Heat Inducible Expression of a Chimeric Maize hsp70CAT Gene in Maize Protoplasts

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Received for publication June 6, 1988

ABSTRACT

The response of maize (Zea mays L.) protoplasts to high temperature stress was investigated. After isolation and electroporation, protoplasts were preincubated for 12 hours at 26°C then incubated for 6 hours at elevated temperatures. The pattern of polypeptides synthesized by these protoplasts during the last hour was monitored by in vito labeling with [35S]-methionine. Incubation at 40°C and 42°C resulted in the synthesis of polypeptides not detectable at 26°C. Introduction of a chimeric maize heat shock protein 70 promoter-chloramphenicol acetyltransferase coding region gene into protoplasts via electroporation resulted in the temperature-dependent induction of chloramphenicol acetyltransferase activity with maximal activity at 40°C. In the same protoplasts, a second chimeric gene, in which the firefly luciferase coding region was under the control of the 35S promoter from cauliflower mosaic virus, did not show an increase in expression after incubation at higher temperatures. Maize protoplasts provide a system to study molecular responses to high temperature stress.

The advantages of the use of plant protoplasts for biochemical and gene transfer studies have been widely demonstrated. The ability to obtain a homogeneous population of cells without the physical barrier of the cell wall has allowed the measurement of ion channels (23, 29), facilitated the isolation of subcellular components such as vacuoles (33), and led to the development of direct DNA-mediated gene transfer technologies (14, 24).

For physiological studies, the question remains whether protoplasts are a suitable experimental system. There are many cases where isolated protoplasts mimic the behavior of cells in situ. Oat aleurone protoplasts undergo several morphological changes, release acid phosphatase, and induce amylase mRNA (15) in response to applied gibberellic acid as was previously observed in intact aleurone layers. Cucumber cotyledon protoplasts resemble intact cotyledons in their chilling sensitivity (25). Maize protoplasts regulate the expression of the Adhl promoter in response to oxygen tension as do whole roots (16, 34).

There are other examples, however, where protoplast behavior does not correspond to the behavior of the organ or tissue from which the protoplasts were isolated. Tobacco mesophyll protoplasts do not synthesize small subunit of ribulose bisphosphate carboxylase (10, 11, 32). Barley leaf protoplasts do not accumulate abscisic acid as a response to decreasing osmotic potential, a phenomenon which is observed in intact leaves (20). Thus, it appears that protoplasts are not universally suitable for experiments in which their behavior must mimic the intact tissue; some responses are adversely affected, possibly because the protoplasts experience a loss of turgor, are incubated in high osmoticum, and/or are exposed to crude digestive enzyme preparations.

The response of plant cells to high temperature stress by the synthesis of a subset of proteins, called heat shock proteins, hsp4 (17), has been documented in undifferentiated plant cell cultures of soybean, tobacco (3), and maize (8). In addition, tobacco mesophyll protoplasts synthesized hsp4 (22). We were interested in using a protoplast system to analyze the expression of the maize hsp70 gene (26) during heat shock. The ability to transiently introduce genes into maize protoplasts via electroporation (13) made it possible to analyze the expression of the maize hsp70 gene in a homologous system. We show here that maize protoplasts, even after electroporation, are capable of a heat shock response and show a temperature dependent induction of an introduced maize hsp70-CAT chimeric gene.

MATERIALS AND METHODS

Materials. pCaMViuc was the gift of Jeff DeWet (Stanford University). pMON9508 was the gift of Dilip Shah (Monsanto Company). Maintenance and manipulation of the BMS suspension line and protoplasts were performed as described by Fromm et al. (12).

Construction of pphp70cat. pMON9508 was sequentially digested with PvuII and Bal31; the ends were filled in with T4 DNA polymerase, and the DNA was digested with EcoRI. The hsp70 promoter fragment was then ligated into pUC8 that had been digested with SmaI and EcoRI. A BamHI fragment containing the hsp70 promoter was ligated into the BamHI site of a derivative pUC8 plasmid containing the CAT coding region (1) and the 260 bp Sau3 A fragment containing the poly A addition site of the nopaline synthase gene (4) downstream of the BamHI site. The extent of the Bal31 deletion of the promoter fragment was determined by dideoxy sequencing (5).

Electroporation and Enzyme Assays. Protoplasts were electroporated in 1 mL batches in the presence of both pphp70cat and pCaMViuc, pooled together, and then re-aliquoted to minimize between sample variation. Electroporation conditions, prepara
tion of the protoplast extracts, and CAT assays were as described previously (12); the CAT assay included a 10 min incubation at 65°C (28). After separation by TLC, the ¹⁴C-chloramphenicol and acetylated products were localized by autoradiography, and the radioactive spots were quantitated individually by scintillation counting. The amount of acetylated ¹⁴C-chloramphenicol was divided by the total radioactivity in each lane to determine the percent conversion. Luciferase assays were performed as described in Callis et al. (6). Luciferase activity was monitored in a Monolight 2000 photometer; the values given represent integration of the luminosity for a 30 s period after addition of substrate.

**In Vivo Labeling.** Twelve h after preparation (control) or 12 h after preparation and electroporation, protoplasts were either maintained at 26°C or were incubated at various temperatures for 6 h. During the last hour, ³⁵S-methionine was added to the media to a final concentration of 20 μCi/ml. Protoplasts were then concentrated by centrifugation, washed one time in incubation media with 1 mM methionine, lysed in SDS buffer (18), and sonicated. Total incorporation into acid insoluble material was determined (21). Equal quantities of radioactivity were loaded on a 10% polyacrylamide SDS gel (18). After electrophoresis, the gel was prepared for fluorography by 2,5-diphenyloxazole treatment.

**RESULTS**

The effect of electroporation on the proteins synthesized by maize (*Zea mays* L.) protoplasts is shown in Figure 1. Protoplasts isolated from BMS suspension cells were electroporated and incubated for 18 h at 26°C, the last hour in the presence of ³⁵S-methionine. The profile of *in vivo* labeled proteins from these electroporated protoplasts (lane 2) is identical to the nonelectroporated control protoplasts (lane 1) which were incubated and labeled identically. The levels of acid precipitable counts were also similar (data not shown). This indicates that after 18 h of incubation, electroporation had not altered the ability of protoplasts to synthesize proteins nor the pattern of proteins synthesized.

Figure 1 also shows the effects of incubation at elevated temperatures on protoplast protein synthesis (lanes 3–6). Electroporated protoplasts were incubated for 12 h at 26°C and then placed at the various temperatures for 6 h. During the last hour of incubation ³⁵S-methionine was added. There was no detectable difference between the proteins synthesized at 26°C (lane 2) and at 30°C (lane 3). At 37°C (lane 4), in addition to the 26°C pattern of protein synthesis, there were barely detectable levels of additional polypeptides. These additional polypeptides were more prominently labeled at 40°C (lane 5) and at 42°C (lane 6) constituted the major polypeptides synthesized. The pattern of polypeptides synthesized at 42°C is very similar to that previously observed in heat shocked BMS cultured cells (8) and in heat shocked maize coleoptiles (27). The rate of protein synthesis was also dramatically affected by incubation at 42°C. On a per cell basis, the amount of acid insoluble counts at 42°C was 2% of that incorporated at 26°C (data not shown). Thus, despite the physical stresses of cell wall removal and electroporation, the protoplasts were still capable of responding to heat shock by increasing the rate of synthesis of a small set of hps.

Because protoplasts were capable of responding to high temperature stress, we asked whether expression of an extra-chromosomal hsp 70 gene would show regulated expression when introduced into protoplasts via electroporation. To assay for the induction of an hsp70 gene, an hsp70-CAT chimeric gene was constructed (Fig. 2). A DNA fragment from pMON9508 (26) containing approximately 1.1 kb upstream of the transcribed region for an hsp70 gene and all but 24 of the 107 nucleotides of the 5′ transcribed, untranslated leader was placed upstream

**Fig. 1.** *In vivo* labeling of BMS protoplasts. Protoplasts were labeled in vivo with ³⁵S-methionine for 1 h at 17°C after either isolation (lane 1) or isolation and electroporation (lanes 2–6). After a 12 h incubation, protoplasts were incubated for 6 h at the temperature indicated above each lane. After 18 h, the protoplasts were lysed, and equal amounts of acid insoluble radioactivity were loaded in each lane of a 10% polyacrylamide SDS gel. Following electrophoresis the polypeptides were visualized by fluorography.
HEAT SHOCK RESPONSE IN MAIZE PROTOPLASTS

![Diagram](image)

**A**

hsp70  cat  nos

**B**

hsp70  tctctgcgtgaaaaaatacca
hsp70cat  tctctgcgtgaaaaaatacca
cgaacatttcgcacagacgcagcagcagctgaggt
cgaacacttctcgacacagacgcagcagcagtgaaggt

tcgaaggatacgagacggcagGGGGATCCGTCGAGAlTr

tcgaagagaattcgcctgagaaaaaaaatcca

cgdC,aLLLaaLCagCwaaCCagCagCwaw&gaCLgLga*ggg

AAGAGATG
TCAGGAGCTAAGGAAGCTAAATG

**Fig. 2.** Map of the hsp70cat construction. (A) The hsp70 promoter fragment from pMON9508 (26) represented by the open box was placed in front of the CAT coding region (filled box) and the nopaline synthase poly A addition site (striped box) in the BamHI site of pUC8. The thin line represents plasmid sequences (not to scale). See “Materials and Methods” for details. (B) The nucleotide sequence of the 5′ leader of the endogenous hsp70 gene (top line) compared to the 5′ leader of the chimeric hsp70cat gene (bottom line). They are identical for the first 83 nucleotides, then the chimeric hsp70cat gene contains 36 different nucleotides instead of the 24 nucleotides present in the hsp70 gene. The sequence ends with the translation initiator ATG for the endogenous hsp70 protein and the CAT protein.

**DISCUSSION**

With the exception of pollen, all plant organs investigated so far (2, 7, 8, 17, 35) have been shown to undergo a heat shock response. When plant tissues are incubated at temperatures approximately 10 to 20°C higher than the normal growing temperature, there is transcriptional induction of mRNAs encoding a subset of proteins (reviewed in Ref. 17). Cultured tissues of maize (8), soybean, and tobacco (3) have previously been shown to alter the pattern of poly peptides synthesized in response to high temperature treatment. Tobacco protoplasts maintained at 25°C showed maximal induction of the heat shock proteins at 40°C and protein synthesis was greatly inhibited at 42°C (22). The response observed here of electroporated maize protoplasts to high temperatures is very similar to that previously observed for maize cells and tobacco protoplasts. Thus, electroporated protoplasts preincubated for 12 h are capable of a heat shock response.

The amount of transcript of the endogenous maize hsp70 gene (pMON9508) increased 40- to 60-fold in maize coleoptiles after 2 h of incubation at 42°C (26). This temperature-dependent transcript accumulation is also seen in transgenic petunia leaves for a chimeric maize hsp70 gene containing the promoter from pMON9508 (27). The observations presented here of the temperature-dependent increase in CAT activity in maize protoplasts as a result of expression from psp70cat are in agreement with these results. Because the mRNA produced from the hsp70 gene and the hsp70cat gene are not identical, a quantitative comparison may not be meaningful.

Introduction into tobacco of a chimeric construct containing a *Drosophila* hsp70 promoter linked to the neomycin phosphotransferase reporter gene resulted in transcriptional induction of the neo gene after heat shock, but the message was not translated at heat shock temperatures (30, 31). In this work, CAT mRNAs synthesized in maize protoplasts from a maize hsp70 promoter
at heat shock temperatures were translated at heat shock temperatures. Therefore, the 24 nucleotides of the 107 nucleotide 5′ leader of the endogenous hsp70 mRNA not present in hsp70cat are not essential for translation at heat shock temperatures. The synthesis of 25°C polypeptides at heat shock temperatures in maize in addition to hsps (7) contrasts with the repression of normal polypeptide synthesis observed in Drosophila incubated at heat shock temperatures (19). Therefore, while the transcriptional induction mechanisms for the heat shock genes are highly conserved across euukaryotic kingdoms, there are other aspects of the heat shock response which do not have a high degree of conservation. The ability to introduce DNA into maize protoplasts and the ability of these protoplasts to respond to incubation at heat shock temperatures provides a useful system for the study of heat stress in maize.

Acknowledgements—We would like to thank Dilip Shah for the gift of pMON9508. J. C. was supported in part by a fellowship from the McKnight Foundation training grant at Stanford University.

LITERATURE CITED

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