Dynamic Changes in the Distribution of Minerals in Relation to Phytic Acid Accumulation during Rice Seed Development¹[W]

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Phytic acid (inositol hexakisphosphate [InsP₆]) is the storage compound of phosphorus in seeds. As phytic acid binds strongly to metallic cations, it also acts as a storage compound of metals. To understand the mechanisms underlying metal accumulation and localization in relation to phytic acid storage, we applied synchrotron-based x-ray microfluorescence imaging analysis to characterize the simultaneous subcellular distribution of some mineral elements (phosphorus, calcium, potassium, iron, zinc, and copper) in immature and mature rice (Oryza sativa) seeds. This fine-imaging method can reveal whether these elements colocalize. We also determined their accumulation patterns and the changes in phosphate and InsP₆ contents during seed development. While the InsP₆ content in the outer parts of seeds rapidly increased during seed development, the phosphate contents of both the outer and inner parts of seeds remained low. Phosphorus, calcium, potassium, and iron were most abundant in the aleurone layer, and they colocalized throughout seed development. Zinc was broadly distributed from the aleurone layer to the inner endosperm. Copper localized outside the aleurone layer and did not colocalize with phosphorus. From these results, we suggest that phosphorus translocated from source organs was immediately converted to InsP₆ and accumulated in aleurone cells and that calcium, potassium, and iron accumulated as phytic acid salt (phytate) in the aleurone layer, whereas zinc bound loosely to InsP₆ and accumulated not only in phytate but also in another storage form. Copper accumulated in the endosperm and may exhibit a storage form other than phytate.

The transport of nutrients into developing seeds has received considerable attention. During the grain-filling stage, plants remobilize and transport nutrients distributed throughout the vegetative source organs into seeds. Plant seeds contain large amounts of phosphorus (P) in organic form, which supports growth during the early stages of seedling development. Most of the P in seeds is stored in the form of phytic acid (inositol hexakisphosphate [InsP₆]). Seeds also accumulate mineral nutrients such as potassium (K), magnesium (Mg), calcium (Ca), iron (Fe), zinc (Zn), copper (Cu), and manganese (Mn), which are used in seedling growth. Phytic acid acts as a strong chelator of metal cations and binds them to form phytate, a salt of InsP₆ (Lott et al., 2002; Raboy, 2009). During germination, phytate is decationized and hydrolyzed by phytases, and then inorganic phosphates, inositol, and various minerals are released from the phytate (Loewus and Murthy, 2000). Phytate accumulates within protein bodies, generally of vacuolar origin, in seed storage cells and is usually concentrated in spherical inclusions called globoids. Many studies of the elemental composition of phytate in seeds have been published. Energy-dispersive x-ray microanalyses of many plant species have revealed that, other than P, globoids contain mainly K and Mg as well as low levels of Ca, Mn, Fe, and Zn (Lott, 1984; Lott et al., 1995; Wada and Lott, 1997). This indicates that phytate is a mixed salt of these cations.

Whether all storage metal elements can bind equally to InsP₆ is not known, although most elements are thought to exist in seeds in the form of phytate (Raboy, 2009). To form phytate, P and the other elements must be present in the same place. Therefore, determination of the precise locations of P and other elements in seed tissues makes it possible to judge whether an element exists in the form of phytate. Differences in metal distribution with P might suggest a storage form other than phytate. For determining distributions, synchrotron-based x-ray microfluorescence (μ-XRF) imaging utilizing an x-ray microbeam is a powerful tool. The microbeam excites the elements, thereby revealing the details of their spatial distribution. The development of focusing optics for high-

¹ This work was supported by Japan Advanced Plant Science Network and the Japan Synchrotron Radiation Research Institute (proposal nos. 2009B1724 and 2010B1695).

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¹[W] The online version of this article contains Web-only data.

www.plantphysiol.org/cgi/doi/10.1104/pp.112.206573
energy x-rays using a Kirkpatrick-Baez mirror raises the imaging resolution of elements in μ-XRF analysis. A focal spot size smaller than 1 μm with x-ray energy as high as 100 keV enables detection of the subcellular distribution of elements in plant tissues (Fukuda et al., 2008; Takahashi et al., 2009).

Whether there is an order in the affinity of elements for phytic acid in plant cells remains unknown. The stability of InsP₆-metal complexes has been estimated by in vitro titration (Maddaiah et al., 1964; Vohra et al., 1965; Persson et al., 1998). The binding strength of InsP₆ with metal is stronger for Zn and Cu than for Fe, Mn, and Ca. We also do not know if the mineral composition of phytate in seeds is determined by the relative abundance of these elements in the seed or by their biochemical characteristics. As a first step to address these issues, we examined the simultaneous changes in the distribution of P and metal elements during seed development using μ-XRF imaging analysis.

Our objective in this study was to observe the dynamic changes in the distribution of some nutritionally important minerals (P, Ca, K, Fe, Zn, and Cu) in relation to the accumulation of phytic acid during rice (Oryza sativa) seed development.

## RESULTS

### Accumulation of Minerals in Developing Seeds

To determine the accumulation patterns of the minerals during seed development, we performed inductively coupled plasma optical-emission spectrometry (ICP-OES) analysis (Fig. 1). Mineral contents, except Ca, rapidly increased until 14 d after flowering (DAF), as the weight of the seeds increased (Fig. 2). Both the fresh and dry weights of seeds increased from 5 to 21 DAF. Levels of P, Fe, and Zn gradually increased between 14 and 21 DAF. In contrast, the increases in K and Cu contents almost stopped after 14 DAF. The increase in P and Zn halted after 21 DAF, as did the increase in seed weight. The Ca content in developing seeds differed greatly from that of other minerals, being nearly constant from 5 to 28 DAF. The Ca concentration on the basis of seed fresh weight decreased until 21 DAF as the fresh weight of seeds increased (Supplemental Fig. S1).

### Phosphate and InsP₆ Levels in Developing Seeds

Concentrations of phosphate (Pi) and InsP₆ during seed development were determined by ion chromatography (Fig. 3). We separately analyzed the inner starchy endosperm and the outer endosperm containing the aleurone and subaleurone layers. Pi concentration did not change greatly during seed development, being maintained at low levels in both the inner and outer endosperms. In contrast, InsP₆ concentration differed greatly between the two parts. The InsP₆ level of the inner starchy endosperm remained at low levels (0.3–1.0 mg g⁻¹) from 7 to 30 DAF, whereas that of the outer endosperm increased rapidly from 10 to 25 DAF. No signal peak of inositol phosphates other than InsP₆ was detected in these chromatography experiments. These results suggest that Pi is assimilated into InsP₆ form immediately following their transfer from vegetative organs to seeds.

After 20 DAF, the levels of Pi decreased slightly in both parts of the endosperm (Fig. 3). By this time, the translocation of P from vegetative organs into seeds had...
stopped (Fig. 1). Thus, the decrease in Pi contents after 20 DAF might be due to further InsP6 synthesis at this time.

Expression of OsIPK1 in Developing Seeds

The last step in phytic acid synthesis is the phosphorylation of inositol 1,3,4,5,6-pentakisphosphate to InsP6, catalyzed by inositol 1,3,4,5,6-pentakisphosphate 2-kinase (IPK1). Previous studies have revealed that levels of transcripts of the rice IPK1 homolog gene OsIPK1 in whole developing seeds increase until 7 DAF and then gradually decrease until 28 DAF (Suzuki et al., 2007). To obtain a general estimation of the localization of phytic acid biosynthesis, we assessed the expression of OsIPK1 by using reverse transcription (RT)-PCR and immunoblot analyses of immature seeds at 10 and 15 DAF, respectively (Fig. 4). First, we carefully separated the embryo, endosperm, and aleurone layer and then compared the expression levels of OsIPK1 in the three tissues. OsIPK1 transcripts were detected in the endosperms as well as in the embryos and the aleurone layers (Fig. 4A). OsIPK1 protein was also detected in the endosperm, although protein levels were rather lower in endosperms than in embryos and aleurone layers (Fig. 4B). This suggests that the biosynthesis of InsP6 progresses not only in the aleurone layer and embryo but also in the starchy endosperm. Because chromatography analysis showed InsP6 levels to be consistently low in the inner endosperm during seed development (Fig. 3), InsP6 synthesized in the inner endosperm may immediately transfer to the aleurone layer, or InsP6 synthesis may occur only in the outer endosperm (i.e. in both the aleurone and subaleurone layers) but not in the inner endosperm.

Mineral Distribution Changes during Rice Seed Development

Next, we determined the localization of P, Ca, K, Fe, Zn, and Cu in developing seeds by μ-XRF analysis. Cross sections of the center of freeze-dried immature rice seeds at 10, 15, 20, and 25 DAF were used for x-ray imaging. Figure 5 shows the elemental maps of each element. From 10 to 25 DAF, P accumulated around the aleurone layer and was most abundant on the lateral side of the seed. The localization of Ca, K, and Fe was similar to that of P, as they also accumulated around the aleurone layer during seed development. The differences in Ca, K, and Fe concentrations between the aleurone layer and the inner starchy endosperm were notable. The distribution patterns of the three elements differed somewhat. Ca accumulated uniformly around the aleurone layer, while K, like P, was abundant in the lateral side, and Fe was abundant in the dorsal side, where several vascular bundles, which act as gates for translocation materials, were present.

In contrast, the distribution patterns of Zn and Cu differed from that of P (Fig. 5). Although Zn and Cu were most abundant around the aleurone layer, they were also distributed broadly in the inner endosperm. Cu was abundant on the ventral side near the aleurone layer until 20 DAF. After 20 DAF, Cu spread deeper into the endosperm. Because the translocation of Cu from the vegetative organs was complete before 15 DAF (Fig. 1), this change in localization may depend on the movement of Cu that exists around the aleurone layer. At 25 DAF, Cu was most abundant on the lateral side of the seed. The distribution of Zn changed rapidly during seed development. At 10 DAF, Zn was abundant around the aleurone layer. After 10 DAF, Zn decreased around the aleurone layer and spread into the inner endosperm. Zn exhibited a broader distribution in the endosperm than did Cu and was detected at similar levels in both the central and outer areas of the seed.
the endosperm. Zn was most abundant in the aleurone layer of the dorsal to lateral sides throughout seed development. After 20 DAF, when the translocation of Zn from the vegetative organs had ended (Fig. 1), Zn accumulation around the aleurone layer decreased slightly.

To obtain high-resolution images, we changed the step size of the μ-XRF analysis from 50 μm to 1 μm. At this size, we could compare the exact positions of each element. Figure 6A compares the elemental maps of P, Ca, K, and Fe around the aleurone layer on the lateral side of developing seeds. The distribution areas of Ca, K, and Fe were quite similar to that of P during seed development from 10 to 25 DAF, even though the regions in which each element was most abundant differed slightly. Figure 6B compares the P, Zn, and Cu elemental maps. The distribution of Zn was similar to that of P but was slightly broader, which is apparent in the image at 25 DAF. The Cu accumulation region did not strictly overlap with that of P throughout seed development, and Cu localization changed gradually from 10 to 25 DAF. The area of most abundant Cu moved from the outer to the inner endosperm, and at 25 DAF it was completely different from the site of P localization.

**DISCUSSION**

In this study, μ-XRF imaging analysis revealed the dynamic spatial distributions of elements (P, Ca, K, Fe, Zn, and Cu) in developing rice seeds. P accumulated to high levels in the outer cell layers of the endosperm in the endosperm. Zn was most abundant in the aleurone layer of the dorsal to lateral sides throughout seed development. After 20 DAF, when the translocation of Zn from the vegetative organs had ended (Fig. 1), Zn accumulation around the aleurone layer decreased slightly.

Marignano et al. (2010) noted that the localization patterns of these elements did not change markedly (Fig. 7A). P, Ca, K, and Fe accumulated mainly in the aleurone layer. However, Zn and Cu broadly accumulated from the aleurone layer to the inner endosperm. Figure 7B shows the fine elemental maps around the aleurone layer on the ventral side of the seeds as determined by 1-μm step-size analysis. As in the case of developing seeds, the distributions of P, Ca, K, and Fe in the mature seed were similar. The localization sites of these elements were mostly restricted to the aleurone layer. In contrast, Zn and Cu localization differed from that of P, as they were most abundant in the endosperm adjacent to the aleurone layer (i.e., the subaleurone layer).

**Mineral Localization in Mature Seeds**

The distribution patterns of minerals in mature seeds were determined by μ-XRF analysis (Fig. 7). Although the concentrations of each element in the accumulation area increased after 25 DAF (Fig. 5), the localization patterns of these elements did not change markedly (Fig. 7A). P, Ca, K, and Fe accumulated mainly in the aleurone layer. However, Zn and Cu broadly accumulated from the aleurone layer to the inner endosperm. Figure 7B shows the fine elemental maps around the aleurone layer on the ventral side of the seeds as determined by 1-μm step-size analysis. As in the case of developing seeds, the distributions of P, Ca, K, and Fe in the mature seed were similar. The localization sites of these elements were mostly restricted to the aleurone layer. In contrast, Zn and Cu localization differed from that of P, as they were most abundant in the endosperm adjacent to the aleurone layer (i.e., the subaleurone layer).

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both immature and mature seeds (Figs. 5 and 7). Using ion chromatography, Pi content was determined to be low (0.2–0.3 mg g⁻¹), and inositol phosphates other than InsP₆ were not detected in seeds throughout their development. The InsP₆ content of the outer endosperm increased rapidly from 10 to 25 DAF (Fig. 3) as P content increased (Fig. 1). Previously, we demonstrated that phytate-containing particles called globoids were present only in the aleurone layer, not in the starchy endosperm, during rice seed development (Yoshida et al., 1999). These results indicate that the majority of P is localized in the aleurone layer as phytic acid throughout seed development. Because translocation of P from vegetative source organs to seeds is active before 21 DAF (Fig. 1), the translocated P is likely converted immediately to InsP₆.

OsIPK1 catalyzes the final step of phytic acid biosynthesis from inositol 1,3,4,5,6-pentakisphosphate to InsP₆ in rice, and its transcript has been abundant in seeds from 5 to 28 DAF (Suzuki et al., 2007). In this study, OsIPK1 mRNAs and OsIPK1 proteins were detected not only in the embryos and the aleurone layers, in which the active phytate accumulation was observed, but also in the endosperms. This strongly suggests that phytic acid biosynthesis also occurs in endosperm cells. The concentration of InsP₆ in the inner endosperm did not increase, remaining at low levels during seed development from 7 to 30 DAF (Fig. 3). Therefore, InsP₆ synthesized in inner endosperm cells may be transported immediately to the aleurone layer, and/or InsP₆ synthesis in the endosperm may be restricted to the subaleurone layer.

Using μ-XRF analysis with a 1-μm step size, the subcellular localizations of the elements could be determined. This high-resolution elemental map showed the precise positions of the elements and whether they occupied the same sites. Because the majority of P in the aleurone layer may exist as InsP₆ and InsP₆ acts as a strong chelator of metal elements, the elements that were detected in the same place as P in the aleurone layer may exist in the form of a phytate salt, making it possible to deduce which elements bind mainly to InsP₆. Ca, K, and Fe were restricted mainly to the aleurone layer, and they colocalized with P in both developing and mature seeds. This strongly suggests that Ca, K, and Fe bind to InsP₆ in the aleurone layer. In contrast, Zn is more loosely bound to InsP₆ because it is localized not only in the aleurone layer but also in the inner endosperm to a considerable extent in immature seeds, and this tendency is stronger in mature seeds. This suggests that Zn binds not only to InsP₆ but also to other compounds. Recently, Persson et al. (2009) reported simultaneous detection of Fe, Zn, S, and P in barley (Hordeum vulgare) grain using size-exclusion chromatography and ICP-mass spectrometry. Fe and Zn were shown to have a different complex form in barley grain tissues, with Fe associated mainly with P (InsP₆) and Zn bound mainly to sulfur (peptides). This conclusion is consistent with our findings. Further research is required to determine the Zn-binding complex during seed development.

Cu localization differed from that of P at almost all seed developmental stages (Figs. 6 and 7), suggesting that Cu binds to a compound other than InsP₆. We suggest that translocated Cu passes rapidly through the aleurone layer to the endosperm tissues without being captured by InsP₆ in the aleurone layer. The concentration of Cu in seeds is too low to detect using the general method. By means of highly sensitive μ-XRF analysis at Super Photon Ring-8, we can, to our knowledge for the first time, suggest that Cu exists in a storage form other than phytate. In vegetative tissues,
Cu accumulates mainly in chloroplasts and is associated with metalloproteins such as plastocyanin. In spite of the essential role of Cu in plants, Cu ions are highly toxic at elevated concentrations. Therefore, excess cytoplasmic Cu is considered to bind to a detoxification peptide, phytochelatin (Epstein and Bloom, 2004). Further studies are needed to determine whether phytochelatin forms a complex with Cu in seed tissues.

In a study of the solubility and relative stability of various InsP₆-metal complexes using potentiometric titration analysis, Vohra et al. (1965) reported that the order of stability was Cu > Zn > nickel > cobalt > Mn > Fe > Ca. Persson et al. (1998) indicated Cu > Zn > cadmium, whereas Maddaiah et al. (1964), using a different titration method, reported the stability order as Zn > Cu > cobalt > Mn > Ca. However, in these studies, Cu and Zn may have had a high affinity for InsP₆. In contrast to these in vitro results, in vivo InsP₆ is stored mainly with Mg and K in rice and corn (Zea mays) seeds (Ogawa et al., 1975; Lin et al., 2005) and with Mg, K, and Ca in wheat (Triticum aestivum) seeds (Bohn et al., 2007). In this study, μ-XRF analysis at a 1-μm step size also indicated that Ca, K, and Fe bind strongly to phytic acid, Zn binds loosely, and Cu scarcely binds to phytic acid. The high affinity of Cu and Zn for InsP₆ that was found in vitro does not coincide with that observed in the in vivo system.

Nicotianamine (NA) is a chelator of metals and is ubiquitous in higher plants (Curie et al., 2009). The major role proposed for NA is that of a facilitator of long-distance transport of metals in the phloem, and it also has an important role in maintaining metal homeostasis. Fe, Zn, and Cu are likely present as their NA complexes in the phloem (von Wieren et al., 1999). Lee et al. (2009) reported that activation of one of the NA synthase genes, OsNAS3, resulted in a significant elevation of Fe, Zn, and Cu contents in rice seeds as well as elevation of NA levels in plants. In contrast, disruption of OsNAS3 caused the Fe, Zn, and Cu contents of seeds to decrease. These findings suggest that the transport of Fe, Zn, and Cu from source organs into seeds is in the form of NA complexes. After transport into seeds, the elements show the various movements and distributions found in our study. Fe remains primarily in the aleurone layer on the dorsal side. Cu moves immediately into the inner endosperm. Zn stays near the aleurone layer and moves more deeply into the inner endosperm. It has been suggested that Fe and Zn in seeds bind mainly InsP₆ and peptides, respectively (Persson et al., 2009). Whether the different movements and distributions of Fe, Zn, and Cu are reflected in differences in the complexes formed by these elements remains to be elucidated.

In this study, Ca content in developing seeds was nearly constant between 5 and 28 DAF (Fig. 1). This is greatly different from the accumulation patterns of the other elements. Ca is considered to move preferentially in xylem and is generally immobile in phloem (Epstein and Bloom, 2004). This is related to the symplastic nature of phloem. The constant Ca contents during seed development suggest that translocation of Ca into a seed via the phloem may be very limited.

Intensive accumulation of seed storage reserves is generally observed during the middle to late stages of seed development (Goldberg et al., 1989). For example, the major storage proteins of rice, glutelins and prolamins, are detected after 10 DAF (Li and Okita, 1993). The onset of phytic acid biosynthesis is unusually early: globoids, phytate-containing particles, have been observed as early as 4 DAF (Yoshida et al., 1999). In this study, we detected active phytic acid accumulation during early seed development, and the concentrations of Pi in seeds were maintained at low levels in both the inner and outer endosperm, which contains the aleurone layer, whereas translocation of P from the vegetative organ into seeds was quite active until 14 DAF (Fig. 1). The accumulation of phytate during the very early stages of seed development must play a role in the developmental process. Plausibly, Pi homeostasis in seed cells is needed throughout seed development, as it is in the vegetative organs. ADP-Glc pyrophosphorylase is a key regulatory enzyme in the biosynthesis of starch (Preiss, 1982). ADP-Glc pyrophosphorylase is allosterically inhibited by Pi in the plant cell (Plaxton and Preiss, 1987). To maintain high starch biosynthesis in endosperm cells throughout seed development, the concentrations of Pi in seed cells must be maintained at low levels. Phytic acid may support active starch synthesis by acting as a reservoir of Pi. Another possible role of phytic acid in early seed development is to protect the seed from oxidative stress. A highly reactive hydroxyl radical is generated from hydrogen peroxide in the presence of Fe by a Fenton or Fenton-like reaction (Prousek, 2007). Reactive species are powerfully toxic oxidants and can damage biomolecules, such as DNA, proteins, and lipids. Phytic acid is one of the natural strong iron chelators and, therefore, may protect seed cells from oxidative stress during seed development. Indeed, Fe was most abundant in the aleurone layer on the dorsal side, which is adjacent to the translocated site (Fig. 5). We suggest that the translocated Fe is immediately captured by phytic acid in the aleurone layer and is prevented from translocating to another location in the seed.

MATERIALS AND METHODS

Plant Materials

Rice (Oryza sativa var japonica, ‘Kitaake’ and ‘Nipponbare’) plants were grown in a greenhouse under natural light in summer. We used only superior carpyoses, as growth rates differ between superior and inferior carpyoses and seed growth in inferior carpyoses is retarded (Kuwano et al., 2009).

ICP-OES Analysis

Immature seeds of Kitaake at 5, 7, 14, 24, and 28 DAF were dehusked and freeze-dried. The freeze-dried samples (90–160 mg) were weighed into a glass tube and digested with 500 μL of HNO₃ for 12 h at 80°C on a heat block. Samples were then dissolved with 100 μL of 30% (w/v) hydrogen peroxide and diluted with 0.08 n HNO₃. The concentrations of P, Ca, K, Fe, Zn, and Cu in the acid-digested materials were determined using an ICP-OES apparatus.

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Distribution Changes in Metals of Developing Seeds

X-ray fluorescence intensity in the μ-XRF analysis is directly proportional to the concentration of the element (Terada et al., 2010). The analysis was repeated at least three times using three independent seeds at the same developmental stage.

Sequence data for OsIPK1 can be found in the Rice Annotation Project Database libraries under accession number Osa04g0661200.

Supplemental Data

The following materials are available in the online version of this article.

**Supplemental Figure S1** Changes in P, Ca, K, Fe, Zn, and Cu concentrations on the basis of seed fresh weight during seed development from 5 to 28 DAF.

**LITERATURE CITED**

We are grateful to Mr. W. Hosokawa, Mr. G. Kataoka, Mr. H. Kusuda, and Mr. D. Shima for technical assistance with the μ-XRF analysis. We thank the members of Laboratory of Plant Molecular Genetics at the University of Tokyo for comments and participation in discussions.

Received September 4, 2012; accepted October 17, 2012; published October 22, 2012.

**RT-PCR Analysis**

For RT-PCR analysis, total RNA was prepared from tissues of immature seeds of Nipponbare at 10 DAF. The method of separating the aleurone layer, embryo, and starchy endosperm from seeds at 10 DAF has been described previously (Kuwano et al., 2011). Total RNA was extracted using the RNeasy Plant Mini Kit (Qiagen). Genomic DNA was eliminated by treatment with DNase I, and then the RNA was purified using the RNeasy Micro Kit (Qiagen). RT-PCR amplification was performed using the One-Step RT-PCR Kit (Qiagen). The Actin gene was used as the control. The gene-specific primer pairs were 5′-CGTCCTGTGTAATCTGGTG-3′ and 5′-AATTTGCTCTGTCTGAG-3′ for OsIPK1 and 5′-GAACACGCTGGAAGCATA-3′ and 5′-CCATTGGAAGTCGAGAG-3′ for the Actin gene.

**Immunoblot Analysis**

For immunoblot analysis, the separated aleurone layer, embryo, and starchy endosperm from immature seeds of Nipponbare at 15 DAF were used. Samples were brushed with Multi-Beads Shockers and the powder of samples was defatted with acetone. Then, proteins were extracted from the defatted samples sequentially with solutions A (1 mM EDTA, 1% Triton X-100, 50 mM β-mercaptoethanol, and 10 mM Tris-HCl, pH 7.5) and B (4 M urea, 10% glycerol, 5% β-mercaptoethanol, 2% SDS, and 62.5 mM Tris-HCl, pH 6.5). The extracted proteins were separated by SDS-PAGE and transferred to Immob-Blot polyvinylidene difluoride membranes (Bio-Rad). Each lane contained the proteins extracted from an identical weight of defatted samples. The C-terminal half of the OsIPK1 protein (264 amino acids) produced in E. coli was used as a positive control. The OsIPK1 protein was visualized with goat anti-rabbit IgG alkaline phosphatase conjugate using X-phosphosphate membranes (Bio-Rad). Each lane contained the proteins extracted from an identical weight of defatted samples. The C-terminal half of the OsIPK1 protein (264 amino acids) produced in E. coli was used as a positive control. The OsIPK1 protein was visualized with goat anti-rabbit IgG alkaline phosphatase conjugate using X-phosphosphate membranes (Bio-Rad).

**μ-XRF Analysis**

cv Kitaake rice seeds were dehusked and freeze dried, cut with a microslicer (Dosaika EM) into 200-μm sections, and subjected to μ-XRF analysis. The μ-XRF imaging analysis was carried out at BL37XU of the Super Photon Ring-8 facility in Hyogo, Japan. Details of the μ-XRF system have been reported by Kamijo et al. (2003) and Hokura et al. (2006). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003). Elemental maps were obtained by scanning the samples with a 10.0-keV monochromatized beam. The monocromatized beam was focused by a Kirkpatrick-Baez mirror system into a 0.5-μm vertical direction microbeam at the entrance of the ion chromatography. The details of the extraction and ion chromatography methods have been described previously (Kuwano et al., 2003).
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