ABSTRACT

The amination of α-ketoglutarate (α-KG) by NADH-glutamate dehydrogenase (GDH) obtained from Sephadex G-75 treated crude extracts from shoots of 5-day-old seedlings was stimulated by the addition of Ca²⁺. The NADH-GDH purified 161-fold with ammonium sulfate, DEAE-Toyopearl, and Sephadex G-200 was also activated by Ca²⁺ in the presence of 160 micromolar NADH. However, with 10 micromolar NADH, Ca²⁺ had no effect on the NADH-GDH activity. The deamination reaction (NAD-GDH) was not influenced by the addition of Ca²⁺.

About 25% of the NADH-GDH activity was solubilized from purified mitochondria after a simple osmotic shock treatment, whereas the remaining 75% of the activity was associated with the mitochondrial membrane fraction. When the lysed mitochondria, mitochondrial matrix, or mitochondrial membrane fraction was used as the source of NADH-GDH, Ca²⁺ had little effect on its activity. The mitochondrial fraction contained about 155 nanomoles Ca per milligram of mitochondrial protein, suggesting that the NADH-GDH in the mitochondria is already in an activated form with regard Ca²⁺. In a simulated in vitro system using concentrations of 6.4 millimolar NAD, 0.21 millimolar NADH, 5 millimolar α-KG, and 5 millimolar glutamate thought to occur in the mitochondria, together with 1 millimolar Ca²⁺, 10 and 50 millimolar NH₄⁺, and purified enzyme, the equilibrium of GDH was in the direction of glutamate formation.

NADH-GDH is easily solubilized from mitochondria from roots (30), shoots (4, 25), and seeds (28) of various plants. Although it is the only mitochondrial enzyme with the potential for the assimilation of NH₄⁺, it has been suggested by Miflin and Lea (21) that the NH₄⁺ released by the conversion of glycine to serine in the mitochondria (3) is reasimilated via GS in the cytosol. They have also suggested that the principal function of mitochondrial GDH is the oxidation of glutamate (21). However, a part of intramitochondrially generated NH₄⁺ from glycine is incorporated into glutamate by mitochondria isolated from pea shoots (11). Chou and Splisttoesser (7), Joy (14), and recently Furuhashi and Takahashi (10) have shown that NADH-GDH (amination reaction), but not NAD-GDH (deamination reaction), is activated by Ca²⁺ in vitro. According to the work by Bowman et al. (4), NAD is the only detectable pyridine nucleotide in mung bean mitochondria. Thus, concentrations of Ca²⁺ and NADH in the mitochondria could be important in the regulation of NADH-GDH.

The purpose of the present study was to characterize the activation of NADH-GDH by Ca²⁺ and to estimate concentrations of Ca²⁺ and NADH for simulation of the GDH reaction in mitochondria isolated from corn shoots.

MATERIALS AND METHODS

Plant Materials. Corn seeds (Zea mays L. var W64A × W182E) were planted in vermiculite and seedlings were grown in a Toshiba Cold Chain growth chamber, model TGS-13L, with a 12-h light period for 5 d. Light (about 490 μE/m²-s) was supplied by Toshiba Yoko lamps DA400 (metal halide lamp) and fluorescent lamps, and the temperature was maintained at 27°C. The plants were watered every morning with a nutrient medium containing 1 mM Ca²⁺ as described previously (18).

Isolation of Mitochondria by Density Gradient Centrifugation. All reagents and buffer solutions were prepared with double glass-distilled H₂O to minimize Ca²⁺ contamination, and all operations were carried out at 4°C. Whole shoots of 5-d seedlings were harvested, washed in water, blotted dry with filter paper, and weighed. The shoots (40 to 60 g in fresh weight) were chopped with razor blades in extraction buffer consisting of 0.4 M mannitol, 0.1 mM Hepes-KOH buffer (pH 7.5), 1 mM EDTA, 0.1% (w/v) BSA, and 0.6% (w/v) insoluble PVP, at a ratio of 1 g fresh weight per 3 ml of buffer. The chopped materials were ground with mortar and pestle.

The mitochondrial fraction was isolated by modification of the method described by Nishimura et al. (26) as described previously by us (32). The purified mitochondria after Percoll discontinuous density gradient were able to oxidize both malate and succinate with a high P/O and RCR ratios and appear to have about 92% of the outer membrane intact (32). Our preparations were almost free of Chl (chloroplast) or catalase (microbody) contamination. These criteria were used by Nishimura et al. for assessing mitochondrial purity (26).

Preparation of Submitochondrial Fractions. The purified mitochondrial pellet was treated hypotonically using 1.5 ml of 20 mM Hepes-KOH buffer (pH 7.5). The suspension was mixed with a Teflon-glass homogenizer and incubated in an ice bath for 30 min. The suspension (1 ml) was then centrifuged in the ultracentrifuge equipped with an SW 50-1 rotor at 150,000 g for 1 h. The pellet, resuspended in 1 ml of the same buffer, was designated the mitochondrial membrane fraction. The supernatant remaining above the pellet consisted of the soluble matrix.
of the mitochondria.

Purification of NADH-GDH from the Crude Mitochondrial Fraction. The crude mitochondrial pellet prepared from 179 g fresh weight of the corn shoots was suspended in about 90 ml of 0.1 M Tris-HCl buffer (pH 8.5) containing 0.1 mM EGTA and 1 mM 2-ME. The suspension was frozen and thawed twice, placed in a salt-ice mixture, and then sonicated with a Branson sonifier, model 200, operating at power setting 5 on a 50% duty cycle for 2 min. Following the disruption treatment, the suspension was centrifuged at 20,000g for 30 min and the supernatant fraction was designated the crude mitochondrial extract. The extract was subjected to 30 to 60% (NH₄)₂SO₄ fractionation. The resulting pellet was dissolved in 0.1 M Tris-HCl buffer (pH 8.5) containing 0.1 mM EGTA and 1 mM 2-ME and then dialyzed against the same buffer. The dialyzed preparation was centrifuged and the mitochondrial supernatant fraction was loaded onto a column (1.6 × 10 cm) of DEAE-Toyopearl, equilibrated with 50 mM Tris-HCl buffer (pH 8.5) containing 5 mM α-KG, 1 mM 2-ME, and 0.1 mM EGTA. The column was washed with the same buffer and the NADH-GDH was eluted with a linear gradient of KCl (0 to 0.5 M). The NADH-GDH activity was recovered in the portion of the eluent containing approximately 0.12 M KCl. Proteins were then sedimented with 70% (NH₄)₂SO₄, and the enzyme was further purified by gel filtration chromatography on Sephadex G-200 column (1.6 × 90 cm) equilibrated with 50 mM Tris-HCl buffer (pH 8.5) containing 0.1 M KCl and 1 mM 2-ME. The NADH-GDH activity was observed as a single peak near the void volume of the column, and was free of contaminating EGTA.

When NADH-GDH activity was assayed without purification, the procedures of enzyme extraction from the corn shoots and desalting with Sephadex G-75 were the same as those described by Oaks et al. (27).

Enzyme Assays and Analytical Methods. Enzyme activities were assayed spectrophotometrically in a double-beam spectrophotometer (Shimadzu, model UV-150-02) at 30°C. NADH-GDH was assayed routinely in 0.2 M Tris-HCl buffer (pH 8.0), containing 0.266 mM (NH₄)₂SO₄, 0.16 mM NADH, and 5 mM α-KG with or without 1 mM CaCl₂ as described previously (27). However, in critical experiments (NH₄)₂SO₄ was used in a concentration range of 2 to 100 mM. NADH-GDH was measured by a modification of the method described by Furuhashi and Takahashi (10). The assay system contained 0.1 M Tris-HCl buffer (pH 8.5), 3 mM NAD, and 50 mM glutamate with or without 1 mM CaCl₂. Succinate:Cyt c reductase activity was assayed by following reduction of Cyt c as described by Douce et al. (9). Malate dehydrogenase was assayed by following the oxidation of NADH as described by Priestley and Brunsma (28). Fumarase was assayed by coupling the reaction with malic enzyme and following the reduction of NADP as described by Hatch (12).

The Ca content in the isolated mitochondrial fraction was determined with an atomic absorption/flame emission spectrophotometer (Shimadzu model AA646, equipped with a Graphite furnace atomizer, model GFA-3). Protein content was determined by the method of Lowry et al. (16).

Extraction and Determination of Pyridine Nucleotides. Pyridine nucleotide content was determined by the cycling assay method described by Matsumura and Miyachi (19). During extraction and isolation of mitochondria, 10 μM rotenone and 5 μM antimycin A were included in all the buffer solutions and in the Percoll gradient solutions. The final mitochondrial pellet after the Percoll density gradient was dissolved in 1.1 ml of 0.1 M Bicine-NaOH buffer (pH 8.0). The mitochondrial suspension (0.5 ml) was then transferred to preheated NaOH or HCl solutions which had a final concentration of 0.1 N. After an incubation at 100°C for 2 min, the suspension was cooled and centrifuged. NADH is specifically extracted with an alkaline treatment, while NAD is extracted with an acid treatment (19). Each extract was neutralized by adding an appropriate amount of NaOH or HCl. The extract (50 to 200 μl) was then incubated in brown test tubes in a reaction system containing 0.1 mM Bicine-NaOH buffer (pH 8.0), 4 mM EDTA, 0.42 mM MTT, 1.676 mM PES, and 0.5 mM ethanol for 5 min at 37°C. The total reaction volume was 1.0 ml. The reaction was started by adding 10 units of alcohol dehydrogenase and was allowed to run for 30 min at 37°C. The reaction was terminated by adding 12 mM iodoacetate and the absorbance was measured at 570 nm. Amounts of NADH were calculated from the standard curves with authentic NAD(H).

A blank test without sample or NAD(H) was also carried out.

Source of Chemicals. Bicine, Heps, Mes, Tris, sucrose, manitol, PES, Na-glutamate, and rotenone were all purchased from Nakarai Chemicals, Ltd., Japan and were of a specially prepared reagent grade. Fatty acid-free BSA, horse heart Cyt c (type III), yeast alcohol dehydrogenase free of NAD(H), chicken liver malic enzyme, and MTT were from Sigma; antimycin A, from Boehringer; Percoll and Sephadex, from Pharmacia. NAD, NADH, NADPH, NADP were purchased from Kojin Co. Ltd., Japan and DEAE-Toyopearl was from Toyo Soda MFG Co. Ltd., Japan. All other chemicals were of the highest quality available.

RESULTS

Localization of NADH-GDH in Mitochondria Isolated from Corn Shoots. NADH-GDH is relatively easily solubilized from plant mitochondria and hence the enzyme is thought to be localized in the soluble matrix of mitochondria (4, 28). However, the method of disrupting the mitochondria in those reports (4, 28) involved a relatively vigorous treatment, such as freezing and thawing followed by sonication. To demonstrate the distribution of NADH-GDH in corn mitochondria, we treated mitochondria, purified by centrifugation through a Percoll density gradient, with osmotic shock, a relatively gentle disruption method. The mitochondrial matrix and membrane fractions were then separated by centrifugation (Table I). Most of the fumarase activity (a matrix marker) and succinate:Cyt c reductase (a membrane marker) were located in the expected fractions (29). NADH-GDH activity of catalytic dehydrogenase, which was thought to be in the mitochondrial matrix (4, 28) were, however, found primarily in the membrane fraction (75%). When the mitochondria were disrupted by harsher treatments (freezing and thawing followed by sonication), about 85% of the NADH-GDH activity was

| Table 1. Localization of NADH-GDH in Mitochondria Isolated from Corn Shoots |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| Fraction        | NADH-GDH        | Malate Dehydrogenase | Fumarase | Succinate:Cyt c Reductase |
| Lysed mitochondrial fraction | 0.476 | 2.03 | 0.024 | 0.116 |
| % of activity assayed with the lysed mitochondria | Mitochondrial matrix | 25.5 | 22.6 | 70.4 | 1.3 |
| Mitochondrial membrane | 77.8 | 74.8 | 7.4 | 89.2 |

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Table II. Activation of NADH-GDH by Ca²⁺

The amination reaction of GDH was measured using a crude preparation desalted on Sephadex G-75 column (27), a 16-fold purified fraction, a lysed mitochondrial fraction, a mitochondrial matrix fraction, and a mitochondrial membrane fraction from shoots of 5-d-old corn. The NADH-GDH activity was measured in the presence or absence of 1 mM CaCl₂ in the standard assay system containing 0.2 M Tris-HCl buffer (pH 8.0), 0.266 mM (NH₄)₂SO₄, 0.16 mM NADH, and 5 mM α-KG.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Desalted Crude Fraction</th>
<th>Purified enzyme</th>
<th>Mitochondrial fractions</th>
<th>Lysed Matrix Membrane</th>
</tr>
</thead>
<tbody>
<tr>
<td>No addition</td>
<td>0.01</td>
<td>1.40</td>
<td>0.68</td>
<td>0.20</td>
</tr>
<tr>
<td>Ca²⁺ (1 mM)</td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>

Ca²⁺ on the amination reaction was tested using fixed concentrations of 200 mM NH₄⁺, 5 mM α-KG, and 10 or 160 μM NADH. In the presence of 160 μM NADH, the activation curve was sigmoidal with significant activation seen with 1 mM Ca²⁺ and maximal activation with 45 μM Ca²⁺ (Fig. 1). The NADH-GDH was thus activated about 9-fold by Ca²⁺. Higher concentrations of Ca²⁺, up to 10 mM, maintained the activity at the saturated level (results not shown). Addition of 2 mM EGTA completely reversed the activation of the enzyme by 1 mM Ca²⁺. When the NADH levels were reduced to 10 μM NADH, Ca²⁺ had no effect on the amination reaction. The oxidative deamination reaction of GDH (NAD-GDH) was not affected by addition of Ca²⁺.

For example, with purified GDH, 2.58 and 2.41 μmol NAD were reduced/min/mg protein in the absence and presence of 1 mM Ca²⁺, respectively.

In animal cells many of the regulatory processes linked to Ca²⁺ are mediated through their interactions with high-affinity Ca²⁺-binding proteins, such as calmodulin (6). Calmodulin also exists in plant cells (17) but its function is not yet known, except perhaps in chloroplasts (13, 23) and the plasma membrane-enriched microsomal fraction (8). Neither bovine brain calmodulin nor the calmodulin antagonist, trifluoperazine, influenced the activity of purified or mitochondrial derived GDH (data not shown).

Saturation curves for the three substrates of NADH-GDH showed the normal Michaelis-Menten kinetics in the presence of 1 mM Ca²⁺ (Fig. 2). Apparent Kₘ values of 63 μM for NADH and 2.22 mM for α-KG were calculated from the curves. The apparent Kₘ for NH₄⁺ decreased when the concentration of NADH was lowered; it was 12.5 mM in the presence of 160 μM NADH and 4.76 mM with 10 μM NADH. Thus, the corn shoot-NADH-GDH has kinetic properties similar to the enzyme from Medicago sativa (24). With low concentrations of NADH, the apparent Vₘₐₓ for NH₄⁺ also decreased. In the absence of Ca²⁺, a marked substrate inhibition was observed with the individual substrates and, hence, the Kₘ value for each could not be measured.

Estimation of Concentrations of Ca²⁺, Pyridine Nucleotides, NH₄⁺, and Glutamate in Mitochondria Isolated from Corn Shoots. The concentrations of Ca²⁺ in purified mitochondria

FIG. 1. Effect of Ca²⁺ on the amination reaction of GDH purified from corn shoots. The assay mixture contained 10 (O) or 160 (●) μM NADH, 5 mM α-KG, 100 mM (NH₄)₂SO₄, and 1.2 μg of the 16-fold purified enzyme in 0.2 M Tris-HCl buffer (pH 8.0). The NADH-GDH activity was 3.38 and 1.69 nmol NAD oxidized-min⁻¹ with 10 and 160 μM NADH, respectively, in the absence of Ca²⁺. Effect of 2 mM EGTA was also tested in the presence of 160 μM NADH and 1 mM Ca²⁺ (▲).

FIG. 2. Substrate saturation curves of the purified NADH-GDH from corn shoots in the presence (●) or absence (O) of 1 mM Ca²⁺. A, NH₄⁺ with 10 μM NADH; B, NH₄⁺ with 160 μM NADH; C, NADH; and D, α-KG. In each system the other components were added at the concentrations used in the assay system as described in Figure 1.
Inhibitors of electron transfer chain, 10 μM rotenone and 5 μM antimycin A, were added to all buffers and Percoll gradient solutions during extraction and purification of mitochondria. The purified mitochondrial pellet was dissolved in a known volume of 0.1 M Bicine-NaOH buffer (pH 8.0) and the suspension was treated with either 0.1 N NaOH or 0.1 N HCl followed by boiling for 2 min for the extraction of NADH or NAD. After neutralization, the pyridine nucleotides were assayed using the cycling method in the presence of alcohol dehydrogenase (19).

<table>
<thead>
<tr>
<th>Experiment</th>
<th>Ca (nmol·mg⁻¹ mitochondrial protein)</th>
<th>NAD (nmol·mg⁻¹ mitochondrial protein)</th>
<th>NH₄⁺ (mm)</th>
<th>Glu (10⁻³ M)</th>
<th>Gly (10⁻³ M)</th>
<th>Ser (10⁻³ M)</th>
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<tr>
<td>I</td>
<td>162</td>
<td>7.22</td>
<td>0.14</td>
<td>5.03</td>
<td>1.21</td>
<td>2.68</td>
</tr>
<tr>
<td>II</td>
<td>149</td>
<td>5.51</td>
<td>0.28</td>
<td>3.64</td>
<td>0.80</td>
<td>1.07</td>
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<td></td>
<td></td>
<td></td>
<td>54</td>
<td>6.37</td>
<td>0.21</td>
<td>4.34</td>
</tr>
</tbody>
</table>

**DISCUSSION**

GDH purified from crude mitochondrial preparations of corn shoots is stimulated by Ca²⁺ over a concentration range of 1 μM to 10 mM, when the NADH concentration is in excess of the Kₘ for NADH. Estimations of the concentrations of Ca and NADH in the mitochondria suggest that NADH-GDH should be in the activated state. Hence NADH-GDH should be in a configuration to support the assimilation of NH₄⁺. In a model experiment using pure GDH and all the substrates for NADH- and NADH-GDH at the concentrations estimated to be present in the mitochondria, the equilibrium of the reaction was in the direction of glutamate formation. In recent experiments by Hartmann and Ehmke (11), approximately 14% of the NH₄⁺ released from glycine in mitochondria obtained from pea shoots was reassimilated within the mitochondria. This was interpreted by them to be insufficient to consider NADH-GDH as a significant point of entry of NH₄⁺. However, no mention was made of optimizing Ca and NADH in their system, hence even this low level of NH₄⁺ assimilation should be taken as an indication of the capacity of the mitochondria to assimilate NH₄⁺. According to the work by Bergman and Ericson (2), the release of NH₄⁺ from glycine occurs at a pH maximum at 8.1 in spinach leaf mitochondria. This pH is almost the same as the optimal pH of NADH-GDH reaction

**Fig. 3.** Simulation of equilibrium of GDH reaction in vitro using concentrations of potential substrates estimated to occur in mitochondria of corn shoots as a function of pH. The purified enzyme fraction (1.2 μg protein) was used as the source of enzyme. According to the estimation of substrates (Table III), the enzyme was incubated with 0.21 mM NADH, 6.4 mM NAD, 5 mM α-KG, 5 mM glutamate, 1 mM CaCl₂, and 10 (C) or 50 mM (O) of NH₄Cl in 0.2 M Tris-Mes buffer at different pH indicated. An oxidation of NADH was observed in all cases. The rate of amination reaction (C) and the deamination reaction (Δ) were also measured with the standard assay systems as described in “Materials and Methods,” except that the Tris-HCl buffers are replaced by the Tris-Mes buffer at different pHs indicated.

from corn shoots was estimated in order to determine its in vivo significance (Table III). NADH is also an important factor for the NADH-GDH activity and its activation by Ca²⁺ (Fig. 1), while NAD has been shown to be an inhibitor of the amination reaction (14, 24). Therefore, the concentration of both NAD and NADH were also determined in the purified mitochondria. Purified corn mitochondria contained 149 to 162 nmol Ca/mg protein. When intramitochondrial Ca distribution was determined, after submitochondrial fractionation following the gentle disruption with osmotic shock, about 35% of the total Ca was distributed in the mitochondrial matrix and the remaining 65% with the mitochondrial membrane. If one assumes a matrix water space of 1 μl/mg mitochondrial protein (4, 15), and the Ca detected in the matrix is all free, the Ca²⁺ concentration in the matrix can be calculated to be 52 to 56 mM. This, together with the fact that the addition of Ca²⁺ had little effect on the NADH-GDH in purified mitochondrial fractions (Table I), suggests that the mitochondrial enzyme is fully activated with respect to Ca²⁺. The purified mitochondria contained 0.21 nmol NADH and 6.37 nmol NAD/mg mitochondrial protein, respectively. Thus, the concentrations of NADH and NAD in the matrix should be 0.21 and 6.37 mM. The concentration calculated for NADH is higher than the apparent Kₘ for NADH obtained in vitro using the purified GDH in the presence of Ca²⁺ and, hence, an activation by Ca²⁺ would be expected to occur in vivo.

Levels of amino acids in the mitochondria were determined by reversed phase chromatography on an Altex Ultrasphere-ODS (5 μM) column as described by Winspear and Oaks (31). Levels of glycine or glutamate in corn mitochondria were 1.88 and 1.01 nmol/mg protein, respectively. NH₄⁺ levels were determined after diffusion of NH₄⁺ in Conway dishes by the method of McCullough (20). Values for NH₄⁺ range from 3.6 to 5.0 mM in corn mitochondria. If NH₄⁺ is as diffusible as people claim, then our values are low values (2, 11). Additions of NH₄Cl up to 30 mM had no effect on the respiration of malate. Thus, mitochondrial respiration is several orders of magnitude less sensitive to NH₄⁺ than is chloroplast electron transport (11, 21).

To estimate the equilibrium of the GDH reaction in corn mitochondria, purified GDH was incubated in the 0.2 M Tris-Mes buffer at a pH range from 6.0 to 9.0 with the estimated in vitro levels of substrate (Fig. 3). According to the results of Bowman et al. (4), mitochondria from mung bean contain both α-KG and glutamate in the millimolar range. From these calculations an assay system containing 0.21 mM NADH, 6.4 mM NAD, 5 mM α-KG, 5 mM glutamate, 1 mM Ca²⁺, and two levels of NH₄⁺ (10 and 50 mM) was tested by measuring oxidation of NADH or reduction of NAD. The NH₄⁺ levels represent concentrations above or below the Kₘ for GDH. With these assay conditions, the equilibrium of GDH was in the direction of amination at all pHs tested. As with the standard NADH-GDH assay, a pH optimum of 8.0 was observed. The deamination reaction rate increased with increasing pH over the pH range used. When various concentrations of NH₄⁺ (2 to 100 mM) were tested with the reaction system containing all of substrates of GDH at pH 8.0, the equilibrium of the reaction was still in the direction of glutamate production (results not shown).
either in the standard assay system or in the simulated assay system as shown in Figure 3. The conversion of glycine to serine is catalyzed by two enzymes, glycine decarboxylase and serine hydroxymethyltransferase, both of which are localized in the mitochondrial membrane (22). This conversion is also coupled to the formation of NADH (1). Our finding that the loose association of NADH-GDH with the mitochondrial membrane in corn shoots coupled with the generation of both NADH and NH₄⁺ in that fraction suggest that GDH could participate in the re-assimilation of NH₄⁺.

It has been proposed by Miflin and Lea (21) that GS should serve in prefered, due to GDH in the assimilation of NH₄⁺, because of its low Km for NH₄⁺. In their scheme, GOGAT allows for the formation of two glutamate molecules from one molecule each of glutamine and α-ketoglutarate. In normal metabolism, one glutamate molecule would be recycled through glutamine and the other would be available for other biosynthetic reactions. In photosynthesis, the two glutamate molecules would be required for the synthesis of two glycine molecules, and a third glutamate molecule from somewhere would be required for the regeneration of glutamine. Our results indicate that levels of NH₄⁺ in the mitochondria are in the 5 to 10 mm range, and that the mitochondria can tolerate these relatively high concentrations of NH₄⁺. At these levels of NH₄⁺, there should be a synthesis of glutamate in the mitochondria mediated by GDH. It is our contention that GDH, by virtue of its isolation in mitochondria, can compete successfully with GS for that NH₄⁺. A GDH fully saturated with Ca²⁺ and NADH may not be sufficient to assimilate all the NH₄⁺ released by the conversion of glycine to serine or, for that matter, produced by the action of nitrate reductase. However, its action would permit the net synthesis of glutamate, glutamine, and, subsequently, of other amino acids required to sustain both the GS/GOGAT cycle itself and the enhanced rate of protein synthesis seen in light-treated leaves (5).

**LITERATURE CITED**

10. Furuhashi, K., Sakaishi 1982 Control of glutamate dehydrogenase from green tobacco callus mitochondria by Ca²⁺ and pH. Plant Cell Physiol 23: 179-184