fig. 4) to determine the benzodioxane structures to get more detailed structural picture of such structures in lignin polymers. First of all, the normal DFRC method results in cleavage of \( \beta \)-ethers but leaves the benzodioxanes intact. After zinc reduction, the phenols released due to \( \beta \)-ether cleavage are methylated, and the phenolic acetates derived from lignins’ originally free phenols remain unaffected. So the benzodioxanes with free phenols on their G/S units in lignins will be acetylated by acetyl bromide and remain acetylated, whereas the benzodioxanes with etherified phenols on their G/S units become methylated. A solid-state extraction step, although not absolutely necessary, is effective in cleaning up the sample and enriching the dimeric products, allowing for more accurate GC analysis. The final step in this modified DFRC procedure, base hydrolysis to remove acetates, converts all benzodioxane dimeric products into compounds 9. Their TMS derivatives were then analyzed by GC.

The partial GC-flame ionization detector profiles of the degradation products of four samples by the modified DFRC procedure are shown in Figure 5 (along with the reference model spectrum in Fig. 5A). The target compounds 9 were identified, as their TMS derivatives, by their mass spectra and GC retention time comparison with synthetic and authentic

---

**Figure 2.** Synthesis of benzodioxane DFRC products 12 (see later in Fig. 6 for their structures). i, NaH, THF. ii, Pyrrolidine. iii, \( 1_6 \) or \( 1_8 \), benzene/acetone (4/1, v/v). iv, DIBAL-H, toluene. v, Iodomethane-K\( \text{CO}_3 \), acetone. vi, Ac\( _2 \)O pyridine.

---

---

---
model compounds in separate GC-MS runs. Theoretically, all of the compounds 9 can have four isomers, but only one or two isomers for each were significant enough to be detected. Based on results from NMR and model studies on free radical coupling of 5-OH-CA or 5-hydroxyferulate with hydroxycinnamyl alcohols, the benzodioxane ring having its two oxygen substituents in the trans-configuration (i.e. the $RR/SS$ pair of enantiomers) would be major. The trans-double bond on the 5H unit is expected to be major according to the mechanism involved in DFRC reactions (Lu and Ralph, 1997b). Therefore the dominant benzodioxane products identified by GC profiles are the ones with a trans-ring and a trans-double bond, although small amounts of isomers having a cis-ring and a trans-double bond were also found by comparing

**Figure 3.** DFRC reactions of benzodioxane lignin model compounds 13, and GC chromatograms of the DFRC products 14 and 15.

![Model compounds and GC chromatograms](image)

**Figure 4.** Benzodioxane compounds 9 released from DFRC degradation (using the modified post-treatment methods described here) of a hypothetical lignin-containing free-phenolic and etherified benzodioxane units.

![Benzodioxane compounds](image)
their retention times and mass spectra with those of authentic models. An internal standard, mono-4-O-methylated 5,5-diferulic acid, which has been used previously (Bunzel et al., 2003) but was synthesized here in pure form by an improved method (F. Lu, unpublished data), was added to samples right before TMS derivatization and GC analysis. At this moment, the GC results reflect the benzodioxanes actually recovered by this procedure after DFRC degradation, uncorrected for any losses. From the similarities among compounds 9, it is expected that the relative ratios among them should not change through the procedure, i.e. they should not be subjected to any differential partitioning.

From the results summarized in the Table I, it can be seen that guaiacyl benzodioxanes released by DFRC were mostly (70%–86%) from the ends of lignin molecules and the released syringyl benzodioxanes were mostly (67%–77%) from internal units. In other words, the released guaiacyl benzodioxanes came from phenolic units whereas the released syringyl benzodioxanes came from those etherified through 4-O-β-linkages. This is fairly typical of observations from analysis of thioacidolysis products from methylated cell walls. For example, in the wild-type Arabidopsis samples in a recent report (Ralph et al., 2008), and in earlier pine (Pinus spp.) and poplar studies (Lapierre and Rolando, 1988), released S-monomers were more highly etherified than their guaiacyl counterparts. These results may be explained, in part, by the lignification mechanism. Guaiacyl units with an additional active site at their 5-positions could, during lignification, form 5-β-, 5-O-γ-, and 5-5-linkages that are not cleavable by DFRC (or thioacidolysis). Therefore, there is less chance for guaiacyl units than syringyl ones to be etherified through 4-O-β-linkages that are cleaved by DFRC. Moreover, it has been shown that cross coupling between a guaiacyl unit and sinapyl alcohol is an unfavorable reaction (Landucci and Ralph, 2001), and that sinapyl alcohol dominates the monolignol supply at the later stages of lignification (Terashima et al., 1993). Therefore, if the supply of 5-OH-CA is similar to that of the sinapyl alcohol it replaces, most of the guaiacyl benzodioxanes that are released here are formed presumably at late stages. However, guaiacyl benzodioxanes formed earlier have more chance to be condensed at their 5-positions and so contribute less to the amount of released benzodioxanes. On the other hand, more internal syringyl benzodioxanes were released by DFRC because syringyl units are able to couple with sinapyl alcohol or cross couple with coniferyl alcohol only through 4-O-β-linkages that are cleavable by DFRC.

By comparing the results from dioxane-water-extracted lignins (so-called milled wood lignins [MWLs]) and the residual lignins remaining after the dioxane-water extraction, it is evident that some partitioning of benzodioxane structures between lignin fractions has occurred during the preparation of the MWL fraction, and that this partitioning is more significant for guaiacyl benzodioxanes than for syringyl ones.

Although guaiacyl benzodioxanes have more chance to have condensed linkages at their 5-positions, more guaiacyl benzodioxanes were released than syringyl ones. The reason for this may be that 5-hydroxy units react more favorably with coniferyl alcohol than with sinapyl alcohol. For example, the yield for compound 8G from coupling of 5-hydroxyferulate with coniferyl alcohol.
Table 1. Yields* and relative ratios of benzodioxanes released by modified DFRC

<table>
<thead>
<tr>
<th>COMT-Deficient Poplar Samples</th>
<th>Relative Ratios</th>
<th>Yields (% Lignin Sample)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>GOMe/GOH 9G/9c</td>
<td>SOMe/SOH 9e/9s</td>
</tr>
<tr>
<td>MWL-1 (antisense)</td>
<td>20.0/80.0</td>
<td>72.0/28.0</td>
</tr>
<tr>
<td>Residual lignin (antisense)</td>
<td>30.0/70.0</td>
<td>71.5/28.5</td>
</tr>
<tr>
<td>Cell wall (antisense)</td>
<td>13.5/86.5</td>
<td>66.7/33.3</td>
</tr>
<tr>
<td>MWL-2 (sense)</td>
<td>26.5/73.5</td>
<td>77.3/22.7</td>
</tr>
</tbody>
</table>

* The measured values listed here are means of replica experimental results and relative errors were within 10%; **, the amount was too low to be determined accurately.

alcohol (Fig. 2) is higher than that for compound 8s from coupling of 5-hydroxyferulate with sinapyl alcohol. Research of this kind desperately needs a method to determine relative cross-coupling propensities between all combinations of natural and novel (from transgenics) units. Also of course, sinapyl alcohol and syringyl units are depleted by COMT deficiency.

One important aspect to note (that also applies to a lesser degree to the thioacidolysis products) is that the overall yields for benzodioxanes released by DFRC are relatively low compared to the high levels found via NMR analysis of the lignins. Since 5-hydroxy units could also couple with further 5-OH-CA monomers producing benzoxydioxane chains that are not cleavable by DFRC, trimers, tetraters, and higher oligomers of benzoxydioxane chains may exist in DFRC products and cannot be measured by the current GC method. In fact, evidence for such benzoxydioxane chains has already been demonstrated in a COMT-deficient alfalfa (Medicago sativa) transgenic (Marita et al., 2003a) and such oligomers have been found in the methanol-soluble phenolics fraction of xylem tissue from COMT-deficient poplar plants (Morreel et al., 2004b). As we discuss next, the isolation of those DFRC trimers or tetraters will give further evidence that 5-OH-CA is truly acting as a monolignol that is well integrated into lignins in COMT-deficient plants.

DFRC Trimmers with Benzodioxane Structures from COMT-Deficient Poplar Lignins

From our work on synthesis of benzoxydioxane lignin model compounds by radical coupling reactions of coniferyl alcohol (1G) and 5-OH-CA (15H), it was evident that the 15H radical generated from one-electron oxidation by silver (I) oxide, or by peroxidase-hydrogen peroxide, has similar chemical reaction propensities to 1G or 1S radicals. Therefore it is reasonable to expect that lignins from COMT-deficient plants in which significant amounts of 15H are produced have such benzoxydioxane structures. The 15H radical or radicals from 5-hydroxyguaiacyl units of oligomers could couple with 15H radical producing 5H-5H benzodioxane structures as we demonstrated in the synthesis of 12 (Figs. 2 and 6). DFRC reactions cleave normal β-O-4-ether linkages between two lignin (guaiaacyl and/or syringyl) units but not the benzoxydioxane linkages. So monomers (acetylated 1H, 1G, and 1S) are produced from DFRC degradation of lignins but not acetylated 15H. Instead, benzoxydioxane (G/S-5H) dimers linked through β-O-4 ethers resulted from DFRC degradation of COMT-deficient poplar lignins. We surmised that benzoxydioxane (G/S-5H-5H) trimers linked with β-O-4 ethers would also be produced from DFRC degradation of COMT-deficient poplar lignins. Such acetylated trimers, if produced, will not be detected by GC-MS due to their low volatility and their likely thermolability. However they could be isolated through a series of liquid chromatography and thin-layer chromatography (TLC) steps (Peng et al., 1998, 1999).

Large-scale DFRC experiments were performed with COMT-deficient poplar wood to isolate the expected trimeric products having benzoxydioxane structures. First of all, the final degradation products recovered by extractions were applied to normal-phase silica-gel flash chromatographic separation to produce fractions having monomers, dimers, trimers, and oligomers. After checking by TLC, fractions having similar compositions were combined. Using synthesized benzoxydioxane trimers to reveal TLC mobilities, fractions containing the expected trimers were pooled out, deacetylated by treatment with pyrrolidine in ethanol, followed by ethyl acetate extraction to remove the contaminating degraded carbohydrates. Second, the ethyl acetate-extracted trimers were acetylated (in acetic anhydride and pyridine) and subjected to further TLC separation. Two fractions (fraction A and fraction B) potentially having the benzoxydioxane trimers were recognized by comparing with synthesized models.

Two-dimensional (2D) NMR experiments were performed on such isolated samples. 2D COSY, HSQC, and HMBC NMR spectra were used to identify the expected benzoxydioxane trimers (Fig. 6). From COSY spectra of fraction A and fraction B (not shown here), two correlation signals, at 4.45/5.08 ppm and 4.39/4.95 ppm, were found. These are diagnostic COSY signals for benzoxydioxane trimeric compounds 12 (Fig. 6). Meanwhile, proton signals at 4.39 and 4.45 ppm...
also see signals at around 4.05 and 4.30 ppm (not shown here), which are consistent with model compounds 12. As shown in HSQC NMR spectra (Fig. 6, A2 and D2), C-H correlations at around 77.0 to 77.5 (δ_C)/4.95 to 5.05 (δ_H) ppm and around 76.0 to 77.0 (δ_C)/4.36 to 4.50 (δ_H) ppm, are also characteristic of benzodioxane structures according to NMR data from model compounds 12 (Fig. 6, B2 and C2). Moreover, C-H correlations at around 77.0 to 77.5 (δ_C)/4.95 to 5.05 (δ_H) ppm were clearly separated into two distinct networks, suggesting that two kinds of benzodioxane structures exist in these compounds. Comparison of the HSQC NMR spectrum of fraction A (Fig. 6, A2) with that of synthesized benzodioxane trimers 12 (Fig. 6, B2 and C2) suggests that these two distinguishable correlations at around 77.0 to 77.5 (δ_C)/4.95 to 5.05 (δ_H) ppm belong to the Cα-Hα correlation of benzodioxanes constructed by a G/S unit and a 5H unit (B ring;
CONCLUSION

Whereas trimer (Fig. 6), and the Cα-Hα correlation of a benzodioxane constructed by two 5H units (B ring and C ring; Fig. 6), respectively. Further evidence for such benzodioxane structures can be found in the aromatic regions of HSQC spectra when we compare A1 (fraction A) and B1 (fraction B) with B1 (fraction A) and C1 (12c). Thus C-H correlations from the 2- and 6-positions of 5H units (B and C rings; Fig. 6) were readily identified. Also found from these spectra are C-H correlations from the 2-, 5-, and 6-positions of G units (12g; Fig. 6) and C-H correlations from 2/6-positions of S units (12s; Fig. 6), and even those from the α- and β-positions of cinnamyl acetate end groups (C ring; Fig. 6). All those C-H correlations found in these HSQC spectra of fraction A and fraction B are fully consistent with those from synthesized benzodioxane trimers 12. However, integration of contour volumes from the well-separated aromatic correlations (at the 6-positions) in the HSQC spectra (Fig. 6, A1 and D1) suggests that fraction A contains almost only benzodioxane trimer 12c whereas trimer 12s was dominant in fraction B.

MATERIALS AND METHODS

General

All chemicals and reagents were from Aldrich and used as supplied. Solvents (analytical reagent grades) were from Mallinckrodt. Evaporations were conducted under reduced pressure at temperatures <40°C. Further removal of organic solvents, as well as drying of residues, was accomplished under higher vacuum (100–200 mTorr) at room temperature. Flash chromatography was performed using FLASH 40-M cartridges on a Biotage Isolera Max under higher vacuum (100–200 mTorr) at room temperature. Flash chromatography was performed using FLASH 40-M cartridges on a Biotage Isolera Max and C176 C. Further

into lignins like the traditional monolignols. 5-OH-CA is cross coupling (at its β-position) with the phenolic end of growing lignin oligomers, and new monolignols (1G, 1S, and 15H) can all cross couple with the newly formed 5-hydroxyguaiaacyl end unit, extending the chain in a typical endwise fashion, and forming benzodioxane structures. Further monolignols will cross couple with the new end unit from this latest addition (G, S, or 5H unit), such that it too becomes further etherified forming new 4-O-β-bonds (for G and S end units) or another benzodioxane (for 5H end units). The isolated trimeric G/S-5H-5H benzodioxane products from DFRC degradation provide further evidence for such lignification mechanisms in COMT-deficient plants.

Table II. NMR data for synthesized benzodioxane trimer acetates 12

<table>
<thead>
<tr>
<th>G-SH-5H, 12c</th>
<th>S-SH-5H, 12s</th>
</tr>
</thead>
<tbody>
<tr>
<td>1H</td>
<td>1H</td>
</tr>
<tr>
<td>Gα 5.06, 1H, d, J = 7.7 Hz</td>
<td>Sα 5.05, 1H, d, J = 7.8 Hz</td>
</tr>
<tr>
<td>GB 4.42, 1H, m</td>
<td>Sβ 4.46, 1H, m</td>
</tr>
<tr>
<td>Gy 4.03, 1H, m, 4.30, 1H, m</td>
<td>Sy 4.06, 1H, m, 4.30, 1H, m</td>
</tr>
<tr>
<td>G1 –</td>
<td>S1 –</td>
</tr>
<tr>
<td>G2 7.25, 1H, d, J = 1.7 Hz</td>
<td>S2 6.90, 2H, s</td>
</tr>
<tr>
<td>G3 –</td>
<td>S3 –</td>
</tr>
<tr>
<td>G4 –</td>
<td>S4 –</td>
</tr>
<tr>
<td>G5 7.12, 1H, d, J = 8.2 Hz</td>
<td>S5 –</td>
</tr>
<tr>
<td>G6 7.70, 1H, dd, J = 8.2, 1.7 Hz</td>
<td>S6 6.90, 2H, s</td>
</tr>
<tr>
<td>Ba 4.94, d, J = 7.6 Hz</td>
<td>Ba 4.96, d, J = 7.6 Hz</td>
</tr>
<tr>
<td>Bb 4.35, 1H, m</td>
<td>Bb 4.39, 1H, m</td>
</tr>
<tr>
<td>By 4.03, 1H, m, 4.30, 1H, m</td>
<td>By 4.06, 1H, m, 4.30, 1H, m</td>
</tr>
<tr>
<td>B1 –</td>
<td>B1 –</td>
</tr>
<tr>
<td>B2 6.77, 1H, d, J = 1.80 Hz</td>
<td>B2 6.79, 1H, brd</td>
</tr>
<tr>
<td>B3 –</td>
<td>B3 –</td>
</tr>
<tr>
<td>B4 –</td>
<td>B4 –</td>
</tr>
<tr>
<td>B5 –</td>
<td>B5* –</td>
</tr>
<tr>
<td>B6 6.73, 1H, d, J = 1.8 Hz</td>
<td>B6 6.74, 1H, d, J = 2.0 Hz</td>
</tr>
<tr>
<td>Ca 6.56, 1H, brd, J = 15.9 Hz</td>
<td>Ca 6.57, 1H, brd, J = 15.9 Hz</td>
</tr>
<tr>
<td>CB 6.23, 1H, dt, J = 15.9, 6.3 Hz</td>
<td>CB 6.24, 1H, dt, J = 15.9, 6.3 Hz</td>
</tr>
<tr>
<td>Cy 4.65, 2H, d, J = 6.3 Hz</td>
<td>Cy 4.66, 2H, d, J = 6.3 Hz</td>
</tr>
<tr>
<td>Cl –</td>
<td>Cl –</td>
</tr>
<tr>
<td>C2 6.76, 1H, d, J = 1.8 Hz</td>
<td>C2 6.77, 1H, d, J = 1.9 Hz</td>
</tr>
<tr>
<td>C3 –</td>
<td>C3 –</td>
</tr>
<tr>
<td>C4 –</td>
<td>C4 –</td>
</tr>
<tr>
<td>C5 –</td>
<td>C5* –</td>
</tr>
<tr>
<td>C6 6.65, 1H, d, J = 1.8 Hz</td>
<td>C6 6.66, 1H, d, J = 1.9 Hz</td>
</tr>
</tbody>
</table>

* The chemical shifts for B5 and C5 are close to each other and may be interchangeable.
**Synthesis of Model Compounds**

Compound 8 (Fig. 2), 5-hydroxyavlin diacetate, was made quantitatively from 5-hydroxyavin by acetylation with acetic anhydride pyridine. Compound 6, ethyl 5-hydroxyferulate diacetate. Sodium hydride (750 mg, 31.25 mmol) was suspended in 250 mL dry triethyl phosphonoacetate (THF) in a 500 mL round-bottom flask, to which THF (70.1, 31.22 mmol) was added slowly (dropwise). A clear solution was formed when hydrogen gas evolution had ceased. Compound 5 (6.0 g, 23.80 mmol) was added into the solution while well stirred. Stirring was continued for another hour before adding 50 mL 3% HCl solution. THF solvent in the resultant mixture was removed by evaporation at 40°C under reduced pressure during which a white solid product precipitated. The crude product was extracted with ethyl acetate (2 x 200 mL). After being dried over anhydrous MgSO4 and filtered, the ethyl acetate solution was evaporated to produce product (7.27 g, 94% yield). NMR analysis indicated that the crude product was pure enough for the next reaction. Compound 6, NMR, δ 1.27 (1H, t, J = 7.15 Hz, Me), 2.25 (3H, s, OAc), 2.26 (3H, s, OAc), 3.91 (3H, s, A-OMe), 4.20 (2H, q, J = 7.15 Hz, CH3), 6.57 (1H, d, J = 15.97 Hz, Aδ); 7.15 (1H, d, J = 1.84, A6), 7.37 (1H, d, J = 1.84 Hz, A2), 7.61 (1H, d, J = 15.97 Hz, Aα); 14.53 (Me), 20.07 (OAc), 39.03 (OAc), 56.79 (A-OMe), 60.84 (CH3), 109.92 (A2), 116.19 (A6), 120.19 (A3), 136.63 (A1), 134.57 (A4), 143.81 (Aa), 144.86 (Aa), 153.78 (Aa), 166.72 (Aa), 167.84 (OAc), 168.38 (OAc).

Compound 7 was made from 6 by deacetylation with neat pyrrolidine (Lu and Ralph, 1998). NMR, δ 1.25 (1H, t, J = 7.14 Hz, Me), 3.36 (3H, s, A-OMe), 4.17 (2H, q, J = 7.14 Hz, CH3), 6.34 (1H, d, J = 15.90 Hz, Aα); 6.85 (1H, brs, A6), 6.88 (1H, brs, A2), 7.53 (1H, d, J = 15.90 Hz, Aα); 14.46 (Me), 56.41 (A-OMe), 60.50 (CH3), 104.30 (A2), 106.50 (A6), 108.45 (A3), 126.37 (A1), 137.31 (A4), 145.83 (Aa), 146.21 (Aa), 149.01 (Aa), 163.47 (Aa).

Compound 8c (707 mg, 3.65 mmol) and 1G (788 mg, 4.38 mmol) were dissolved in benzenecacete (45 mL, 21, 1/1) in 100 mL round bottom flask and silver carbonate (Ag2CO3, 2.52 g, 9.13 mmol) was added. The resulting mixture was stirred overnight by which time no starting materials remained, as shown by TLC (cyclodioxane ethyl acetate, 3:1, v/v). After filtering off the solids, the organic solvents were evaporated at 40°C under reduced pressure. The residual products were redissolved in 5 mL dichloromethane and applied to a flash chromatography with cyclodioxane ethyl acetate (3:1, v/v) as eluting solvent. The major product, compound 8c (white solid, 800 mg), was obtained in 53% yield. NMR: δ 3.50 (1H, m, Aγy), 3.80 (1H, m, Aγy), 3.85 (3H, s, A-OMe), 3.89 (3H, s, B-OMe), 4.09 (1H, m, Aγ), 4.98 (1H, d, J = 8.01 Hz, Aα), 6.11 (1H, d, J = 1.84, A2), 6.54 (1H, d, J = 15.78 Hz, Aδ), 6.88 (1H, d, J = 8.01 Hz, Aα), 6.95 (1H, d, J = 1.76 Hz, B2), 6.96 (1H, dd, J = 8.0, 1.73 Hz, Aδ), 7.01 (1H, d, J = 1.73 Hz, A2); δ 56.19 (A-OMe), 56.29 (B-OMe), 61.55 (Aγ), 76.85 (Aα), 79.52 (Aδ), 104.70 (B2), 111.12 (B1), 117.33 (Aδ), 115.68 (A5), 116.98 (B12), 121.43 (A6), 123.77 (B1), 128.80 (A1), 136.44 (B5), 145.22 (B2), 145.34 (B4), 148.33 (Aa), 148.30 (Aa), 150.10 (B3), and 163.27 (B2).

Compound 8a was made in 45% yield from 1S and 7 in the analogous way as described above for compound 8c. NMR, δ 3.50 (1H, m, Aγy), 3.82 (1H, m, Aγy), 3.85 (3H, s, A-OMe), 3.89 (3H, s, B-OMe), 4.09 (1H, m, Aγ), 4.98 (1H, d, J = 8.01 Hz, Aα), 6.11 (1H, d, J = 1.84, A2), 6.54 (1H, d, J = 15.78 Hz, Aδ), 6.88 (1H, d, J = 8.01 Hz, Aα), 6.95 (1H, d, J = 1.76 Hz, B2), 6.96 (1H, dd, J = 8.0, 1.73 Hz, Aδ), 7.01 (1H, d, J = 1.73 Hz, A2); δ 56.19 (A-OMe), 56.29 (B-OMe), 61.55 (Aγ), 76.85 (Aα), 79.52 (Aδ), 104.70 (B2), 111.12 (B1), 117.33 (Aδ), 115.68 (A5), 116.98 (B12), 121.43 (A6), 123.77 (B1), 128.80 (A1), 136.44 (B5), 145.22 (B2), 145.34 (B4), 148.33 (Aa), 148.30 (Aa), 150.10 (B3), and 163.27 (B2).
Synthesis of compound 13c (Fig. 3) has been reported previously (Ralph et al., 2001a). Compound 13c was made from 13c by methylation with iodomethane and K₂CO₃ in acetone. NMR, δ: 5.30 (H, m, A), 3.80 (H, m, A), 3.80 (H, br-s, A/B-OMe), 4.02 (H, m, A), 4.51 (H, m, A), 4.71 (H, m, A), 4.97 (H, d, J = 7.85 Hz), 6.35 (H, br-d, B), 6.90 (H, br-d, B), 6.95 (H, d, J = 8.30 Hz, A5), 7.01 (H, dd, J = 8.30, 1.60 Hz, A6), 7.08 (H, d, J = 1.60 Hz, A2); δ²: 61.78 (Ay), 64.62 (Bz), 76.81 (Af), 79.20 (Az), 104.09 (Bz), 108.56 (Bz), 117.82 (Az), 121.51 (A5), 123.01 (A3), 133.17 (B4), 131.39 (B1), 149.05 (B3), 149.78 (B3), 150.18 (A4), 150.39 (A3).

Compounds 14 were obtained from DFRC treatment of compounds 13. Since these were almost quantitative conversions, compounds 14 were characterized directly after DFRC reactions, without any purification. Compound 14a: NMR, δ: 2.22 (3H, s, A), 3.79 (3H, s, B-OMe), 3.84 (3H, s, A-OMe), 4.01 (H, m, A), 4.26 (H, m, A), 4.33 (H, m, A), 5.01 (H, d, J = 7.65 Hz, Aa), 6.38 (H, d, J = 1.80 Hz, Bb), 6.44 (H, d, J = 1.80 Hz, Bb), 7.06 (H, dd, J = 8.04, 8.04 Hz, Aa), 7.11 (H, d, J = 8.04 Hz, Aa), 7.24 (H, d, J = 1.80 Hz, Ab); δ²: 21.18 (Ba), 56.29, 56.34, 63.48 (Ay), 75.85 (Af), 77.01 (Aa), 110.18 (Bb), 110.60 (Bb), 112.74 (A2), 120.68 (Ab), 123.84 (A5), 130.87 (B1), 131.65 (B4), 136.27 (A1), 141.37 (Ab), 144.81 (Bz), 152.54 (A3).

Modified DFRC Procedure for COMT-Deficient Lignins and Cell Wall Isolates

DFRC

Lignin (8–10 mg) or 40 mg extracted wood cell wall was used. The acetyl bromide treatment and zinc reduction steps were performed as per the standard DFRC procedure (Lu and Ralph, 1997a). After the zinc reduction step the degraded products were methylated as follows, instead of using the acetylation step from the original procedure.

Methylation

The above residue was methylated using iodomethane (50 mL) and cesium carbonate (100 mg) in 3 mL acetonitrile for 30 min. The excess reagents were quenched by addition of 1 mL acetic acid. The organic solvents were evaporated under reduced pressure. The methylated products were transferred into a separatory funnel containing 6 mL saturated sodium chloride aqueous solution with 15 mL dichloromethane. After shaking the mixture, the dichloromethane phase was collected and the water phase was extracted with further dichloromethane (2 × 6 mL). The combined dichloromethane solution was dried over anhydrous sodium sulfate, filtered, and evaporated under reduced pressure.

Solid Phase Extraction

The above residue was transferred with 2 mL dichloromethane into a 10 mL round-bottom flask. This mixture was gently stirred at 50°C for 3.5 h. After removal of all solvent and reagents by rotary evaporation at below 40°C, the residues were dissolved in 150 mL dioxane:AcOH:water (5:4:1, v/v/v) solution. Zinc dust, 8.0 g, was added in two parts while this solution was well stirred. Stirring was continued for 30 min, and then the mixture let stand for 30 min before the liquid contents were decanted with the help of some added acetone. The resulting liquid fraction was concentrated to about 100 mL and diluted with 200 mL water, the DFRC degradation products were extracted with dichloromethane (200 × 2 mL). After evaporation of the dichloromethane solution, the residues were acetylated with 20 mL acetic anhydride/pyridine (1:1, v/v) for 16 h.

Large-Scale DFRC on COMT-Deficient Poplar Cell Walls

To cryogenically (liquid N₂) milled-wood sawdust (extractive free, 8.5 g) were added 150 mL 20% (v/v) acetyl bromide/acetic acid solution in a 250 mL round-bottom flask. This mixture was gently stirred at 50°C for 3.5 h. After removal of all solvent and reagents by rotary evaporation at below 40°C, the residues were dissolved in 150 mL dioxane:AcOH:water (5:4:1, v/v/v) solution. Zinc dust, 8.0 g, was added in two parts while this solution was well stirred. Stirring was continued for 30 min, and then the mixture let stand for 30 min before the liquid contents were decanted with the help of some added acetone. The resulting liquid fraction was concentrated to about 100 mL and diluted with 200 mL water, the DFRC degradation products were extracted with dichloromethane (200 × 2 mL). After evaporation of the dichloromethane solution, the residues were acetylated with 20 mL acetic anhydride/pyridine (1:1, v/v) for 16 h.

Silica-Gel Flash Chromatographic Fractionation

The resulting products were purified by flash chromatography by applying to a normal-phase silica gel column (FLASH 40 mm cartridge), and eluting with cyclohexane:acetic acid (3:1, 1 L; 1:1, 1 L; 1:2, 0.75 L; and 0:1, 0.5 L) successively. A total of 12 fractions was collected.

TLC Separation

TLC comparison of the isolated fractions with synthesized benzoxadizone trimers 12 indicated that fractions 9 to 10 potentially contained the expected benzoxadizone products. NMR examination of these fractions showed that they were heavily contaminated with degraded carbohydrates and further purification was needed. Thus fractions 9 and 10 were combined and deacetylated with 2 mL pyridoline in 10 mL ethanol overnight. The lignin degradation products were recovered by ethyl acetate extraction. After being dried over MgSO₄ and filtered off, the ethyl acetate solvent was removed on a rotary evaporator at 40°C under reduced pressure. The residues were acetylated in Ac₂O pyridine producing about 80 mg products, after removing the acetylation reagents by evaporation. These acetylated products were applied to a TLC plate (silica gel, 1 mm) using cyclohexane:acetic acid (1:5, v/v) as eluant. Two fractions (A, 2.5 mg and B, 2.0 mg) with the same Rf values as trimers 12 were collected for NMR analysis.

ACKNOWLEDGMENT

The authors are grateful to the U.S. Department of Agriculture Dairy Forage Research Center for allowing access to Bruker AMX-360 NMR and HP 5980 GC chromatography, which was essential to this work.

Received February 2, 2010; accepted April 26, 2010; published April 28, 2010.
LITERATURE CITED


Freudentberg K, Neish AC (1968) Constitution and Biosynthesis of Lignin. Springer-Verlag, Berlin


Benzodioxane Structures in COMT-Deficient Poplar Lignins


579

Downloaded from on July 13, 2017 - Published by www.plantphysiol.org
Copyright © 2010 American Society of Plant Biologists. All rights reserved.