

Table S1: ROS-responsive SAA gene list. ROS-responsive gene sets obtained from Gadjev et al. (2006) and compared with SAA-specific genes from Rossel et al. (2007)

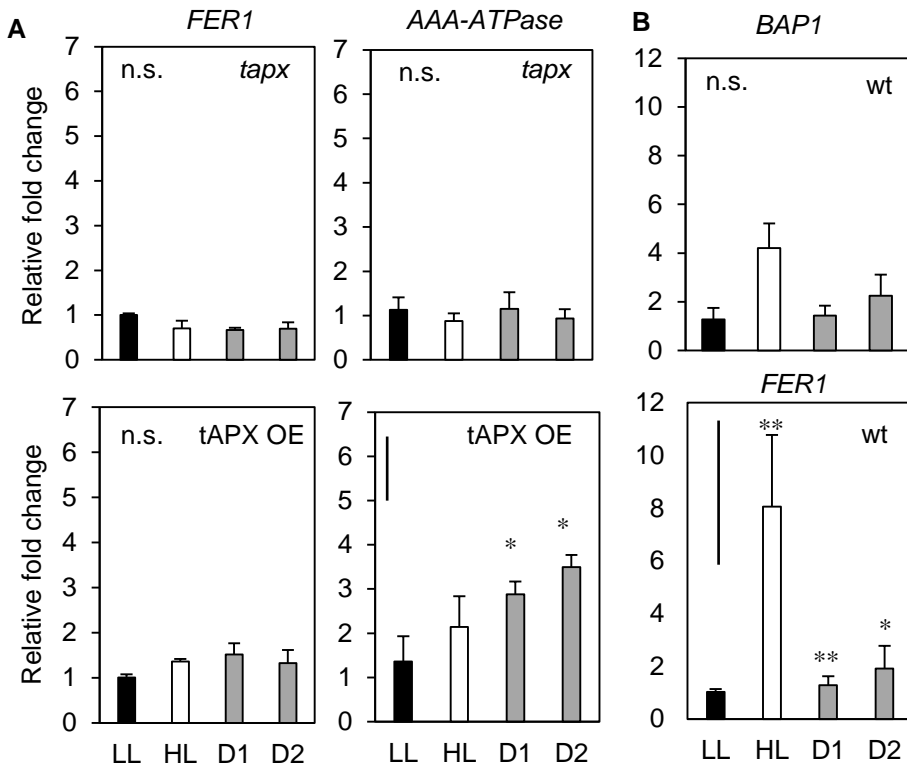
Accession number	Description
Shared O₂⁻ responsive genes	
AT2G17840	Upregulated by high light, drought, cold and salt stress.
Shared H₂O₂ responsive genes	
AT5G54300	Unknown
AT2G30500	Kinase interacting family protein
AT1G70140	Encodes a group I formin. Involved in the initiation and tip growth of root hairs through regulation of the actin cytoskeleton.
Shared ¹O₂ responsive genes of known function	
At1g16370	putative transport protein
At4g34410	AP2 domain transcription factor
At4g27654	Expressed protein
At5g47240	mutT domain protein-like
At1g61610	receptor-like kinase, putative
At4g08170	inositol 1,3,4-trisphosphate 5/6-kinase-like protein
At5g63450	cytochrome P450-like protein
At2g44840	ethylene response element binding protein
At5g22630	chorismate mutase/prephenate dehydratase-like protein
At1g49450	En/Spm-like transposon protein, putative
At4g10390	putative protein kinase CLV1 receptor kinase
At3g59350	protein kinase-like protein Pto kinase interactor 1
At5g67210	putative protein
At5g56980	putative protein
At1g18710	MYB transcription factor
At2g35710	putative glycogenin
At1g73080	receptor protein kinase, putative
At4g35480	RING-H2 finger protein RHA3b
At3g15990	putative sulfate transporter
At1g09940	putative glutamyl-tRNA reductase 2
At2g01150	RING-H2 finger protein RHA2b
At2g37900	putative peptide/amino acid transporter
At1g07500	hypothetical protein
At2g30360	putative protein kinase
At1g74430	AtMYB95

Table S1: ROS-specific SAA gene list (continued)

Accession number	Description
Shared ¹⁸O₂ responsive genes of known function (cont.)	
At2g39420	putative phospholipase
At5g24590	NAC2-like protein
At1g74930	AP2 domain transcription factor, putative
At5g65280	G protein-coupled receptor-like protein
At5g66210	calcium-dependent protein kinase
At1g74100	putative flavonol sulfotransferase
At2g20880	AP2 domain transcription factor
At3g10300	unknown protein
At3g44260	CCR4-associated factor 1-like protein
At5g26030	ferrochelataase-I
At2g47180	putative galactinol synthase
At3g46930	protein kinase 6-like protein
At3g53780	membrane protein
At1g74450	unknown protein
At5g60300	receptor like protein kinase
At2g20340	putative tyrosine decarboxylase
At5g60890	MYB transcription factor (ATR1)
At2g29450	glutathione S-transferase
At2g47060	putative protein kinase
At5g64310	arabinogalactan-protein
At1g63830	unknown protein
At1g67060	hypothetical protein
At4g27940	mitochondrial carrier protein
At5g17490	RGL3 transcription factor
At1g76380	hypothetical protein
At3g56260	putative protein
At3g56880	putative protein
At3g05640	putative protein phosphatase-2C
At1g75230	3-methyladenine DNA glycosylase, putative
At1g50640	ethylene responsive element binding factor 3 (AtERF3)
At5g64260	phi-1-like protein
At3g05580	putative serine threonine protein phosphatase
At5g09440	putative protein phi-1

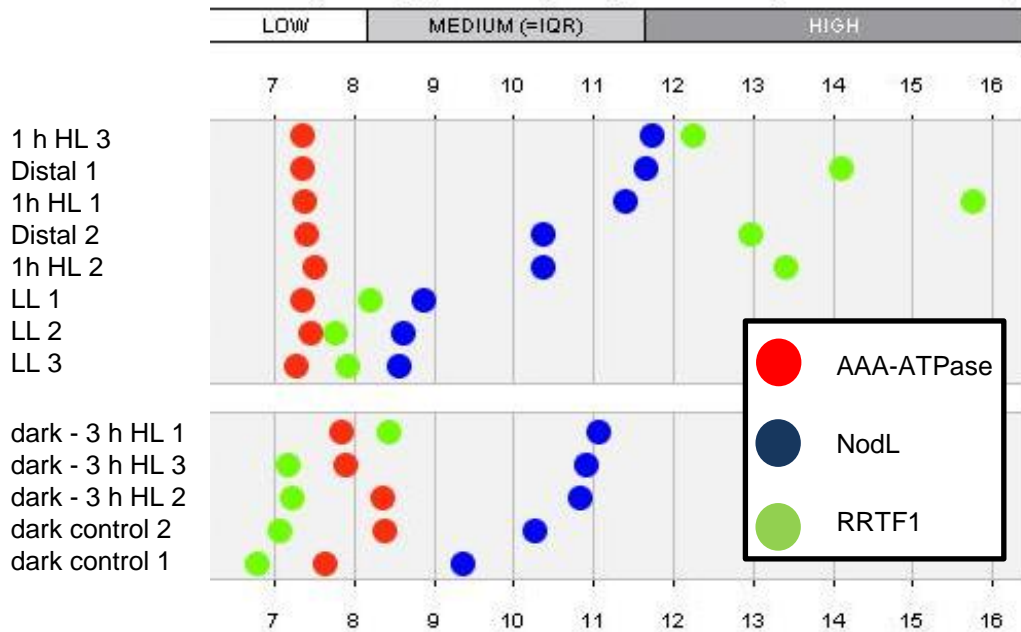
Table S1: ROS-specific SAA gene list (continued)

Accession number	Description
Shared ¹O₂ responsive genes of known function (cont.)	
At1g27770	envelope Ca ²⁺ -ATPase
At1g78080	AP2 domain transcription factor RAP2.4
At2g43020	putative amine oxidase
At5g55120	putative protein
At1g28380	unknown protein
At3g17860	unknown protein
At1g11960	unknown protein
At1g28050	CONSTANS family zinc finger protein
At1g01260	bHLH transcription factor
At3g26980	geranylgeranylated protein, putative
At1g70480	unknown protein
At1g08930	zinc finger protein ATZF1
At1g52720	hypothetical protein
Shared general stress response genes	
AT1G27730	SALT TOLERANCE ZINC FINGER, TF
AT1G80840	pathogen induced TF
AT4G17230	Encodes a scarecrow-like protein (SCL13).Member of GRAS gene family.
AT4G27410	Encodes a NAC transcription factor induced in response to desiccation.
AT4G17500	Encodes a member of the ERF (ethylene response factor) subfamily B-3 of ERF/AP2 transcription factor family
AT1G52890	encodes a NAC transcription factor whose expression is induced by drought, high salt, and abscisic acid.
AT5G47220	Encodes a member of the ERF (ethylene response factor) subfamily B-3 of ERF/AP2 transcription factor family.
AT5G59820	zinc finger protein involved in high light and cold acclimation, TF
AT1G43160	encodes a member of the ERF (ethylene response factor) subfamily B-4 of ERF/AP2 transcription factor family (RAP2.6).
AT5G04340	putative C2H2 zinc finger TF
AT2G38470	TF, involved in response to various abiotic stresses - especially salt stress.
AT2G40140	Salt-inducible zinc-finger protein
AT1G77450	NAC domain containing protein 32
AT1G19020	unknown, response to oxidative stress



C

Level of expression (signal intensity on Affymetrix Arabidopsis ATH1 Genome Array)



created with GENEVESTIGATOR

Figure S1: Commonly used ROS marker genes, *FER1* and *AAA-ATPase* are not induced by LED HL in stable silenced and overexpressing tAPX lines. (A) Transcriptional analysis of *FER1* (H₂O₂ marker), and *AAA-ATPase* (commonly used to demonstrate RB but not MV induction) in *tapx 2/1* and the tAPX -14/2 overexpression lines. (B) *FER1* and *BAP1* (¹O₂ marker) are shown for wt tissue, as well as (C) Geninvestigator data demonstrating the differences between *AAA-ATPase*, *NodL* (¹O₂ responsive SAA marker), and *RRTF1* (SAA marker) from the Rossel et al., (2007) microarray (AT-00345) and a Shao et al., (2013) microarray (AT-00691) of seedlings grown on plates under different growth conditions. The latter tissue was from seedlings grown under continuous (100 μmol photons m⁻² s⁻¹) light on plates, transferred at 12 days to the dark for 3 days, and treated with 3 h HL (1000 μmol photons m⁻² s⁻¹). LL, low light; HL, high light; D1, lower distal leaf position 5; D2, upper distal leaf position 7. One-way ANOVAs were performed on each *ZAT10* data set, *post hoc* Fisher's LSD results are displayed as a bar in the top left corner of each plot. t-test were performed on *ELIP2* data sets. *p < 0.05, ** p < 0.005 n.s., not significant. Error bars represent standard error, N = 3.

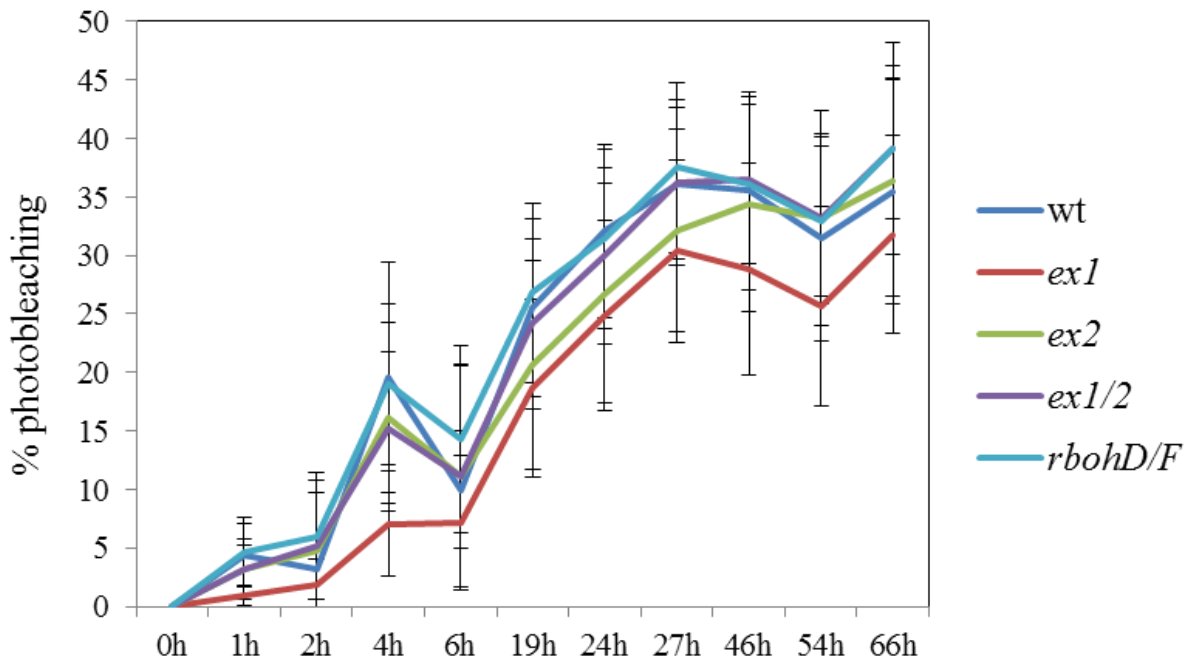


Figure S2: Comparison of photobleaching development between ROS signaling mutants. (A) Bleaching percentages were obtained through the Scanalyser and were normalised to 0% at time 0. Dark blue, *wt*; red, *ex1*; green, *ex2*; purple, *ex1/ex2*; light blue, *rbohD/F*. Error bars represent standard deviation, n = 15. (B) representative images of photobleaching on a plant undergoing continuous HL and detection by the Scanalyzer software.

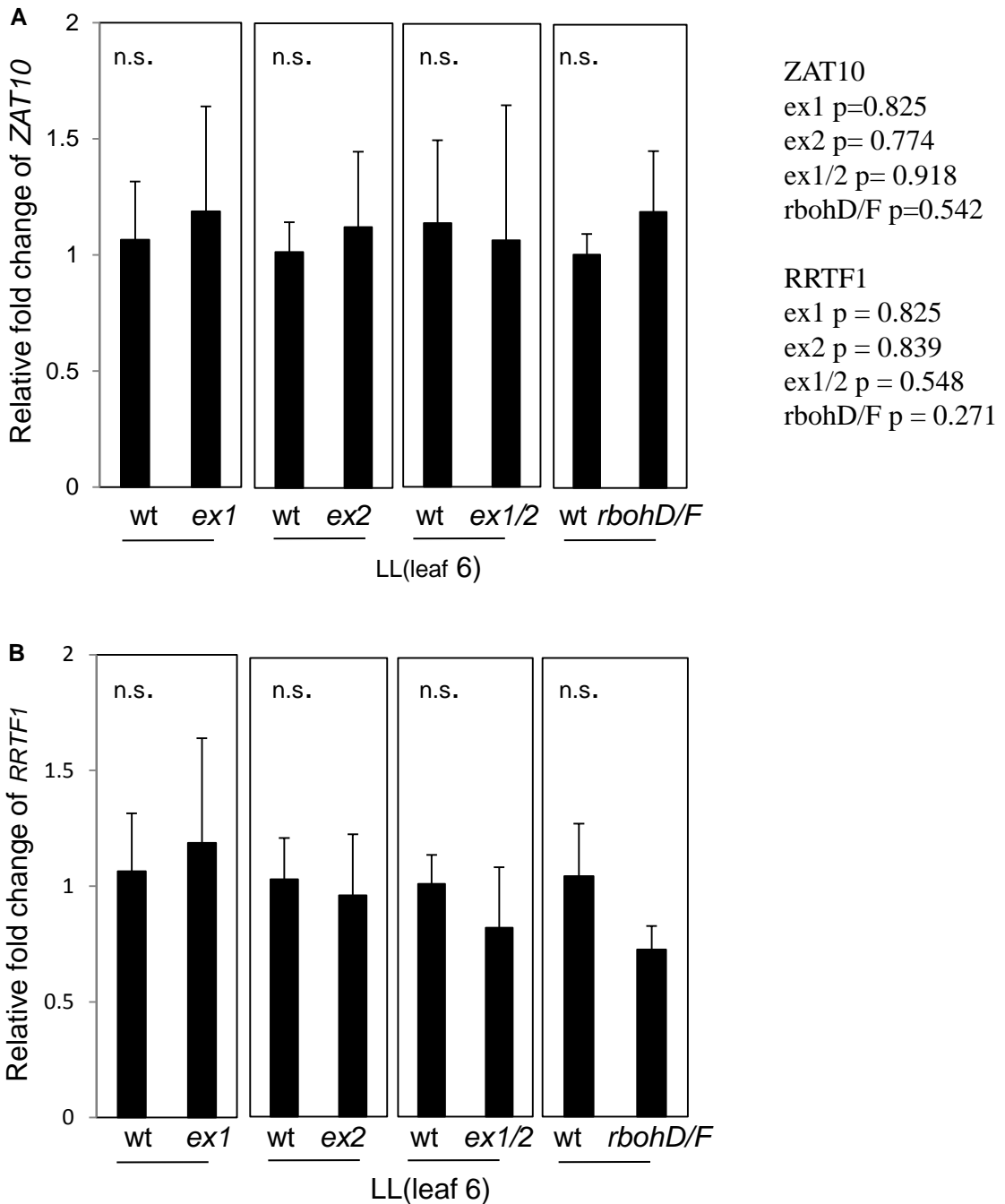


Figure S3: Basal *ZAT10* and *RRTF1* induction in LL tissue of wt and ROS signaling mutants. (A) *ZAT10* induction comparing wt-Col with *ex1*, *ex2*, *ex1/2*, and *rbohD/F* LL tissue, (B) *RRTF1* for the same comparisons. One-way ANOVAs were performed on each data set, *post hoc* Fisher's LSD statistics are displayed as n.s. (not significant) in the top left corner of each plot. Error bars represent standard error; N = 3.

Table S2 A: Statistical tests for differences within ZAT10 ROS-signaling mutant data sets. Statistical results of one-way ANOVAs and post hoc Fischer LSD and Holm-Sidak tests within mutant datasets. N = 3.

Data set	one-way ANOVA	Fischer LSD	Holm-Sidak
ex1	p = 0.0021	0.924	HL vs LL (0.004), D1(0.019), D2 (0.031)
ex1/2	p = 0.043	0.638	HL vs D2 (0.006)
ex2	p = 0.005	1.071	D1 vs LL (0.002), D2 (0.015)
rbohD/F	p = 0.004	2.104	HL vs LL (0.001), D2(0.003), D1 (0.017)

Table S2 B: Statistical tests for differences within RRTF1 ROS-signaling mutant data sets. Statistical results of one-way ANOVAs and post hoc Fischer LSD and Holm-Sidak tests within mutant data sets. N = 3.

Data set	one-way ANOVA	Fischer LSD	Holm-Sidak
ex1	p = 0.076	n.s.	-
ex1/2	p = 0.397	n.s.	-
ex2	p = 0.001		D1 vs LL, HL vs LL
rbohD/F	p < 0.001	9.659	HL vs LL (<0.001), D2(<0.001), D1 (<0.001)

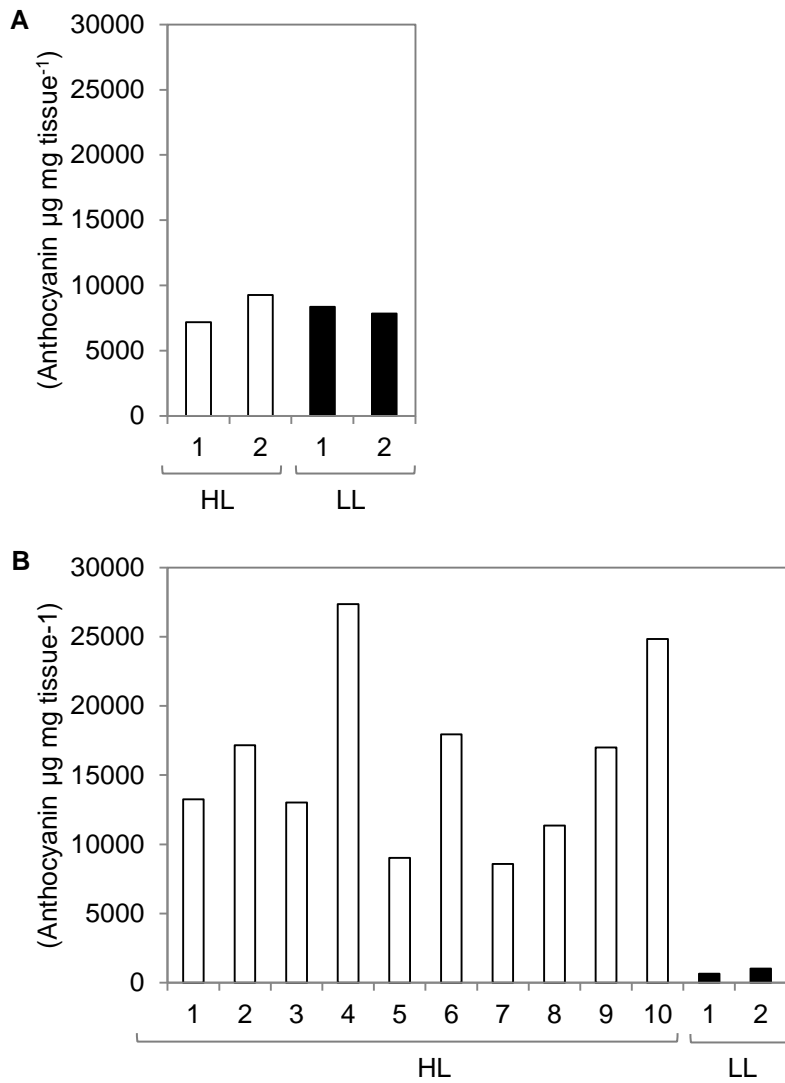


Figure S4: Anthocyanin Test Assays. (A) anthocyanin accumulation in tissues exposed to 1 h $1000 \mu\text{mol photons m}^{-2} \text{s}^{-1}$ HL; (B) anthocyanin accumulation after 3 days of HL-treatment. White bars represent HL-treated tissue; black bars represent LL controls.

Time (h)	Plant	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
2.5	2	2	0	T	0	0	3	0	0	0	0	0	0	0			
20	2	2	1	T	2	1	3	0	1	1	1	1	1	1	1	1	
2.5	4	0	0	T	0	0	0	0	0	0	0	0	0	0			
20	4	0	0	T	3	2	2	0	1	0	1	1	1	1	0		
2.5	5	0	0	T	0	0	0	0	0	0	0	0	0	0	0	0	
20	5	0	0	T	0	0	0	0	0	0	0	0	0	0	0	0	
2.5	3	0	0	1	T	1	0	3	1	0	1	0	1	0			
20	3	0	0	1	T	1	0	2	0	0	1	1	1	1	1		
2.5	1	2	1	3	1	0	T	0	2	3	0	1	0	0	2	?	?
20	1	2	1	3	2	0	T	0	2	3	0	2	1	1	1	1	1
2.5	7	0	2	1	0	1	T	1	2	3	2	3	1	0	2	1	0
2.5	8	1	0	3	0	1	T	2	3	3	0	3	1	0	2	1	1
2.5	11	1	0	3	0	3	T	1	2	3	0	3	1	1	1	1	0
2.5	9	0	0	1	1	0	3	2	2	T	0	2	2	1	3?	1?	1?
2.5	10	0	0	1	0	0	3	3	0	T	0	2	3	0	0	0	0

Figure S5: Rhodamine B infiltrations. 2.5 and 20 h after infiltration of Rhodamine B fed through petioles of leaf insertion 6. In general, the dye infiltrates tissues in an orthostichous pattern.

Table S3: Statistical analysis results for connected and unconnected leaf data sets.
 Statistical results of one-way ANOVAs and post hoc Fischer LSD and Holm-Sidak tests within experimental data sets.

Data set	one-way ANOVA	Fischer LSD	Holm-Sidak
ZAT10 vasc. 5 min	p < 0.001	1.795	HL vs all tissue (< 0.001)
ZAT10 vasc. 1 h	p < 0.001	2.128	HL vs all tissue (<0.001), D1 vs O1 (0.033), O2 (0.022)
RRTF1 vasc. 5 min	p < 0.001	2.155	HL vs LL (<0.001), D1(0.004), D2 (<0.001), O2 (<0.001) O1 vs O2 (<0.001), D2(0.001), LL (0.002),D1 (0.011)
RRTF1 vasc. 30 min	n.s. 0.461	- - -	
RRTF1 vasc. 1 h	p = 0.006	14.433	HL vs LL (p< 0.001), D1(0.004), O2 (0.003) O1 vs LL (p< 0.011) D2 vs LL (0.032)
NodL vasc. 5 min	n.s. p = 0.718	- - -	
NodL vasc. 30 min	p < 0.001	6.325	HL vs all tissue (p < 0.001)
NodL vasc. 1 h	p < 0.001	10.805	HL vs all tissue (p < 0.001) O2 vs LL (0.005), D1(0.013), D2 (0.012), O1 vs LL (0.013), D2(0.037), D1 (0.041)

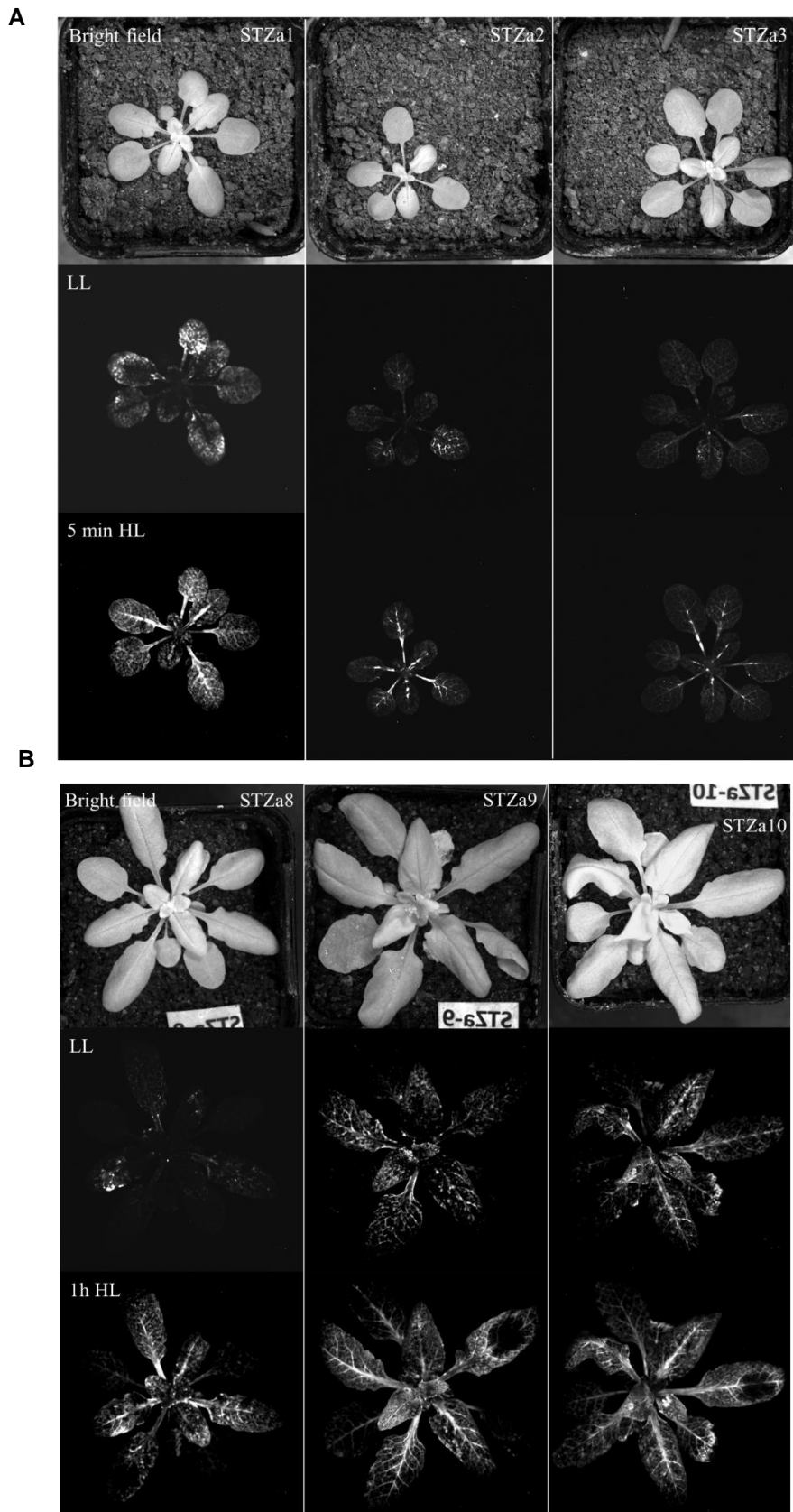


Figure S6: 5 min and 1h timepoints from the HL spot treatment of ZAT10::LUC HL-specific transgenics. CCD camera images of three biologicals, including the bright field image, under LL, and after either (A) 5 min or (B) 1h HL spot treatment of leaf position 6.

Table S4: Shared β -cyclocitral and SAA genes. Shared genes from the β -cyclocitral (Ramel et al., 2012) and SAA (Rossel et al., 2007) microarrays.

Accession number	Gene name	Compartment
AT1G17420	LOX3	chloroplast
AT4G36010		anchored to membrane, extracellular region
AT3G25780	AOC3	chloroplast, plasma membrane, vacuolar membrane
AT5G05600		cytoplasm
AT1G80840	WRKY40	nucleus
AT4G17230	SCL13	nucleus
AT4G27410	RD26, ANAC072	nucleus
AT2G06050	OPR3	peroxisome
AT5G10695		extracellular region, mitochondrion
AT4G17500	ERF1	nuclear pore, nucleus
AT3G51450		chloroplast, endomembrane system
AT5G47220	ERF2	nucleus
AT1G32920		endomembrane system
AT1G28480	GRX480, ROXY19	cytoplasm
AT5G27520	PNC2	mitochondrial inner membrane, mitochondrion, peroxisome
AT3G55980	SZF1	nucleus
AT3G29575	AFP3	nucleus
AT5G09440	EXL4	chloroplast, membrane, plasmodesma
AT3G15210	ERF4	nuclear body, nucleus
AT1G19300	PARVUS/GLZ1	endoplasmic reticulum
AT2G42540	COR15A	chloroplast, chloroplast envelope, chloroplast stroma
AT5G59820	ZAT12	intracellular, nucleus
AT2G29440	GSTU6/GST24	cytoplasm, cytosol
AT1G66090		chloroplast
AT3G56880		nucleus
AT1G73480		cytoplasm
AT4G18010	5PTASE2/IP5PII	cytoplasm
AT1G70700	JAZ9	nucleus
AT4G20860		cytoplasm
AT4G15210	ATBETA-AMY, BAM5, BMY1	chloroplast, cytoplasm
AT2G23810	TET8	golgi apparatus, plasma membrane, plasmodesma
AT1G29330	ERD2	golgi apparatus, ER
AT1G72450	JAZ6	nucleus
AT5G04340	ZAT6	nucleus
AT2G36590	PROT3	plasma membrane
AT1G05575		endoplasmic reticulum
AT1G09070	(AT)SRC2	ER, plasma membrane, vacuole
AT2G26530	AR781	chloroplast, plasma membrane
AT2G38470	WRKY33	nucleus
AT1G01720	ATAF1	nucleus

Table S4: Shared β -cyclocitral and SAA genes (continued)

Accession number	Gene name	Compartment
AT2G41100	TCH3	mitochondrion, plasmodesma, vacuolar membrane
AT5G13190	GILP	plasma membrane
AT5G06870	PGIP2	cytosol
AT1G51090		nucleus
AT5G62570		
AT4G12720	NUDT7	cytoplasm, plasma membrane
AT5G14700		
AT4G34710	ADC2	nucleus
AT5G42050		nucleus
AT4G34150		cytosol, plasmodesma
AT1G14130		cytoplasm, nucleus
AT1G71100	RSW10	cytoplasm
AT2G46600		nucleus
AT5G35735		plasma membrane
AT2G40140	CZF1, SZF2	nucleus
AT5G26030	FC1	chloroplast, mitochondrion
AT2G33380	RD20, CLO3	chloroplast membrane, vacuole
AT2G23320	WRK15	nucleus
AT1G01140	CIPK9, PKS6	cytoplasm, plasma membrane
AT1G42990	BZIP60	ER, nucleus
AT1G73500	MKK9	mitochondrion
AT5G25930		plasma membrane
AT1G77450	ANAC032	nucleus
AT3G52800		nucleus
AT1G09970	RLK7	micropyle, plasma membrane
AT1G30620	HSR8, MUR4, UXE1	golgi apparatus
AT4G16760	ACX1	peroxisome, plasmodesma
AT2G01450	MPK17	nucleus
AT4G36990	HSF4	nucleus
AT3G23920	BAM1, BMY7	chloroplast, cytosol, nucleus
AT4G24160		unknown
AT2G29450	GSTU5	plasma membrane, plasmodesma
AT5G10380	RING1	plasma membrane
AT5G47060		nucleus

Table S5: qRT-PCR primers and used for Roche (with UPL probes) and SybrGreen for quantitative transcript analysis.

Target	Accession	Probe	Primer sequence (5' -3')
CYP5	At2g29960	UPL 103	GGCAGTTCCTAAAACCTGCAGAA
			TTCCCTTGTAGTGTAGAGGTTTCC
PP2A	At1g13320	UPL 120	GACCGGAGCCAACCTAGGAC
			AAATGCGACAAAACCTGGTAACT
ZAT10	At1g27730	UPL 31	GGACAAAGGGTAAGCGATCTAA
			AGAAGCATGAGGCAAAAAGC
RRTF1	At4g34410	UPL 68	TCGGGTATGCATTATCCTAACA
			AAGCTCTTGCTCCGGTGA
NodL	At5g64870	UPL144	TCGTGTTCTTGTTGCTTCCA
			TGAACCTTGTCAAACACTTCCTT
ELIP2	At4g14690	UPL 101	CCACCACAAATGCCACAG
			GCAAATCTCCAAACTTCGTACTC
FER1	At5g01600	UPL 21	CTGAAAAAGGAGATGCTTTATATGC
			TGAGGCCACTTTGTGAACG
BAP1	At3g61190	UPL 152	TAAACCGGAGACCCATCAAG
			TCGACATTTCTCGTCGATTTT