Use of Lipophilic Cations to Measure the Membrane Potential of Oat Leaf Protoplasts

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ABSTRACT

Uptake of the lipophilic cation triphenylmethylphosphonium into mesophyll protoplasts of oat (Avena sativa L. cv. "Garry") approaches equilibrium at 3 to 4 hours. The resulting external and internal concentrations are then used with the Nernst equation to obtain a membrane potential of −62 millivolts, inside negative. Potentials calculated in this manner are depolarized by adding 2 mM sodium azide and 50 μM carbonyl cyanide-m-chlorophenylhydrazone as well as by increasing the external proton and potassium concentrations. The depolarizations are qualitatively similar to those seen when oat mesophyll cells are measured in situ with microelectrodes. It is concluded that due to the lack of turgor and fragility of protoplasts, estimations of their membrane potential may be made more reliably, under some conditions, with lipophilic cations than with microelectrodes.

It is becoming increasingly evident that protoplasts are suitable material for studies of physiological properties of membranes. One reason is that free space determinations are usually unnecessary for these easily manipulated cells; thus, more quantiative estimates can be made for membrane-regulated events such as proton, mineral ion, and solute fluxes as well as for investigations into hormone transport and mode of action.

An important parameter for these studies is the membrane potential which is both a result of, and tends to regulate, transport of charged and uncharged species (6). Membrane potentials of +10 to +15 mV, inside positive, have been obtained for Acer pseudoplatanus protoplasts (5) and for protoplasts of six other species (14). These values were obtained by impaling cells with microelectrodes, and are strikingly different from membrane potentials of cells in intact tissues which are negative inside.

Even though the results for seven species of protoplasts were consistent (5, 14), the possibility exists that these cells cannot withstand microelectrode penetration as effectively as fully turgid cells in situ. Furthermore, fluxes of Rb⁺ and amino acids, which depend in part on a membrane potential which is inside negative (2, 8), showed similar characteristics in both intact tissues and protoplasts (ref. 13; and Rubinstein and Tattar, in preparation). I decided, therefore, to estimate protoplast membrane potential without using microelectrodes.

The procedure selected was described first for organelles by Grinius et al. (3) and has been used subsequently for bacteria (4, 15), Thermoplasma (9), and Chlorella (11) among other cell types. It involves the passive uptake of a lipophilic cation whose concentration ratio across the cell membrane at equilibrium is converted to membrane potential by the Nernst equation:

\[ E_m = 59 \log \frac{C_e}{C_i} \]

Activity coefficients are omitted and a temperature of 25 C is assumed. It will be shown that the radioactive lipophilic cation triphenylmethylphosphonium (TPMP⁺) is accumulated in oat leaf protoplasts against a concentration gradient and the membrane potential is estimated to be in the order of −62 mV, inside negative.

MATERIALS AND METHODS

Protoplasts were isolated and purified by a modification of the procedures of Kanai and Edwards (10). Leaves of 7-day-old light-grown oats (Avena sativa L. cv. "Garry") with the lower epidermis removed were plasmolysed for 15 min on a solution containing 0.6 M sorbitol, 29 mM sucrose, 20 mM MES-KOH (pH 5.5) and 5 mM MgCl₂. Cellulysin (Calbiochem) and DTT were then added to a final concentration of 0.5% (w/v) and 2 mM, respectively, for 2.5 hr at 33 C. The released protoplasts were filtered through 70-μm nylon mesh and centrifuged at 400 g for 2.5 min. The pellet was first resuspended in 0.6 M sorbitol, 29 mM sucrose, 50 mM HEPES-KOH (pH 8.0), 5 mM MgCl₂; then 1.2 ml was added to 10.8 ml of a mixture of PEG 6000 (Baker), Dextran T₂₀ (United States Biochemicals), Na-phosphate buffer (pH 7.5), and sorbitol [final concentrations: 5.5% (w/w), 2.5 mM, 400 mM, 0.6 M, respectively], and centrifuged at 300 g for 5 min. The purified protoplasts banding at the interface were removed and washed twice by centrifuging at 400 g for 60 sec in the buffer to be used for the uptake experiments. The cells were then stored overnight in the refrigerator. Cell counts were made with a hemocytometer.

Uptake was measured by adding an aliquot of protoplasts (1-5 x 10⁶) which had been at room temperature for 1 hr to a buffered solution (usually at pH 7.0) of sugars and ions [5 mM each of Tris, HEPES, and MES, 0.6 sorbitol, 29 mM sucrose, 1 mM glucose, 1 mM KH₂PO₄, 1 mM Ca(NO₃)₂, 0.25 mM MgSO₄ containing 3H-triphenylmethylphosphonium bromide (310 mCi/mmol; final concentration, 12.6 μM; supplied by New England Nuclear); the volume of incubation medium was usually 250 μl. After the desired incubation period at 25 C under fluorescent light of 200 ft-c, a 200-μl aliquot was removed and suspended in a calibrated tube containing 10 ml of cold incubation buffer. For the experiments in Table I, the tubes were poured onto 0.45-μm filters, the filters were then dried and placed into scintillation vials to which 4 ml of Scintisol (Isolab) had been added. For all other experiments, the tubes were centrifuged at 45 g for 6 to 8 min, the supernatant aspirated to 0.2 ml of the pellet, and, after adding 10 ml more cold incubation buffer, it was centrifuged again. A final aspiration to 0.2 ml was performed, 0.2 ml of buffer added, the tube vortexed, and 0.3 ml was placed in scintillation vials. Counts were converted to dpm with a quench curve for the isotope and the radioactivity associated with cells at zero time (about 3-5% of total uptake by 3 hr) was subtracted from each time point. Internal concentrations were calculated on the basis of a 40-μm diameter for the average protoplast.
Membrane potentials of leaf cells in situ were determined by inserting a 3 mM KCl-filled microelectrode (tip diameter less than 1 μm; resistance less than 25 megohm) into a peeled leaf segment. The signal was amplified and recorded as previously described (16). During these measurements, tissues were bathed in a flowing solution of 1 mM HEPES-KOH buffer (pH 6.5) containing 1 mM CaCl₂.

RESULTS

Two typical time courses for TPMP⁺ uptake are shown in Figure 1. The isotope approached equilibrium somewhat beyond 3 hr, but the curves were usually not flat even after 6 to 8 hr. Experiments were terminated at 3 to 4 hr, however, since protoplast numbers began to decrease sharply at that time. If the C₀ and Cₐ for TPMP⁺ are determined at 210 min for 11 different experiments, the average membrane potential using the Nernst equation is -62 ± 9 mv, inside negative. This value is thus an underestimate since uptake may still not be at equilibrium and the cells may become depolarized somewhat during the uptake period.

To see if the TPMP⁺ was free within the cells or was tightly bound to or dissolved in the cell components, protoplasts were osmotically broken and filtered before or after the uptake period (Table I). All of the radioisotope within the protoplast is probably not tightly bound since counts associated with cells ruptured at the start of the uptake period were not significantly different from counts associated with cells ruptured 210 min later.

External concentrations of H⁺ and K⁺ were altered to see if changes in Eₘ which are known to result (6) could be detected by measuring concentrations of TPMP⁺ close to equilibrium. A pH study (Table II) shows that the TPMP⁺-determined Eₘ is depolarized 12 mv as the pH decreased from 7.3 to 5.5. This depolarization can be compared with depolarizations determined with microelectrodes of 23 mv (from -148 mv to -125 mv) for oat mesophyll cells over the same pH span and 25 mv (from -120 to -95 mv) between pH 6.5 and 4.0 for oat coleoptile cells (2).

The addition of KCl depolarizes the TPMP⁺-determined Eₘ from -60 mv at 1 mM KCl to -44 mv at 30 mM KCl in the experiments shown in Table III. These data can be compared with a 58 mv depolarization (from -140 to -82 mv) seen with microelectrode methods when the KCl concentration around oat mesophyll cells is increased from 1 mM to 25 mM and a 60 mv depolarization for oat coleoptiles over the same concentration range (7).

Using microelectrodes, depolarizations of the Eₘ have been reported after the addition of respiratory poisons (6). In the experiment shown in Table IV, addition of 2 mM NaN₃ or 50 μM CCCP for 210 min decreased the TPMP⁺-determined Eₘ 29 mv and 33 mv, respectively. These results can be compared with a 30-mv depolarization (−146 mv to −116 mv) for 1 mM NaN₃ and with a 45-mv depolarization (−148 mv to −103 mv) for 50 μM CCCP measured by impaling mesophyll cells in situ with microelectrodes.

DISCUSSION

One criterion for using the Nernst equation with a passively accumulated ion to estimate the Eₘ is that the internal concentration is determined after equilibrium has been attained. While the time courses (Fig. 1) show that uptake does not always level off

![Time course of ³H]TPMP⁺ uptake into oat leaf protoplasts](https://www.plantphysiol.org/)

![Table I. Association of ³H]TPMP⁺ with the non-filterable portion of oat leaf protoplasts](https://www.plantphysiol.org/)

<table>
<thead>
<tr>
<th>Time after addition</th>
<th>Treatment</th>
<th>Isotope</th>
<th>Intact</th>
<th>Burst</th>
</tr>
</thead>
<tbody>
<tr>
<td>Min</td>
<td>dpm10⁻⁶</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>0.5</td>
<td>1.14</td>
<td>2.00</td>
<td></td>
<td></td>
</tr>
<tr>
<td>210</td>
<td>3.63</td>
<td>2.05</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

![Table II. Effect of pH on TPMP⁺ uptake into oat leaf protoplasts](https://www.plantphysiol.org/)

<table>
<thead>
<tr>
<th>pH</th>
<th>C₀/Cₐ</th>
<th>Eₘ</th>
</tr>
</thead>
<tbody>
<tr>
<td>7.3</td>
<td>11.7</td>
<td>-63</td>
</tr>
<tr>
<td>6.3</td>
<td>10.0</td>
<td>-59</td>
</tr>
<tr>
<td>5.5</td>
<td>7.3</td>
<td>-51</td>
</tr>
</tbody>
</table>

![Table III. Effect of KCl concentration on TPMP⁺ uptake into oat leaf protoplasts](https://www.plantphysiol.org/)

<table>
<thead>
<tr>
<th>KCl added (mM)</th>
<th>C₀/Cₐ</th>
<th>Eₘ</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>10.2</td>
<td>-60</td>
</tr>
<tr>
<td>10</td>
<td>7.5</td>
<td>-52</td>
</tr>
<tr>
<td>30</td>
<td>5.6</td>
<td>-44</td>
</tr>
<tr>
<td>100</td>
<td>3.7</td>
<td>-33</td>
</tr>
</tbody>
</table>

![Table IV. Effect of respiratory poisons on TPMP⁺ uptake into oat leaf protoplasts](https://www.plantphysiol.org/)

<table>
<thead>
<tr>
<th>Addition</th>
<th>C₀/Cₐ</th>
<th>Eₘ</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>15.3</td>
<td>-70</td>
</tr>
<tr>
<td>2mM NaN₃</td>
<td>5.0</td>
<td>-61</td>
</tr>
<tr>
<td>50μM CCCP</td>
<td>4.2</td>
<td>-37</td>
</tr>
</tbody>
</table>
Plant reasonbly deteriorate related concentration regarding relative negative inside erably stimulatory treatments. 

Another criterion for the use of TPMP\(^+\) is that the cation is distributed uniformly within the cell. It appears that the cation is not bound to cellular components (Table I), but a uniform distribution within a multicompartmented structure such as a protoplast is unlikely; it must be assumed, therefore, that the value for internal concentration of TPMP\(^+\) represents the average of all compartments and is largely that of the vacuole which occupies over 90% of cell volume. Similar problems regarding the identity of the membrane across which potential measurements are made also plague investigations using microelectrodes.

Compared to the microelectrode-determined \(E_m\) of mesophyll cells in situ (\(-140\) to \(-150\) mv), protoplasts at \(-62\) mv are depolarized, but they are still electronegative with respect to the external solution. An \(E_m\) with the inside negative is consistent with predictions made from uptake studies of Rb\(^+\) (13) and \(\alpha\)-amino-isobutyric acid (Rubinstein and Tattar, in preparation), but is opposed to literature values showing the inside of the protoplast to be electropositive compared to the outside (5, 14). This contradiction may be related to the method of measurement, i.e. the insertion of a microelectrode. It is possible that protoplast turgor pressure, which is near zero, changes the elasticity of the plasma membrane such that it makes an incomplete seal around the microelectrode; if the microelectrode tip were in the vacuole, the \(E_m\) would then be measured across the tonoplast. Positive potentials have been recorded for isolated vacuoles (12).

One disadvantage for the use of lipophilic cations to measure protoplast \(E_m\) is the length of time necessary to reach equilibrium. This has been obviated by Komor and Tanner (11) who plotted a calibration curve of uptake rate versus \(E_m\) for a range of potentials. Another way to measure \(E_m\) at shorter time intervals would be the flow dialysis method described by Ramos et al. (15). Finally, estimates of \(E_m\) using TPMP\(^+\) could be made at shorter times by measuring influx (\(J_i\)) and efflux (\(J_o\)) from preloaded protoplasts. These values could then be used in the Usling-Teorell flux-ratio equation which can be written:

\[
E_m = 59 \log \frac{C_o - J_o}{C_i - J_i}
\]

The chief advantage of measuring protoplast \(E_m\) with lipophilic cations rests on the comparative harmlessness of the method. Oat leaf protoplasts are extremely labile (1) and may not be able to survive the trauma of penetration by microelectrodes. Even if the absolute values of \(E_m\) cannot be determined accurately because of the length of time for the cation to reach equilibrium, it may still be possible to study the qualitative effects of various treatments on \(E_m\).

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LITERATURE CITED

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